

The Proteome Profile of the Human Osteosarcoma U2OS Cell Line

KATERINA N. NIFOROU¹, ATHANASIOS K. ANAGNOSTOPOULOS², KONSTANTINOS VOUGAS², CHRISTOS KITTAS¹, VASSILIS G. GORGULIS¹ and GEORGE T. TSANGARIS²

¹Department of Histology-Embryology, School of Medicine, University of Athens;

²Proteomics Research Unit, Centre of Basic Research II,
Biomedical Research Foundation of the Academy of Athens, Athens, Greece

Abstract. The human osteosarcoma U2OS cell line is one of the first generated cell lines and is used in various areas of biomedical research. Knowledge of its protein expression is limited and no comprehensive study on the proteome of this cell line has been reported to date. Proteomics technology was used in order to analyse the proteins of the U2OS cell line. Total protein extracts were separated by two-dimensional gel electrophoresis (2-DE) and analysed by matrix-assisted laser desorption ionisation-mass spectrometry (MALDI-MS) and MALDI-MS-MS following in-gel digestion with trypsin and, finally, protein identification was carried out by peptide mass fingerprint (PMF) and post source decay (PSD), respectively. Approximately 3,000 spots were excised from two 2-DE gels and were analysed, resulting in the identification of 237 different gene products. The majority of the identified proteins were enzymes, regulatory proteins and RNA-associated proteins, while leukocyte markers and oncogenes were also present. Our findings include 11 protooncogenes (FKBP4, SRC8, PSD10, FUBP1, PARK7, NPM, PDIA1, OXRP, SET, TCTP and GRP75) related to the cancerous state of the U2OS cell line. The U2OS 2-DE database provides the basis for future protein studies.

Human osteosarcoma is a primary malignant tumor of the bone affecting children and young adults (age 15-29 years), as well as adults in later life (age >60 years). It is the 6th most common type of cancer in children and appears during

the second decade of life in the adolescent growth period, with a high incidence in teenage boys who are experiencing growth spurts. The cause of osteosarcoma is unknown; however irradiation, genetic influences and rapid bone growth are considered to be related to its development.

In order to further understand the molecular basis of this disease, we applied proteomics technology to the human osteosarcoma U2OS cell line. Proteomics technologies are powerful analytical tools and, in combination with other high throughput screening techniques, are widely used for the analysis of biological specimens. Protein analysis by two-dimensional gel electrophoresis (2-DE) coupled to mass spectrometry nowadays comprises the standard approach for the construction of comprehensive protein databases (1). Protein identification is greatly enhanced and improved by robotic systems and automated mass spectra acquisition devices, which allow thorough analysis of biological specimens such as cell lines.

Proteome databases are primarily used to illuminate normal and disease states, and the information acquired can be used in diagnosis and treatment, and in biomarker discovery (2). Cancer cell lines are generally used for proteomic analysis since they exhibit equivalent changes to the tissue they derive from. Hence the screening of the proteome of some cancer cell lines has been performed, e.g. human immature T-cell line CCRF-CEM, HeLa cell line and osteosarcoma Saos2 cell line (3-5).

The human osteosarcoma U2OS cell line was derived in 1964 from a moderately differentiated sarcoma of the tibia of a 15-year-old girl. It is one of the first generated cell lines and is used very frequently. Spectral karyotyping analysis and cytogenetic analysis has revealed chromosomal instability, structural rearrangements and alterations and high incidence of aneuploidy (6). Spectral analysis indicated the near-triploid state of U2OS cells which appeared to be a combination of tetraploidization and chromosomal losses. Other rearrangements mainly involved chromosomes 20 and 8, from simple translocations to highly complex rearrangements, but

Abbreviations: PMF, Peptide mass fingerprint; PSD, post source decay; DMEM, Dulbecco's modified Eagle's medium; FCS, fetal calf serum.

Correspondence to: George Th. Tsangaris, Proteomics Research Unit, Centre of Basic Research II, Biomedical Research Foundation of the Academy of Athens, Soranou Ephessius 4, 11527 Athens, Greece. Tel: +30 210 6597075, Fax: +30 210 6597545, e-mail: gthsangaris@bioacademy.gr

Key Words: Osteosarcoma, U2OS, proteomics, 2-DE protein database, mass spectrometry, MALDI-MS.

also to a less extent chromosome 17. Most of the centromeric breakpoints occurred in chromosomes 6, 13 and 17, but at least four in chromosomes 2, 4, 5, 8, 19 and 21.

Two tumor suppressive genes, *p53* and *pRb*, are functional in U2OS cells, whereas in other, more aggressive, osteosarcoma cell lines such as Saos2 these genes are mutated (7-9). Analysis of other oncogenes and tumor suppressor genes signalized 5 times amplification of *c-myc* with no apparent serum stimulations; additionally *c-fos* had no visible alterations. Compared with other osteosarcoma cell lines, U2OS cells have the lowest level of chromosomal numerical variations and only 2% of the cells have multipolar mitoses, similar to normal control fibroblasts, probably due to functional *p53* and *pRb* (9).

The U2OS cell line is widely used in biomedical research such as biochemistry, molecular biology, bone formation, arthritis (10-14). In this study, the proteome profile of U2OS cells was constructed, consisting of 237 single gene products.

Materials and Methods

Materials and reagents. Immobilized pH-gradient (IPG) strips and IPG buffers were purchased from Bioworld Laboratories (Hercules, CA, USA). Acrylamide/piperazine-di-acrylamide (PDA) solution (37.5:1, w/v) was purchased from Biosolve Ltd. (Valkenswaard, the Netherlands) and the other reagents for the polyacrylamide gel preparation from BioRad. CHAPS was obtained from Roche Diagnostics (Mannheim, Germany), urea from AppliChem (Darmstadt, Germany), thiourea from Fluka (Buchs, Switzerland), 1,4-dithioerythritol (DTE) and EDTA from Merck (Darmstadt, Germany). Except for CHAPS, which was kept at 23°C, the other reagents were kept at 4°C.

Cell cultures. Cells were grown in Dulbecco's modified Eagle's medium (DMEM; Biochrome, Berlin, Germany) supplemented with 10% fetal calf serum (FCS) (Biochrome), at 37°C in a humidified atmosphere supplemented with 5% CO₂.

Two-dimensional gel electrophoresis. U2OS cells (40x10⁶) were washed with normal saline and resuspended in 0.5 ml of urea buffer, consisting of 20 mM Tris, 7 M urea, 2M thiourea, 4% CHAPS, 10 mM 1,4-dithioerythritol, 1 mM EDTA and a mixture of protease inhibitors [1 mM PhenylMethaneSulphonylFluoride (PMSF) and 1 tablet Complete™ (Roche Diagnostics, Basel, Swiss) per 50 ml of suspension buffer] and phosphate inhibitors (0.2 mM Na₂VO₃ and 1 mM NaF). The cells were lysed by sonication at 3x60 s in 35% amplification and the suspension was centrifuged at 13000xg for 30 min at 4°C. The protein content of the supernatant was determined using the EXPERION Automated Electrophoresis Station (BioRad) in combination with Protein 260 Analysis Kit™ (BioRad) according to the manufacturer's instructions.

2D-Gel electrophoresis was performed as reported elsewhere (4). Total protein (1 mg) was applied to immobilized pH 3-10 non-linear gradient strips in sample cups at their acidic and basic ends. Focusing started at 250 V for 30 min and the voltage was gradually increased to 8000 V at 3 V/min and remained constant for a further 22 h. The second-dimensional separation was performed in 12%

SDS-polyacrylamide gels (180x200x1.5 mm), running at 40 mA per gel in a PROTEAN apparatus (Biorad). After fixation with 50% methanol, containing 10% acetic acid for at least 2 h, the gels were stained overnight with colloidal Coomassie blue (Novex, San Diego, CA, USA), washed twice with water and scanned in a densitometer (GS-800 Calibrated Densitometer; Biorad).

Peptide mass fingerprint (PMF) and post source decay (PSD). Peptide analysis and protein identification were performed as described elsewhere (15). Spots were automatically detected by Melanie 4.02 (GeneBio, Geneve Bioinformatics S.A., Geneva, Swiss) software on the Coomassie blue-stained gel, excised by the Proteiner SPII (Bruker Daltonics, Bremen, Germany), destained with 30% acetonitrile in 50 mM ammonium bicarbonate and dried in a speed vacuum concentrator (MaxiDry Plus; Heto, Allered, Denmark). Each dried gel piece was rehydrated with 5 µl of 1 mM ammonium bicarbonate containing 50 ng trypsin (Roche Diagnostics) and left in the dark overnight at room temperature. Twenty µl of 50% acetonitrile, containing 0.3% trifluoroacetic acid were added to each gel piece and incubated for 15 min with constant shaking. The resulting peptide mixture (1.5 µl) was simultaneously applied with 1 µl of matrix solution, consisting of 0.025% α-cyano-4-hydroxycinnamic acid (Sigma-Aldrich, USA), standard peptides *des*-Arg-bradykinin, (904.4681 Da; Sigma-Aldrich, USA) and adrenocorticotropin hormone fragment 18-39, (2465.1989 Da; Sigma-Aldrich, USA) in 65% ethanol, 35% acetonitrile and 0.03% trifluoroacetic acid. Samples were analyzed for PMF with matrix-assisted laser desorption-mass spectrometry (MALDI-MS) in a time-of-flight mass spectrometer (Ultraflex II, Bruker Daltonics, Bremen, Germany). Matching peptide and protein searches were performed automatically, as described by Berndt *et al.* (15). Each spectrum was interpreted by the Mascot Software (Matrix Sciences Ltd., London, UK). For peptide identification, the monoisotopic masses were used and a mass tolerance of 0.0025% (25 ppm) was allowed. Unmatched peptides, or peptides with up to one miscleavage site were not considered. The peptide masses were compared with the theoretical peptide masses of all available proteins from all species using SWISS-PROT (<ftp://ftp.expasy.org/databases/uniprot/knowledgebase>), IPI (<ftp://ftp.ebi.ac.uk/pub/databases/IPI/current/>), and MSDB (<ftp://ftp.ebi.ac.uk/pub/databases/MassSpecDB>) databases which are updated bimonthly. The probability score identified by the software was used as the criterion of the identification. Samples not identified by PMF (probability significance of *p*<0.05) were automatically selected for post-source decay (PSD) MS-MS analysis or MALDI-MS-MS. The peptide masses chosen for PSD-MS-MS analysis had a signal intensity of >600 counts and were excluded from the trypsin autodigest, matrix and keratin peaks. The resulting PSD spectra were also interpreted by the Mascot Software and Mascot probability-based scores of *p*<0.02 were considered significant. The identified proteins were annotated on the gel image by hand.

Results

2-DE database. The protein extract from the osteosarcoma U2OS cell line was separated by 2-DE electrophoresis on IPG strips and the spots were visualized with colloidal Coomassie blue staining. In Figure 1, a representative example is shown of the proteins present in U2OS cells as separated on a pH 3-10 NL gel. Four pH 3-10 NL IPG gels were analyzed and

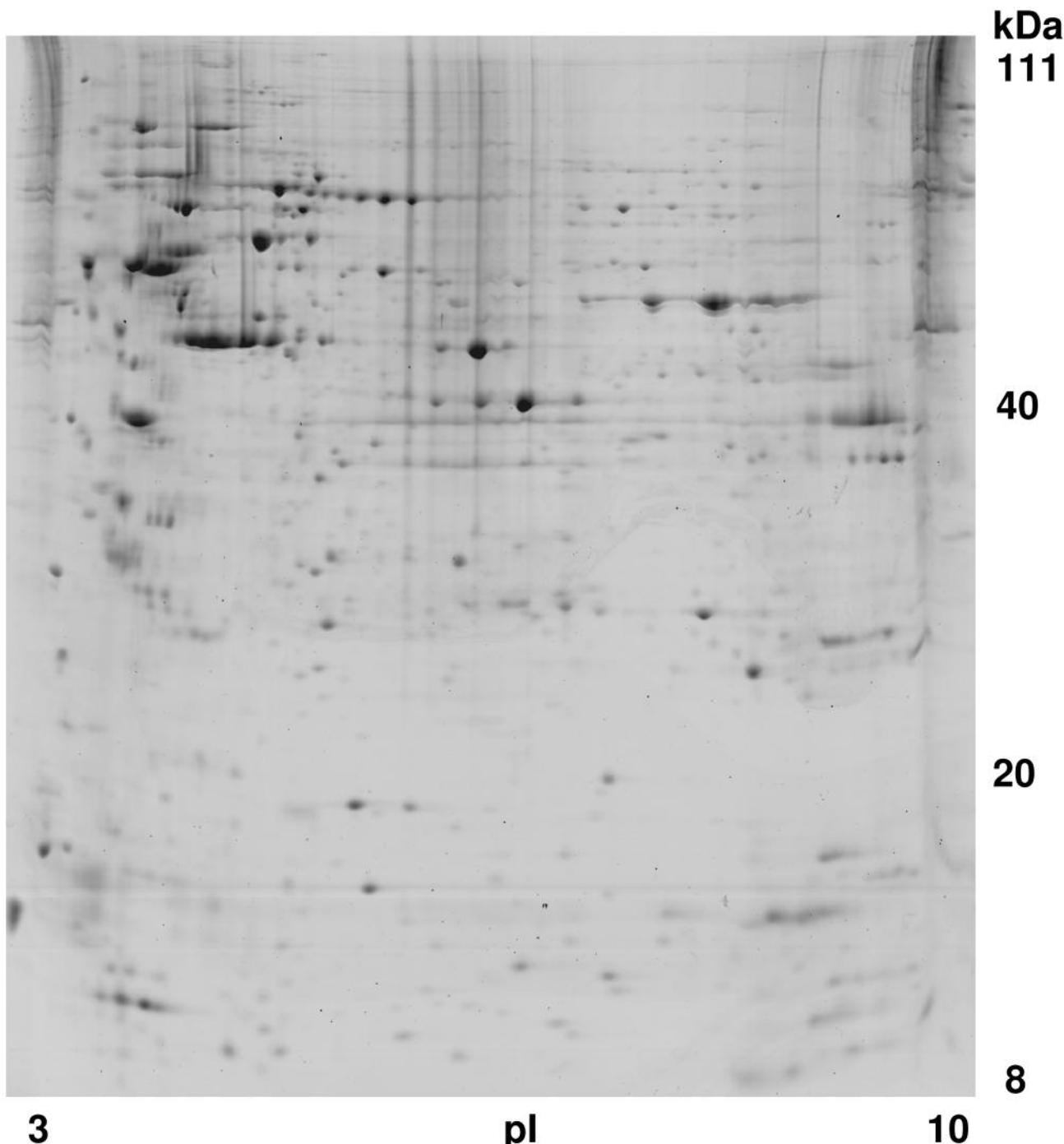


Figure 1. Two-dimensional gel analysis of total protein extract from U2OS cells. Proteins were extracted and separated on IPG strip pH 3-10 non-linearly, followed by a 12% SDS-polyacrylamide gel. The gel was stained with Coomassie blue.

approximately 2,500 spots were detected using the Melanie 4.02 software. Those spots were excised from the pH 3-10 gel and analyzed for protein identification following in-gel digestion with trypsin. Each spot was analyzed for PMF with MALDI-MS in a time-of-flight mass spectrometer and

proteins were identified automatically by the peptide mass matching. Proteins not identified by PMF were subsequently selected for PSD-MS-MS and analyzed with MALDI-MS-MS. Using an internal peptide standard to correct the measured peptide masses, we were able to use very narrow windows of

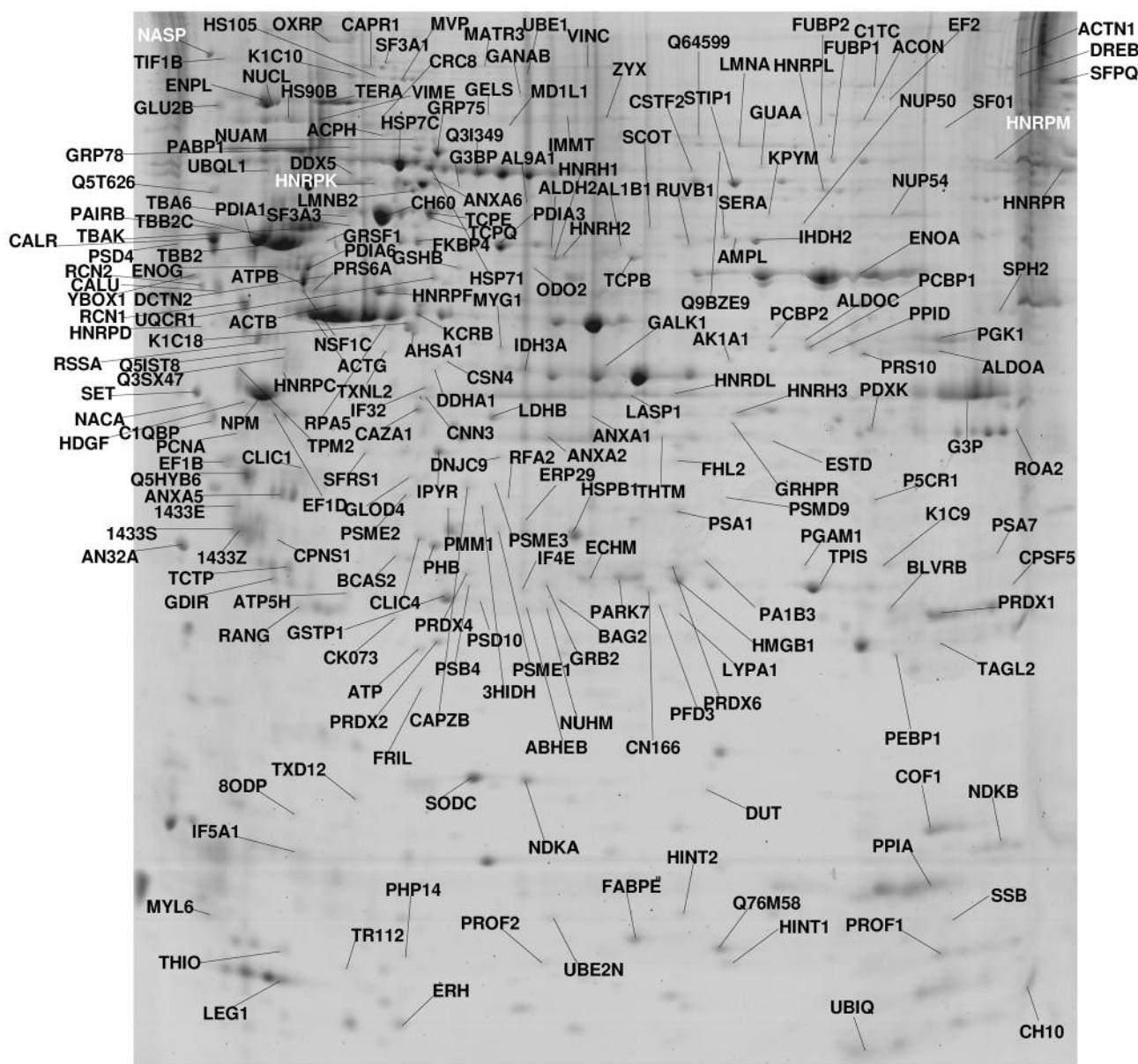


Figure 2. Identification of protein spots present in the gel of Figure 1, analyzed by PMF and/or PSD-MS-MS. The identified proteins are annotated by their abbreviated names and are listed in Table I.

mass tolerance (0.0025%) and hence, increase the confidence of identification, as well as the total identification rate up to 85%. This resulted in the identification of 237 different gene products which are annotated on the gel shown in Figure 2.

The SWISS-PROT accession numbers, the abbreviated and full names of the proteins, the theoretical MW as well as data from the mass spectrometry analysis, *i.e.* the numbers of matching peptides and the probability that the identification is random, are listed in Table I. Furthermore, information on the subcellular localization and function of these proteins as given in publicly accessible databases are shown.

From the 2,500 spots analyzed 673 spots were identified resulting in 237 different gene products. Thus the most abundant proteins present in U2OS cells were 62 members of the heat shock protein family, 34 members of the actin family, 19 members of the lamin family, 18 members of the tubulin family and 16 members of the annexin family of proteins.

Subcellular localization. For 5% of the identified proteins, no data were found regarding their subcellular localization. Most of the proteins are cytoplasmic (37%), nuclear (27%), mitochondrial (12%) and endoplasmic reticular (8%).

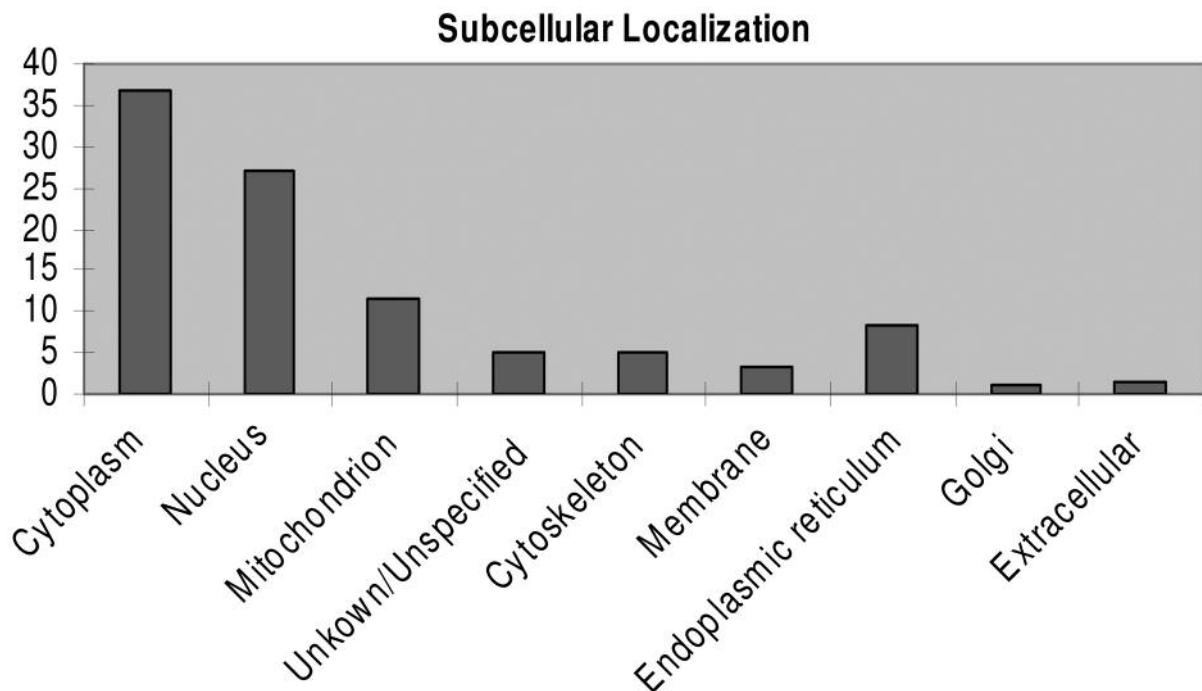


Figure 3. Subcellular localization of the human osteosarcoma U2OS cell line proteins. The proteins in Table I were categorized according to their localization in the cells.

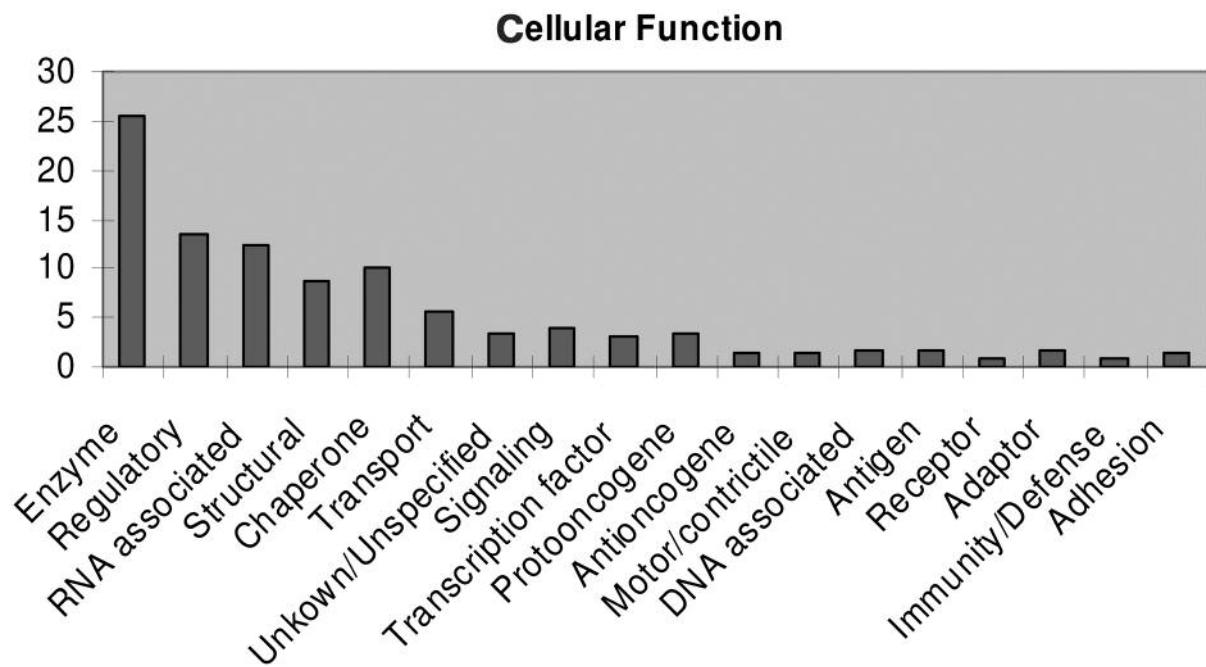


Figure 4. Classification of the proteins present in U2OS cells into functional groups. Protein molecules and their function are given in Table I.

Cytoskeletal proteins constitute 5% of the proteome and 3% are membranic proteins. The subcellular localization is also presented in Figure 3.

Function. As expected, most of the proteins are enzymes (25%), regulatory (13%) and RNA associated proteins (12%) as seen in Figure 4. There are chaperone/stress

Table I. Proteins from the human osteosarcoma U2OS cell line were extracted and separated by 2-D gel electrophoresis, as described in Materials and Methods. The proteins were identified by PMF and/or PSD, following in-gel digestion with trypsin. The spots representing the identified proteins are indicated in Figures 2 and 3 and are designated with their abbreviated names, or the SWISS-PROT accession numbers, or the accession numbers of the other databases. The theoretical Mr, the matching peptides and the probability of a random identification (Score), as well the annotated subcellular location and function are listed. Score is -10*log(P), where P is the probability that the observed match is a random event (MASCOT, <http://www.matrixscience.com>). Score >53 indicate $p < 0.05$.

Accession no	Protein name	Protein symbol	Protein description	MW score	Mascot score	Peptides matched	Cellular function	Subcellular localization
O00299	CLIC1_HUMAN	CLIC1	Chloride intracellular channel protein 1, NCC27	26792	141	11	Transport Channel	Membranic, Nuclear membrane
P30086	PEBP1_HUMAN	PEBP1	Phosphatidylethanolamine-binding protein 1, Raf kinase inhibitor protein	21057	94	9	Antioncogene	Cytoplasmic
P08238	HS90B_HUMAN	HS90B	Heat-shock protein HSP 90-beta	83264	223	30	Chaperone/Stress	Cytoplasmic
P48643	TCP1_HUMAN	TCP1	T-complex protein 1 subunit epsilon	59671	180	34	Chaperone/Stress	Cytoplasmic
P50990	TCPQ_HUMAN	TCPQ	T-complex protein 1 subunit theta, TCP-1-theta	59621	163	27	Chaperone/Stress	Cytoplasmic
P78371	TCPB_HUMAN	TCPB	T-complex protein 1 subunit beta, CCT-beta	57488	284	20	Chaperone/Stress	Cytoplasmic
Q12790	FKBP4_HUMAN	FKBP4	FK506-binding protein 4	51805	282	33	Chaperone/Stress, Enzyme, Protooncogene, Regulatory	Cytoplasmic
P61289	PSME3_HUMAN	PSME3	Proteasome activator complex subunit 3, Ki nuclear autoantigen, PA28g	29506	81	10	Chaperone/stress, Regulatory	Regulatory
Q06323	PSME1_HUMAN	PSME1	Proteasome activator complex subunit 1, PA28alpha	28723	86	9	Chaperone/stress, Regulatory	Cytoplasmic
Q9UL46	PSME2_HUMAN	PSME2	Proteasome activator complex subunit 2, PA28beta	27362	94	9	Chaperone/stress, Regulatory	Cytoplasmic
O00764	PDXK_HUMAN	PDXK	Pyridoxal kinase	35102	60	5	Enzyme	Cytoplasmic
O75608	LYPA1_HUMAN	LYPA1	Acyl-protein thioesterase 1	24670	58	4	Enzyme	Cytoplasmic
P00558	PGK1_HUMAN	PGK1	Phosphoglycerate kinase 1	44615	160	18	Enzyme	Cytoplasmic
P04075	ALDOA_HUMAN	ALDOA	Fructose-bisphosphate aldolase A	39420	107	14	Enzyme	Cytoplasmic
P07195	LDHB_HUMAN	LDHB	L-lactate dehydrogenase B chain	36638	130	14	Enzyme	Cytoplasmic
P07741	APT_HUMAN	ATP	Adenine phosphoribosyltransferase	19608	137	12	Enzyme	Cytoplasmic
P09211	GSTP1_HUMAN	GSTP1	Glutathione S-transferase P	23356	113	12	Enzyme	Cytoplasmic
P09972	ALDOC_HUMAN	ALDOC	Fructose-bisphosphate aldolase C	39456	111	12	Enzyme	Cytoplasmic
P10599	THIO_HUMAN	THIO	Thioredoxin	11737	70	6	Enzyme	Cytoplasmic
P10768	ESTD_HUMAN	ESTD	S-formylglutathione hydrolase, Esterase D	31463	72	6	Enzyme	Cytoplasmic
P11586	CITC_HUMAN	CITC	C-1-tetrahydrofolate synthase, cytoplasmic	101559	151	19	Enzyme	Cytoplasmic
P12268	IMDH2_HUMAN	IMDH2	Inosine-5'-monophosphate dehydrogenase 2	55805	168	13	Enzyme	Cytoplasmic
P12277	KCRB_HUMAN	KCRB	Creatine kinase B-type	42644	88	10	Enzyme	Cytoplasmic
P13798	ACPH_HUMAN	ACPH	Acylamino-acid-releasing enzyme, Acyl-peptide hydrolase	81225	89	15	Enzyme	Cytoplasmic
P14550	AK1A1_HUMAN	AK1A1	Alcohol dehydrogenase [NADP+]	36573	92	10	Enzyme	Cytoplasmic
P14618	KPYM_HUMAN	KPYM	Pyruvate kinase isozymes M1/M2	57937	128	14	Enzyme	Cytoplasmic
P18669	PGAM1_HUMAN	PGAM1	Phosphoglycerate mutase 1, Phosphoglycerate mutase isozyme B	28804	103	10	Enzyme	Cytoplasmic
P28838	AMPL_HUMAN	AMPL	Cytosol aminopeptidase	56166	132	13	Enzyme	Cytoplasmic
P30043	BLVRB_HUMAN	BLVRB	Flavin reductase, Biliverdin reductase B	22119	88	9	Enzyme	Cytoplasmic
P32119	PRDX2_HUMAN	PRDX2	Peroxiredoxin-2	21892	109	7	Enzyme	Cytoplasmic
P49189	AL9A1_HUMAN	AL9A1	4-trimethylaminobutyraldehyde dehydrogenase	53802	66	7	Enzyme	Cytoplasmic
P49915	GUAA_HUMAN	GUAA	Gamma-aminobutyraldehyde dehydrogenase	76715	265	25	Enzyme	Cytoplasmic
P51570	GALK1_HUMAN	GALK1	GMP synthase [glutamine-hydrolyzing] Galactokinase	42272	74	6	Enzyme	Cytoplasmic

Table I. *continued*

Table I. continued

Accession no	Protein name	Protein symbol	Protein description	MW	Mascot score	Peptides matched	Cellular function	Subcellular localization
P55036	PSD4_HUMAN	PSD4	26S proteasome non-ATPase regulatory subunit 4	40737	87	10	Enzyme	Cytoplasmic
Q06830	PRDX1_HUMAN	PRDX1	Peroxiredoxin-1	22110	201	20	Enzyme	Cytoplasmic
Q13162	PRDX4_HUMAN	PRDX4	Peroxiredoxin-4	30540	105	9	Enzyme	Cytoplasmic
Q15102	PA1B3_HUMAN	PA1B3	Platelet-activating factor acetylhydrolase IB subunit gamma	25734	63	7	Enzyme	Cytoplasmic
Q15181	IPYR_HUMAN	IPYR	Inorganic pyrophosphatase	32660	180	19	Enzyme	Cytoplasmic
Q92871	PMM1_HUMAN	PMM1	Phosphomannomutase 1, PMM 1	29747	66	5	Enzyme	Cytoplasmic
Q9BX68	HINT2_HUMAN	HINT2	Histidine triad nucleotide-binding protein 2, HINT-2	17162	69	4	Enzyme	Cytoplasmic
P04632	CPNS1_HUMAN	CPNS1	Calpain small subunit 1, Calcium-dependent protease small subunit 1	28316	56	5	Enzyme, Regulatory	Cytoplasmic
P30041	PRDX6_HUMAN	PRDX6	Peroxiredoxin-6, 1-Cys peroxiredoxin, Antioxidant protein 2	25035	191	17	Enzyme, Regulatory	Cytoplasmic
P06733	ENOA_HUMAN	ENOA	Alpha-enolase, C-myc promoter-binding protein, MBP-1	47169	211	20	Enzyme, Structural, Antigen, Antioncogene	Cytoplasmic
Q14247	SRC8_HUMAN	SRC8	Sic substrate contractin, Oncogene EMS1	61636	75	11	Protooncogene	Cytoplasmic
Q76003	TXNL2_HUMAN	TXNL2	Thioredoxin-like protein 2, PKC-theta-interacting protein	37432	90	11	Regulatory	Cytoplasmic
P31947	1433S_HUMAN	1433S	14-3-3 protein sigma	27774	137	19	Regulatory	Cytoplasmic
P35080	PROF2_HUMAN	PROF2	Profilin-2	15046	56	4	Regulatory	Cytoplasmic
P63104	1433Z_HUMAN	1433Z	14-3-3 protein zeta/delta	27745	122	18	Regulatory	Cytoplasmic
Q9BT178	CSN4_HUMAN	CSN4	COP9 signalosome complex subunit 4	46269	107	12	Regulatory	Cytoplasmic
Q9UMX0	UBQL1_HUMAN	UBQL1	Ubiquilin-1, hPLIC-1	62519	77	14	Regulatory	Cytoplasmic
P52565	GDIR_HUMAN	GDIR	Rho GDP-dissociation inhibitor 1, Rho GDI 1	23207	74	9	Regulatory, Signaling	Cytoplasmic
Q9Y696	CLIC4_HUMAN	CLIC4	Chloride intracellular channel protein 4, Intracellular chloride ion channel protein p64H1	28772	145	16	Regulatory, Transport channel	Cytoplasmic
P13639	EF2_HUMAN	EF2	Elongation factor 2	95338	84	12	RNA associated	Cytoplasmic
P62988	UBIQ_HUMAN	UBIQ	Ubiquitin	8565	75	5	RNA associated	Cytoplasmic
Q12849	GRSF1_HUMAN	GRSF1	G-rich sequence factor 1, GRSF-1	50170	72	10	RNA associated	Cytoplasmic
Q3SX47	Q3SX47_BOVIN	Q3SX47	Heterogeneous nuclear ribonucleoprotein C	32428	66	9	RNA associated	Cytoplasmic
Q76M58	Q76M58_HUMAN	Q76M58	40S ribosomal protein S12	14515	102	10	RNA associated	Cytoplasmic
Q13347	IF32_HUMAN	IF32	Eukaryotic translation initiation factor 3 subunit 2, eIF3i, TRIP-1	36502	133	12	RNA associated, Antioncogene	Cytoplasmic
Q51ST8	Q51ST8_MACFA	Q51ST8	Heterogeneous nuclear ribonucleoprotein C [Fragment]	29266	65	8	RNA associated, DNA associated	Cytoplasmic
P02792	FRIL_HUMAN	FRIL	Ferritin light chain	202020	97	7	Storage	Cytoplasmic
P07437	TBB2_HUMAN	TBB2	Tubulin beta chain	49671	238	37	Structural	Cytoplasmic
P12814	ACTN1_HUMAN	ACTN1	Alpha-actinin-1	103058	59	16	Structural	Cytoplasmic
P63261	ACTG_HUMAN	ACTG	Actin, cytoplasmic 2, Gamma-actin	41793	172	21	Structural	Cytoplasmic
Q16643	DREB_HUMAN	DREB	Drebrin	71425	92	15	Structural	Cytoplasmic
Q01469	FABPE_HUMAN	FABPE	Fatty acid-binding protein, epidermal,	15164	154	18	Transport Carrier	Cytoplasmic
Q53FT3	CK073_HUMAN	CK073	Psoriasis-associated fatty acid-binding protein homolog	21628	57	5	Unknown/Unspecified	Cytoplasmic
Q9BZE9	Q9BZE9_HUMAN	Q9BZE9	Uncharacterized protein C11orf73	60183	86	10	Unknown/Unspecified	Cytoplasmic
			Tether containing UBX domain for GLUT4, Alveolar soft part sarcoma locus					

Table I. continued

Table I. *continued*

Accession no	Protein name	Protein symbol	Protein description	MW	Mascot score	Peptides matched	Cellular function	Subcellular localization
P09104	ENOG_HUMAN	ENOGENE	Gamma-enolase	47269	106	11	Enzyme	Cytoplasmic, Cell membrane
O94760	DDAH1_HUMAN	DDAH1	NG,NG-dimethylarginine dimethylaminohydrolase 1, DDAH-1 Activator of 90 kDa heat shock protein ATPase homolog 1, p38, AHAI	31122	72	8	Enzyme	Cytosolic, Cytoplasmic, Endoplasmic reticulum
O95433	AHSA1_HUMAN	AHSA1		38274	59	7	Chaperone/stress, Regulatory	Cytoplasmic, Membrane
O75832	PSD10_HUMAN	PSD10	26S proteasome non-ATPase regulatory subunit 10, Gankyrin Annexin A5	24428	103	8	Regulatory, Protooncogene	Cytoplasmic, Extracellular
P08758	ANXA5_HUMAN	ANXA5		35937	221	18	Regulatory, Transport channel	Cytoplasmic, Membrane
P23528	COF1_HUMAN	COF1	Coflin-1, p18	18502	128	12	Structural	Cytoplasmic, Mitochondrial, Nuclear
Q14847	LASPI_HUMAN	LASPI	LIM and SH3 domain protein 1, LASP-1	29717	82	13	Adaptor	Cytoplasmic, Nuclear
P31948	STIP1_HUMAN	STIP1	Stress-induced-phosphoprotein 1, STI1, Hsc70/Hsp90-organizing protein	62639	264	43	Adaptor, Chaperone/Stress	Cytoplasmic, Nuclear
Q14192	FHL2_HUMAN	FHL2	Four and a half LIM domains protein 2, Skeletal muscle LIM-protein 3	32193	93	8	Adaptor, Regulator, Signaling	Cytoplasmic, Nuclear
P04792	HSPB1_HUMAN	HSPB1	Heat-shock protein beta-1, Heat-shock 27 kDa protein	22783	120	13	Chaperone/stress	Cytoplasmic, Nuclear
P11142	HSP7C_HUMAN	HSP7C	Heat-shock cognate 71 kDa protein	70898	247	35	Chaperone/stress	Cytoplasmic, Nuclear
P49321	NASP_HUMAN	NASP	Nuclear autoantigenic sperm protein, NASP	85238	144	23	Chaperone/stress	Cytoplasmic, Nuclear
P61758	PFD3_HUMAN	PFD3	Prefoldin subunit 3, Von Hippel-Lindau-binding protein 1, VBP-1	22658	74	6	Chaperone/stress	Cytoplasmic, Nuclear
P62333	PRS10_HUMAN	PRS10	26S protease regulatory subunit S10B	44173	94	12	Chaperone/Stress, Enzyme	Cytoplasmic, Nuclear
P17980	PRS6A_HUMAN	PRS6A	26S protease regulatory subunit 6A, Tat-binding protein 1	49204	56	14	Chaperone/Stress, Transcription factor	Cytoplasmic, Nuclear
O14818	PSA7_HUMAN	PSA7	Proteasome subunit alpha type 7	27887	73	7	Enzyme	Cytoplasmic, Nuclear
P04406	G3P_HUMAN	G3P	Glyceraldehyde-3-phosphate dehydrogenase, GAPDH	36053	149	15	Enzyme	Cytoplasmic, Nuclear
P22392	NDKB_HUMAN	NDKB	Nucleotide diphosphate kinase B	17298	97	12	Enzyme	Cytoplasmic, Nuclear
P25786	PSA1_HUMAN	PSA1	Proteasome subunit alpha type 1	29556	75	10	Enzyme	Cytoplasmic, Nuclear
P28070	PSB4_HUMAN	PSB4	Proteasome subunit beta type 4 [Precursor]	29204	69	7	Enzyme	Cytoplasmic, Nuclear
P48637	GSHB_HUMAN	GSHB	Glutathione synthetase	52385	75	9	Enzyme	Cytoplasmic, Nuclear
Q9Y265	RUVBL1_HUMAN	RUVBL1	RuvB-like 1, TIP49a	50228	104	15	Enzyme, DNA associated, Antigen	Cytoplasmic, Nuclear
Q96AE4	FUBP1_HUMAN	FUBP1	Far upstream element-binding protein 1, FUBP	67560	210	26	Protooncogene	Cytoplasmic, Nuclear
Q99497	PARK7_HUMAN	PARK7	Protein DJ-1, Oncogene DJ1	19891	134	19	Protooncogene, Signaling neurotransmitter	Cytoplasmic, Nuclear
Q92945	FUBP2_HUMAN	FUBP2	Far upstream element-binding protein 2, KSRP, FUSE-binding protein 2	72709	135	15	Regulatory, RNA associated	Cytoplasmic, Nuclear
Q15637	SF01_HUMAN	SF01	Splicing factor 1, Transcription factor ZFM1	68330	61	7	Regulatory, Transcription factor	Cytoplasmic, Nuclear
O43390	HNRPR_HUMAN	HNRPR	Heterogeneous nuclear ribonucleoprotein R	70943	61	17	RNA associated	Cytoplasmic, Nuclear

Table I. *continued*

Table I. *continued*

Accession no	Protein name	Protein symbol	Protein description	MW	Mascot score	Peptides matched	Cellular function	Subcellular localization
P07910	HNRPC_HUMAN	HNRPC	Heterogeneous nuclear ribonucleoproteins C1/C2	33670	60	8	Cytoplasmic, Nuclear	
P14866	HNRPL_HUMAN	HNRPL	Heterogeneous nuclear ribonucleoprotein L	60187	174	20	Cytoplasmic, Nuclear	
P52272	HNRPM_HUMAN	HNRPM	Heterogeneous nuclear ribonucleoprotein M, hnRNP M	77516	108	30	Cytoplasmic, Nuclear	
P63241	IF5A1_HUMAN	IF5A1	Eukaryotic translation initiation factor 5A-1, eIF-5A-1	16832	77	7	Cytoplasmic, Nuclear	
Q14103	HNRPD_HUMAN	HNRPD	Heterogeneous nuclear ribonucleoprotein D0, AUF1	38434	62	5	Cytoplasmic, Nuclear	
Q15365	PCBP1_HUMAN	PCBP1	Poly(rC)-binding protein 1, Alpha-CP1	37498	133	15	Cytoplasmic, Nuclear	
Q15366	PCBP2_HUMAN	PCBP2	Poly(rC)-binding protein 2, Alpha-CP2	38580	78	6	Cytoplasmic, Nuclear	
Q8NC51	PAIRB_HUMAN	PAIRB	Plasminogen activator inhibitor 1 RNA-binding protein	44965	99	16	Cytoplasmic, Nuclear	
P11940	PABP1_HUMAN	PABP1	Polyadenylate-binding protein 1	70671	108	15	Cytoplasmic, Nuclear	
P19338	NUCL_HUMAN	NUCL	Nucleolin	76614	142	22	Cytoplasmic, Nuclear	
P39687	AN32A_HUMAN	AN32A	Acidic leucine-rich nuclear phosphoprotein 32 family member A, Mapmodulin, Acidic nuclear phosphoprotein pp32	28585	70	7	Cytoplasmic, Nuclear	
P51858	HDGF_HUMAN	HDGF	Hepatoma-derived growth factor	26788	87	10	Signaling growth factor	
Q14764	MVP_HUMAN	MVP	Major vault protein	99327	150	24	Structural, RNA associated, Transport	
O96IU4	ABHEB_HUMAN	ABHEB	Alhydrolase domain-containing protein 14B	22346	68	5	Cytoplasmic, Nuclear	
O00233	PSMD9_HUMAN	PSMD9	26S proteasome non-ATPase regulatory subunit 9	24654	81	10	Unknown/Unspecified Enzyme	
P08865	RSSA_HUMAN	RSSA	40S ribosomal protein SA, 34/67 kDa laminin receptor	32854	118	12	Adhesion, Signaling, Receptor	
P06396	GELS_HUMAN	GELS	Gelsolin [Precursor], ADF	85698	63	7	Regulatory, Structural	
P18206	VINC_HUMAN	VINC	Vinculin	123799	70	19	Adhesion	
P07951	TPM2_HUMAN	TPM2	Tropomyosin beta chain	32851	88	12	Motor/Contractile	
P60709	ACTB_HUMAN	ACTB	Actin, cytoplasmic 1	41737	133	22	Motor/Contractile, Structural	
P05783	K1C18_HUMAN	K1C18	Keratin, type I cytoskeletal 18, Cytokeratin-18	48058	110	19	Structural	
P08670	VIME_HUMAN	VIME	Vimentin	53652	334	39	Structural	
P13645	K1C10_HUMAN	K1C10	Keratin, type I cytoskeletal 10	59519	67	12	Structural	
P35527	K1C9_HUMAN	K1C9	Keratin, type I cytoskeletal 9, Keratin-9	62129	69	7	Structural	
P47756	CAPZB_HUMAN	CAPZB	F-actin capping protein subunit beta	31350	103	10	Structural	
P52907	CAZAI_HUMAN	CAZAI	F-actin capping protein subunit alpha-1	32923	147	11	Structural	
Q15417	CNN3_HUMAN	CNN3	Calponin-3	36414	120	18	Structural	
P06748	NPM_HUMAN	NPM	Nucleophosmin	32575	74	12	Protooncogene	
Q13561	DCTN2_HUMAN	DCTN2	Dynactin subunit 2	44231	69	8	Motor/Contractile, Transport	
P49773	HINT1_HUMAN	HINT1	Histidine triad nucleotide-binding protein 1, Protein kinase C inhibitor 1	13802	76	5	Regulatory, Regulatory	
O9Y6D9	MDIL1_HUMAN	MDIL1	Mitotic spindle assembly checkpoint protein MAD1	83067	72	15	Antioncogene	
Q03252	LMNB2_HUMAN	LMNB2	Lamin B2	67689	181	23	Structural	

Table I. *continued*

Table I. *continued*

Accession no	Protein name	Protein symbol	Protein description	MW	Mascot score	Peptides matched	Cellular function	Subcellular localization
Q15084	PDIA6_HUMAN	PDIA6	Protein disulfide-isomerase A6 [Precursor]	48121	136	17	Chaperone/Stress	Endoplasmic reticulum
P14625	ENPL_HUMAN	ENPL	Endoplasm [Precursor], Heat-shock protein 90 kDa beta member 1, GRP94 Caereticulin [Precursor]	92469	151	29	Chaperone/Stress, Antigen	Endoplasmic reticulum
P27797	CALR_HUMAN	CALR	Superoxide dismutase [Cu-Zn]	48142	163	19	Chaperone/Stress, Antigen	Endoplasmic reticulum
P00441	SODC_HUMAN	SODC	Protein disulfide-isomerase A3 [Precursor], ERp60	15936	102	8	Chaperone/stress, Enzyme	Endoplasmic reticulum
P30101	PDIA3_HUMAN	PDIA3	Protein disulfide-isomerase A3 [Precursor], ERp60 [Precursor], ERp28	56782	204	24	Chaperone/Stress, Enzyme	Endoplasmic reticulum
P30040	ERP29_HUMAN	ERP29	Endoplasmic reticulum protein ERp29	28993	86	5	Chaperone/Stress, Enzyme	Endoplasmic reticulum
P62937	PPIA_HUMAN	PPIA	Pepidyl-t-prolyl cis-trans isomerase A, Cyclophilin A	18012	161	13	Enzyme, Transport, Enzyme, Immunity/defense	Endoplasmic reticulum
P07237	PDIA1_HUMAN	PDIA1	Protein disulfide-isomerase [Precursor]	57116	214	26	Enzyme, Protooncogene	Endoplasmic reticulum
P50454	SPH2_HUMAN	SPH2	Serpin H1 [Precursor], Collagen-binding protein, Colligin	46441	68	10	Enzyme, Regulatory	Endoplasmic reticulum
O95881	TXD12_HUMAN	TXD12	Thioredoxin domain-containing protein 12 [Precursor]	19206	68	8	Enzyme, Transport	Endoplasmic reticulum
Q15293	RCN1_HUMAN	RCN1	Reticulocalbin-1 [Precursor]	38890	130	17	Regulatory	Endoplasmic reticulum
P24534	EF1B_HUMAN	EF1B	Elongation factor 1-beta, EF-1-beta	24764	68	7	RNA associated, Transcription factor	Endoplasmic reticulum
Q14257	RCN2_HUMAN	RCN2	Reticulocalbin-2 [Precursor], E6-binding protein	36876	130	13	Structural	Endoplasmic reticulum
P14314	GLU2B_HUMAN	GLU2B	Glucosidase 2 subunit beta [Precursor]	59425	95	16	Unknown/Unspecified	Endoplasmic reticulum
Q9Y4L1	OXRP_HUMAN	OXRP	150 kDa oxygen-regulated protein [Precursor]	111335	111	21	Chaperone/Stress, Protooncogene	Endoplasmic reticulum, Golgi apparatus
Q14697	GANAB_HUMAN	GANAB	Neutral alpha-glucosidase AB [Precursor]	106874	150	15	Enzyme	Endoplasmic reticulum, Cyttoplasmic
O43852	CALU_HUMAN	CALU	Calumenin [Precursor]	37107	152	14	Regulatory, Transport	Golgi apparatus
P55072	TERA_HUMAN	TERA	Transitional endoplasmic reticulum ATPase, VCP, Valosin-containing protein Protein SET	89322	283	41	Chaperone/Stress, Structural, Transport	Endoplasmic reticulum, Nuclear
Q01105	SET_HUMAN	SET		33489	56	7	Regulatory, DNA associated, Transcription factor, Protooncogene	Endoplasmic reticulum, Nuclear
Q9UKX7	NUP50_HUMAN	NUP50	Nucleoporin 50 kDa	50144	75	9	Structural	Endoplasmic reticulum Nuclear

Table I. *continued*

Table I. *continued*

Accession no	Protein name	Protein symbol	Protein description	MW	Mascot score	Peptides matched	Cellular function	Subcellular localization
Q31349 Q64599 P13693	Q31349_BOVIN Q64599_RAT TCTP_HUMAN	Q31349 Q64599 TCTP	Serum albumin [Fragment] Hemiferritin Translationally-controlled tumor protein, Fortilin	53925 24091 19595	237 74 70	25 7 12	Transport Transport Enzyme, Protooncogene	Extracellular Extracellular Extracellular, Cytoplasmic Extracellular, Membranous, Endoplasmic reticulum
P11021	GRP78_HUMAN	GRP78	78 kDa glucose-regulated protein [Precursor], GRP 78, BiP	72333	312	35	Chaperone/Stress, Signaling	Membranous
O15942 Q9NRX4 P62993	ZYX_HUMAN PHP14_HUMAN GRB2_HUMAN	ZYX PHP14 GRB2	Zyxin 14 kDa phosphohistidine phosphatase Growth factor receptor-bound protein 2, Adapter protein GRB2	61277 13833 25206	71 73 113	10 6 10	Adhesion Regulatory Regulatory, Signaling growth factor	Membranous Membranous Membranous
P07355	ANXA2_HUMAN	ANXA2		38604	123	14	Regulatory, Signaling, Transport channel	Membranous
Q14444 P09382	CAPR1_HUMAN LEG1_HUMAN	CAPR1 LEG1	Caprin-1, p137/GPI Galecin-1	72752 14716	60 154	10 13	Transport Adhesion, Regulatory	Membranous
Q13283	G3BP_HUMAN	G3BP	Ras GTPase-activating protein-binding protein 1, HDH-VIII, G3BP-1	52164	177	20	Enzyme	Cytoplasmic, Nuclear Membranous, Nuclear
P10809 P05091 P19404	CH60_HUMAN ALDH2_HUMAN NUHM_HUMAN	CH60 ALDH2 NUHM	60 kDa heat shock protein, mitochondrial [Precursor] Aldehyde dehydrogenase, mitochondrial NADH dehydrogenase [ubiquinone] flavoprotein 2, mitochondrial [Precursor]	61055 56381 27392	266 60 58	34 8 7	Chaperone/Stress Enzyme Enzyme	Mitochondrial Mitochondrial Mitochondrial
P25325 P28331	THTM_HUMAN NUAM_HUMAN	THTM NUAM	3-mercaptopyruvate sulfurtransferase NADH-ubiquinone oxidoreductase 75 kDa subunit, mitochondrial [Precursor]	33178 79468	79 159	6 17	Enzyme Enzyme	Mitochondrial Mitochondrial
P30084 P30837 P31930	ECHM_HUMAN AL1B1_HUMAN UQCRI_HUMAN	ECHM AL1B1 UQCRI	Enoyl-CoA hydratase, mitochondrial [Precursor] Aldehyde dehydrogenase X, mitochondrial [Precursor] Ubiquinol-cytochrome c reductase complex core protein 1, mitochondrial [Precursor], Core I protein	31387 57238 52646	102 76 154	13 7 21	Enzyme Enzyme Enzyme	Mitochondrial Mitochondrial Mitochondrial
P31937	3HIDH_HUMAN	3HIDH	3-hydroxyisobutyrate dehydrogenase, mitochondrial [Precursor]	35329	69	9	Enzyme	Mitochondrial
P32322 P36639	P5CR1_HUMAN 8ODP_HUMAN	P5CR1 8ODP	Pyrrrole-5-carboxylate reductase 1 7,8-dihydro-8-oxoguanine triphosphatase, 8-oxo-dGTPase	33361 22552	101 60	8 5	Enzyme Enzyme	Mitochondrial Mitochondrial
P36957	ODO2_HUMAN	ODO2	Dihydrolipooylysine-residue succinyltransferase component of 2-oxoglutarate dehydrogenase complex, mitochondrial [Precursor], E2K	48640	76	9	Enzyme	Mitochondrial
P50213	IDH3A_HUMAN	IDH3A	Isocitrate dehydrogenase [NAD] subunit alpha, mitochondrial [Precursor], NAD (+)-specific ICEDH	39592	56	7	Enzyme	Mitochondrial
P55809	SCOT_HUMAN	SCOT	Succinyl-CoA:3-ketoacid-coenzyme A transferase 1, mitochondrial [Precursor], Scot-S	56158	59	8	Enzyme	Mitochondrial
Q99798 Q08752	ACON_HUMAN PPID_HUMAN	ACON PPID	Aconitase hydratase, mitochondrial [Precursor] 40 kDa peptidyl-prolyl cis-trans isomerase, Cyclophilin-40, CYP-40	85425 40764	98 65	11 5	Enzyme, Immunity/defense, Receptor	Mitochondrial Mitochondrial

Table I. *continued*

Table I. *continued*

Accession no	Protein name	Protein symbol	Protein description	MW	Mascot score	Peptides matched	Cellular function	Subcellular localization
P06576 Q16891 P07737 P61088 Q04837	ATPB_HUMAN IMMT_HUMAN PROFL_HUMAN UBE2N_HUMAN SSB_HUMAN	ATPB IMMT PROFL UBE2N SSB	ATP synthase subunit beta, mitochondrial [Precursor] Mitochondrial inner membrane protein, Mitofillin Profilin-1 Ubiquitin-conjugating enzyme E2 N, Ubcl3 Single-stranded DNA-binding protein, mitochondrial [Precursor] 14-3-3 protein epsilon, 14-3-3E	56560 83678 15054 17138 17260	184 69 74 105 100	26 8 9 8 9	Enzyme, Transport Motor/Contractile, Structural Regulatory Regulatory Regulatory	Mitochondrial Mitochondrial Mitochondrial Mitochondrial Mitochondrial
P62258 O43175	1433E_HUMAN SERA_HUMAN	1433E SERA	D-3-phosphoglycerate dehydrogenase	29174	111	14	Adaptor, Regulator, Signaling neurotransmitter Enzyme	Mitochondrial, Cytoskeletal Mitochondrial, Endoplasmic reticulum
P38646	GRP75_HUMAN	GRP75	Stress-70 protein, mitochondrial [Precursor], Mortalin	73680	286	40	Chaperone/Stress, Transcription factor, Regulatory, Protooncogene	Mitochondrial, Endoplasmic reticulum,
P35232	PHB_HUMAN	PHB	Prohibitin	29804	132	12	Antioncogene	Cytoplasmic Mitochondrial, Endoplasmic reticulum,
P08107	HSP71_HUMAN	HSP71	Heat-shock 70 kDa protein 1	70052	161	18	Chaperone/Stress	Nuclear Mitochondrial, Endoplasmic reticulum,
P61604	CH10_HUMAN	CH10	10 kDa heat-shock protein, mitochondrial	10932	137	11	Chaperone/stress	Nuclear
Q13765	NACA_HUMAN	NACA	Nascent polypeptide-associated complex subunit alpha, NAC-alpha	23384	58	5	Chaperone/Stress	Mitochondrial, Matrix
P15531	NDKA_HUMAN	NDKA	Nucleoside diphosphate kinase A, nm23-HI	17149	152	15	Enzyme	Mitochondrial, Nuclear
P33316	DUT_HUMAN	DUT	Deoxyuridine 5'-triphosphate nucleotidohydrolase, mitochondrial [Precursor], dUTP pyrophosphatase	26706	133	12	Enzyme	Mitochondrial, Nuclear
Q07021	C1QBP_HUMAN	C1QBP	Complement component Q subcomponent-binding protein, mitochondrial [Precursor], Glycoprotein gC1qBP, p33	31362	57	7	Immunity/Defence, RNA associated, Signaling Regulatory	Mitochondrial, Nuclear
Q92598	HS105_HUMAN	HS105	Heat-shock protein 105 kDa	96865	101	14		Mitochondrial, Nuclear
Q9Y224	CNI66_HUMAN	CNI66	Protein C14orf166	28068	138	14	Regulatory	Mitochondrial, Nuclear
P29692	EFID_HUMAN	EFID	Elongation factor 1-delta	31122	179	16	RNA associated	Mitochondrial, Nuclear
Q8WXX5 P15927 O15160 P17844	DNJC9_HUMAN RFA2_HUMAN RPA5_HUMAN DDX5_HUMAN	DNJC9 RFA2 RPA5 DDX5	Dna J homolog subfamily C member 9 Replication protein A 32 kDa subunit, RP-A DNA-directed RNA polymerase I 40 kDa polypeptide Probable ATP-dependent RNA helicase DDX5, RNA helicase p68	29910 29247 39250 69148	65 68 112 60	6 4 12 9	Chaperone/Stress DNA associated Enzyme Enzyme	Nuclear Nuclear Nuclear Nuclear
P12004 P43487	PCNA_HUMAN RANG_HUMAN	PCNA RANG	Proliferating cell nuclear antigen Ran-specific GTPase-activating protein, RanBP1	28769 23310	89 60	10 9	Regulatory, DNA associated, Antigen Regulatory, Signaling	Nuclear Nuclear

Table I. *continued*

Table I. continued

Accession no	Protein name	Protein symbol	Protein description	MW	Mascot score	Peptides matched	Cellular function	Subcellular localization
Q13263	TIF1B_HUMAN	TIF1B	Transcription intermediary factor 1-beta, KRAB-associated protein 1, KAP-1	88550	129	17	Regulatory, Transcription factor	Nuclear
O43809	CPSF5_HUMAN	CPSF5	Cleavage and polyadenylation specificity factor 5	26227	56	5	RNA associated	Nuclear
P06730	IF4E_HUMAN	IF4E	Eukaryotic translation initiation factor 4E	25097	63	6	RNA associated	Nuclear
P22626	ROA2_HUMAN	ROA2	Heterogeneous nuclear ribonucleoproteins A2/B1	37430	148	11	RNA associated	Nuclear
P31942	HNRH3_HUMAN	HNRH3	Heterogeneous nuclear ribonucleoprotein H3	36926	118	18	RNA associated	Nuclear
P31943	HNRH1_HUMAN	HNRH1	Heterogeneous nuclear ribonucleoprotein H	49229	124	17	RNA associated	Nuclear
P33240	CSTF2_HUMAN	CSTF2	Cleavage stimulation factor 64 kDa subunit, Cstf-64	60959	90	13	RNA associated	Nuclear
P52597	HNRPF_HUMAN	HNRPF	Heterogeneous nuclear ribonucleoprotein F, hnRNP F	45672	68	11	RNA associated	Nuclear
P55795	HNRH2_HUMAN	HNRH2	Heterogeneous nuclear ribonucleoprotein H'	49264	129	13	RNA associated	Nuclear
P61978	HNRPK_HUMAN	HNRPK	Heterogeneous nuclear ribonucleoprotein K, hnRNP K	50976	124	16	RNA associated	Nuclear
Q07955	SFRS1_HUMAN	SFRS1	Splicing factor, arginine/serine-rich 1, ASF-1	27745	155	13	RNA associated	Nuclear
Q12874	SF3A3_HUMAN	SF3A3	Splicing factor 3A subunit 3, SF3a60	58849	83	13	RNA associated	Nuclear
Q15459	SF3A1_HUMAN	SF3A1	Splicing factor 3 subunit 1, SF3a120	88886	116	18	RNA associated	Nuclear
Q96S43	HNRDL_HUMAN	HNRDL	Heterogeneous nuclear ribonucleoprotein D-like, hnRNP-DL, JK741-binding protein	46438	64	7	RNA associated	Nuclear
P23246	SFPQ_HUMAN	SFPQ	Splicing factor, proline- and glutamine-rich, PSF	76149	85	20	RNA associated, Antionogene	Nuclear
P67809	YBOX1_HUMAN	YBOX1	Nuclease sensitive element-binding protein 1, YB-1	35924	123	13	RNA associated, Transcription factor	Nuclear
O75934	BCAS2_HUMAN	BCAS2	Breast carcinoma amplified sequence 2	26131	81	8	Structural	Nuclear
P43243	MATR3_HUMAN	MATR3	Matrin-3	94623	159	19	Structural	Nuclear
P09429	HMGB1_HUMAN	HMGB1	High mobility group protein B1	24894	77	8	Transcription factor	Nuclear
Q7Z3B4	NUP54_HUMAN	NUP54	Nucleoporin p54	55435	123	13	Transport	Nuclear
Q9UNZ2	NSF1C_HUMAN	NSF1C	NSFL1 cofactor p47	40573	90	10	Adaptor	Golgi apparatus
P02545	LMNA_HUMAN	LMNA	Lamin-A/C	74139	295	36	Structural	Nuclear, Nuclear matrix
P04083	ANXA1_HUMAN	ANXA1	Annexin A1	38714	68	4	Regulatory, Transport Chaperone/stress	Membranic
O95816	BAG2_HUMAN	BAG2	BAG family molecular chaperone regulator 2, BAG-2	23772	77	8	Regulatory, Transport Enzyme	
O75947	ATPSH_HUMAN	ATPSH	ATP synthase D chain, mitochondrial	18491	115	12	Chaperone/stress	
P22314	UBE1_HUMAN	UBE1	Ubiquitin-activating enzyme E1	117849	102	13	Enzyme	Mitochondrial
P60174	TPIS_HUMAN	TPIS	Triosephosphate isomerase	26669	210	14	Enzyme	
Q9UBQ7	GRHPR_HUMAN	GRHPR	Glyoxylate reductase/hydroxypyruvate reductase	35668	94	8	Enzyme	
P84090	ERH_HUMAN	ERH	Enhancer of rudimentary homolog	12259	64	7	Regulatory	
P08133	ANXA6_HUMAN	ANXA6	Annexin A6, P68	75873	184	30	Regulatory, Transport	
P60660	MYL6_HUMAN	MYL6	Myosin light polypeptide 6, Smooth muscle and nonmuscle myosin light chain alkali 6	16930	94	8	Structural	
P68363	TBAK_HUMAN	TBAK	Tubulin alpha1B chain	50152	207	26	Structural	
P68371	TBB2C_HUMAN	TBB2C	Tubulin beta2C chain	49831	236	32	Structural	
P9HB07	MYG1_HUMAN	MYG1	UPF0160 protein MYG1	42445	82	9	Unknown/Unspecified	
P37802	TAGL2_HUMAN	TAGL2	Transgelin-2	22391	121	14	Unknown/Unspecified, Transcription factor	
O5HYB6	O5HYB6_HUMAN	O5HYB6	Hypothetical protein DKFZp686J1372	27176	112	15		
Q5T626	Q5T626_HUMAN	Q5T626	Nuclear autoantigenic sperm protein	48804	70	11		
Q96B89	GLOD4_HUMAN	GLOD4	Glyoxalase domain-containing protein 4	34793	123	13		
Q9BQE3	TBA6_HUMAN	TBA6	Tubulin alpha1C chain	49895	68	8		
Q9U130	TR112_HUMAN	TR112	TRM112-like protein	14199	64	4		

proteins, structural proteins, such as tubulins and actins, and other major classes of identified proteins, such as transcription factors, transport/carrier and signal transduction. In addition, we found a number of protooncogenes and antioncogenes, representing 3% and 1%, respectively of the total proteome.

Discussion

For the comprehensive analysis of the U2OS osteosarcoma cell line we decided to apply proteomics technology as it provides an insight into the relationships between genes, their products and cell function (16). Analytical methods used for proteomic research result most commonly in inclusive databases such as 2-DE maps. Genome sequence databases, complete catalogues of proteins expressed in organisms, mass spectrometry and software that match MS data with protein sequences databases helped us to successfully complete the construction of the protein database of the osteosarcoma U2OS cell line.

We identified 237 different gene products with several functions including regulatory, signal transduction, protooncogenes and antioncogenes, chaperone/stress and nucleic acid-binding proteins. The use of internal peptide standards allowed narrow windows of mass tolerance (0.0025%), increasing the confidence of identification by PMF and PSD mode of mass spectrometry. Thus the identification was based on three or more (up to 69) matching peptides. Most of them were localized in the cytoplasm, nucleus, mitochondria and some in the membrane.

Eleven protooncogenes were identified among them, TCTP, SRC8, FUBP1, NPM, PARK7, SET, FKBP4 and OXRP. The SRC8 (*EMS1*) protooncogene encodes a human homologue of cortactin, a c-Src substrate associated with cortical cytoskeleton (Q14247). This protein binds components of the actin-related protein (Arp) 2/3 complex which regulates the assembly and structure of actin networks. Cortactin also interacts with a variety of proteins depending on the cell type. The gene encoding SRC8, named as *EMS1*, is very often overexpressed and amplified in many tumors. Dysregulation may lead to increased tumor cell motility and invasiveness (17, 18). FUBP1, a far upstream element binding protein1 (Q96AE4), complexes with FUSE and inhibits c-myc expression which is also involved in cell growth, proliferation, differentiation and apoptosis (19). The location of the FUBP1 gene is in chromosome 1 p31.1, which is very often amplified in osteosarcomas (20). In addition, in this amplified chromosome area, (1p36.33-p36.12) is located protooncogene PARK7 (DJ-1). DJ-1 protein was shown (21) to be a potent inhibitor of the

Daxx/ASK1 cell death signaling pathway, thus protecting cells from oxidative stress, and functions as a survival factor, hence promoting tumor growth.

Another protooncogene identified is nucleophosmin (NPM, P06748), which is present in actively proliferating cells including tumor cells. NPM is a multifunctional protein involved in ribosome assembly, pre-ribosomal RNA processing, DNA duplication, nucleocytoplasmic protein trafficking and centrosome duplication (22). It is induced by genotoxic stress and stabilizes certain conformers of p53, binds pRb and synergistically stimulates DNA polymerase α (23). In addition, NPM protects cells from death and stress-induced apoptosis through inhibition of p53 (24, 25). TCTP is a ubiquitously expressed protein and is regulated both at the transcriptional and translational level. Thus TCTP protein cellular levels are highly regulated in response to numerous extracellular signals and cellular conditions. TCTP is believed to play an important role in cell growth and division as it is considerably up-regulated upon entry of cells into the cell cycle. Therefore, TCTP down-regulation was associated with reversion of transformed cells to a normal phenotype and suppression of malignant transformation (26). This strongly suggests an implication of TCTP in the malignant phenotype and tumor growth. This is strongly supported by the fact that TCTP is stabilized by the anti-apoptotic protein MCL1 and by a correlation of its level with drug resistance in melanoma cells (26).

A comparison of the U2OS proteome with that of Saos2 (5) revealed some differences despite the same origin of the two cancer cell lines. Concerning the whole proteome, 141 proteins were identical, while 304 proteins were different. Nevertheless both cancer cell lines have great similarities regarding the protooncogenes; they share 7 of the same protooncogenes; while 3 are exclusive to Saos2 and 4 to U2OS. Additionally, there were some differences in the presence of antioncogenes as 6 exist in U2OS and 9 in Saos2; five of these were identical. Consequently, in spite of similarities and the osteogenic origin of these two cell lines there were many differences concerning the proteome, which reflects the differences of the genome and character of the two types of cancer: Saos2 is more aggressive than U2OS.

Eleven proteins with unknown/unspecified subcellular function and/or localization were also identified. One was a hypothetical protein (Q5HYB6) with an interest for further investigation.

Summarizing, in the present study, we created the 2-DE database for the human osteosarcoma U2OS cell line. The 237 different gene products were identified using MALDI-MS and MALDI-MS-MS analysis of approximately 3,000 spots out of four 2-DE gels. This 2-DE database creates a useful tool in the study of molecular carcinogenesis.

References

- 1 Görg A, Weiss W and Dunn MJ: Current two-dimensional electrophoresis technology for proteomics. *Proteomics* 4: 3665-3685, 2004.
- 2 Friedman KM and Fox BA: The promising future of proteomics in cancer diagnosis and treatment. *Eur J Gastroenterol Hepatol* 17: 701-703, 2005.
- 3 Anagnostopoulos AK, Vougas K, Kolialexi A, Mavrou A, Fountoulakis M and Tsangaris GT: The protein profile of the human immature T-cell line CCRF-CEM. *Cancer Genom Proteom* 2: 1-29, 2005.
- 4 Fountoulakis M, Tsangaris G, Oh J, Maris A and Lubec G: Protein profile of the HeLa cell line. *J Chrom A* 1038: 247-265, 2004.
- 5 Niforou KN, Anagnostopoulos AK, Vougas K, Kittas C, Gorgoulis VG and Tsangaris GT: The proteome profile of human osteosarcoma Saos2 cell line. *Cancer Genom Proteom* 3: 325-346, 2006.
- 6 Bayani J, Zielenska M, Pandita A, Al-Romaih K, Karaskova J, Harrison K, Bridge JA, Sorensen P, Thorner P and Squire JA: Spectral karyotyping identifies recurrent complex rearrangement of chromosomes 8, 17, 20 in osteosarcomas. *Genes Chromosomes Cancer* 36: 7-16, 2003.
- 7 Wesierska-Gadek J and Schmid G: The subcellular distribution of the p53 tumor suppressor, and organismal ageing. *Cell Mol Biol Lett* 10: 439-453, 2005.
- 8 Zhu L: Tumour suppressor retinoblastoma protein Rb: a transcriptional regulator. *Eur J Cancer* 41: 2415-2427, 2005.
- 9 Isfort RJ, Cody DB, Lovell G and Doersen CJ: Analysis of oncogenes, tumor suppressor genes, autocrine growth factor production and differentiation state of human osteosarcoma cell lines. *Mol Carcinog* 14: 170-178, 1995.
- 10 Furuya K, Ozaki T, Hanamoto T, Hosoda M, Hayashi S, Barker PA, Takano K, Matsumoto M and Nakagawara A: Stabilization of p73 by nuclear I κ B kinase-{alpha} mediates cisplatin-induced apoptosis. *J Biol Chem* 282(25): 18365-18378, 2007.
- 11 Kinsey CG, Bussolati G, Bosco M, Kimura T, Pizzorno MC, Chernin MI, Cassoni P and Novak JF: Constitutive and ligand-induced nuclear localization of oxytocin receptor. *J Cell Mol Med* 11(1): 96-110, 2007.
- 12 Mancini L, Paul-Clark MJ, Rosignoli G, Hannon R, Martin JE, Macintyre I and Perretti M: Calcitonin and prednisolone display antagonistic actions on bone and have synergistic effects in experimental arthritis. *Am J Pathol* 170(3): 1018-1027, 2007.
- 13 Gorgoulis VG, Vassiliou LV, Karakaidos P, Zacharatos P, Kotsinas A, Liloglou T, Venere M, Ditullio RA Jr, Kastrinakis NG, Levy B, Kletsas D, Yoneta A, Herlyn M, Kittas C and Halazonetis TD: Activation of DNA-damage checkpoint and genomic instability in human precancerous lesions. *Nature* 434: 907-913, 2005.
- 14 Bartkova J, Rezaei N, Liontos M, Karakaidos P, Kletsas D, Issaeva N, Vassiliou LVF, Kolettas E, Niforou K, Zoumpourlis VC, Takaoka M, Nakagawa H, Tort F, Fugger K, Johansson F, Sehested M, Andersen CL, Dyrskjot L, Ørntoft T, Lukas J, Kittas C, Helleday T, Halazonetis TD, Bartek J and Gorgoulis VG: Oncogene-induced senescence is part of the tumorigenesis barrier imposed by DNA-damage checkpoints. *Nature* 444(7119): 633-637, 2006.
- 15 Berndt P, Hobohm U and Langen H: Reliable automatic protein identification from matrix-assisted laser desorption/ionization mass spectrometric peptide fingerprints. *Electrophoresis* 20: 3521-3526, 1999.
- 16 Plebani M: Proteomics: The next revolution in laboratory medicine? *Clin Chim Acta* 357: 113-122, 2005.
- 17 Ormandy CJ, Musgrove EA, Hui R, Daly RJ and Sutherland RL: Cyclin D1, EMS1 and 11q13 amplification in breast cancer. *Breast Cancer Res Treat* 78(3): 323-335, 2003.
- 18 Yuan BZ, Zhou X, Zimonjic DB, Durkin ME and Popescu NC: Amplification and overexpression of the EMS1 oncogene, a possible prognostic marker, in human hepatocellular carcinoma. *J Mol Diagn* 5(1): 48-53, 2003.
- 19 He L, Liu J, Collins I, Sanford S, O'Connell B, Benham CJ and Levens D: Loss of FBP function arrests cellular proliferation and extinguishes c-myc expression. *EMBO J* 19(5): 1034-1044, 2000.
- 20 Sandberg AA and Bridge J: Updates in the cytogenesis and molecular genetics of bone and soft tissue tumors: osteosarcoma and related tumors. *Cancer Genet Cytogenet* 145: 1-30, 2003.
- 21 Junn E, Taniguchi H, Jeong BS, Zhao X, Ichijo H and Mouradian MM: Interaction of DJ-1 with Daxx inhibits apoptosis signal-regulating kinase 1 activity and cell death. *Proc Natl Acad Sci USA* 102: 9691-9696, 2005.
- 22 Tarapore P, Shinmura K, Suzuki H, Tokuyama Y, Kim SH, Mayeda A and Fukasawa K: Thr¹⁹⁹ phosphorylation targets nucleophosmin to nuclear speckles and represses pre-mRNA processing. *FEBS Lett* 580: 399-409, 2006.
- 23 Lambert B and Buckle M: Characterization of the interface between nucleophosmin (NPM) and p53: Potential role in p53 stabilization. *FEBS Lett* 580: 345-350, 2006.
- 24 Li J, Zhang X, Sejas DP and Pang Q: Negative regulation of p53 by nucleophosmin antagonizes stress-induced apoptosis in human normal and malignant hematopoietic cells. *Leuk Res* 29: 1415-1423, 2005.
- 25 Li J, Zhang X, Sejas DP, Bagby GC and Pang Q: Hypoxia-induced nucleophosmin protects cell death through inhibition of p53. *J Biol Chem* 279: 41275-41279, 2004.
- 26 Bommer UA and Thiele BJ: The translationally controlled tumor protein (TCTP). *Int J Biochem Cell Biol* 36(3): 379-385 2004.

Received October 24, 2007

Revised December 10, 2007

Accepted January 7, 2008