

Review

Immunogenic Chemotherapy: Discovery of a Critical Protein through Proteomic Analyses of Tumor Cells

LIONEL APETOH^{1,2,3}, MICHEL OBEID^{1,2,4}, ANTOINE TESNIERE^{1,2,4}, FRANÇOIS GHIRINGHELLI^{1,2,3}, GIAN MARIA FIMIA⁵, MAURO PIACENTINI^{1,2,4}, GUIDO KROEMER^{1,2,4} and LAURENCE ZITVOGEL^{1,2,3}

¹Institut Gustave Roussy, Villejuif;

²Faculté Paris Sud-Université Paris XI, Villejuif;

³Institut National de la Santé et de la Recherche Médicale, U805, CIC BT507, Villejuif;

⁴Institut National de la Santé et de la Recherche Médicale, U848, Villejuif, France;

⁵National Institute for Infectious Diseases Lazzaro Spallanzani, Via Portuense 292, I-00149 Rome, Italy

Abstract. *The aim of chemotherapy and radiotherapy is to eliminate tumor cells. While the outcomes of these cytotoxic treatments have previously been assigned to their direct effects on tumor cells, recent findings have shown that the host's immune system also contributes to the success of chemotherapeutic and radiotherapeutic regimens. The finding that some cytotoxic antitumor compounds such as anthracyclines were capable of triggering a potent T-cell-dependent antitumor response has prompted the search for molecular determinants responsible for the immunogenicity of anthracyclines. Proteomic analyses of anthracycline-treated tumor cells have recently revealed the critical involvement of calreticulin in mediating the immunogenicity of dying tumor cells. Here, we focused on the molecular study of immunogenic chemotherapy which led to the characterization of calreticulin as a critical protein in immunogenic cancer cell death.*

Genetic mutations that lead to altered protein signaling pathways within cells may trigger their transformation, eventually leading to tumor formation. However, immunosurveillance of tumor cells might allow, in some cases, the detection and the destruction of these pre-malignant cells by the immune system before clinical detection. Factors involved in the early elimination of tumor cells include innate effectors and innate molecules, such as gamma delta T-cells (1),

natural killer cells (NK) (2), dendritic cells (DCs) (3) and natural killer T-cells (NKT) (4), TRAIL (5), perforin/ granzyme (6) as well as members of adaptive immune responses (B- and T-cells) (7). It has been shown that mice lacking recombination-activating gene 2 (RAG2) and the signaling pathway of IFN γ R or STAT-1 are more likely to develop spontaneous and chemically induced tumors, suggesting the critical involvement of the immune system in antitumor responses, a concept called "tumor immunoeediting" (7). Nonetheless, tumors evoke several mechanisms to escape immunosurveillance. Immunosubversion by tumors of the immune system could be considered as the seventh hallmark of cancer (8).

One of the goals of cytotoxic treatments could be to induce a potent antitumor immunity to apoptotic or dying tumors and restore an effective immune memory to tumor antigens. Radiotherapeutic and chemotherapeutic regimens both have the ability to induce potent tumor cell death. Although the efficacy of these cytotoxic treatments on tumor growth is generally attributed to their direct killing efficacy, the release of cellular components from dying cells such as "damage-associated molecular patterns" (9) has previously been shown to enhance immune responses (10-12). However, to elicit an efficient cytotoxic T-lymphocyte (CTL) response, several immunological steps are required to trigger links between innate and cognate immunity (13). First, the phagocytosis/capture of exogenous antigens by dendritic cells is critical in enabling the second phase *i.e.* antigen processing/presentation of the phagocytic cargo in late endosomes by DC. DCs then need to receive an "activation stimulus" which will promote their migratory capacity from tumor beds to draining lymph nodes and their maturation to the state of antigen-presenting cells (14). Cross-presentation of antigenic peptides on DC major histocompatibility (MHC) class I molecules to antigen

Correspondence to: Laurence Zitvogel, U805 and CIC BT507 Institut National de la Santé et de la Recherche Médicale, Institut Gustave Roussy, 39 rue Camille Desmoulins, F-94805 Villejuif, France. Tel: +33 1 42 11 50 41, Fax: +33 1 42 11 60 94, e-mail: zitvogel@igr.fr

Key Words: Calreticulin, proteomics, tumor cell, antitumor vaccination, chemotherapy, review.

specific T-cells will promote T-cell activation (signal 1). The recognition of co-stimulatory molecules (signal 2) and the production of cytokines (signal 3) leads to the polarization and full activation of T-cells by DCs (15, 16). Finally, DC imprinting of T-cell chemokine receptor expression (signal 4) (17, 18) associated with inflammatory processes located at the effector phase (tumor beds) will induce T-cell migratory patterns leading to tumor eradication (19, 20).

The pioneering work of Casares *et al.* (21) first showed that anthracyclines, a subclass of chemotherapeutic cytotoxic agents, can induce an immune response depending on caspases controlling antigen uptake and not DC maturation.

The exact molecular mechanisms underlying the immunogenicity were recently unraveled by Obeid *et al.* (22) who identified the role of surface calreticulin (ecto-calreticulin) in the immunogenicity of anthracycline-treated tumor cells using proteomic analyses. This finding delineates novel approaches for the generation of cancer vaccines and therapeutic intervention.

Immunogenic Chemotherapy

In addition to its direct cytotoxic antitumor effect, chemotherapy would ideally be able to harness the host's immune system to combat the tumor. However, the type of cell death mainly induced by chemotherapy is apoptosis, which is often defined as a non-immunogenic or even tolerogenic cell death modality (12). By contrast, some studies have suggested that rather than the type of cell death, the tumor cells themselves or the nature of the death-inducing stimulus would influence the immunogenicity of dying cells (23, 24). The latter hypothesis drove Casares *et al.* (21) to discover that drugs inducing endoplasmic reticulum (thapsigargin, tunicamycin, brefeldin), lysosome (bafilomycin A1) or mitochondrial stress (arsenite, betulinic acid, C2 ceramide), or drugs inhibiting the proteasome (MG132, lactacystin, ALLN), NF-KB (Bay 11-7082), or components causing DNA damage (Hoechst 33342, camptothecin, etoposide, mitomycin C) did not promote antitumor immunity. Indeed, a mouse model of vaccination was set up to assess the immunogenicity of dying tumor cells (Figure 1A). In this model, BALB/c mice were injected in one flank with CT26 colon adenocarcinoma tumor cells induced to die with various pharmacological drugs and were then rechallenged one week later with live syngeneic tumor cells. A screening of components commonly used in the clinic of the oncological armamentarium was performed and X-ray- or anthracycline-treated tumor cells were found to selectively elicit a protection against a tumor challenge. In contrast, mitomycin C- or etoposide-treated cells failed to do so, although both treatments exhibited comparable direct cytotoxic activity. This antitumor response could be attributed to T-cells, as the effects of vaccination with anthracycline- or X-ray-treated tumor cells were completely lost in athymic

nude mice lacking T-lymphocytes. Interestingly, antitumor responses in immunocompetent mice were long lasting as mice had mounted a memory response against CT26 tumor cells. Anthracycline treatment appeared to be an immunogenic chemotherapy, *i.e.* a cytotoxic therapy triggering the activation of the host immune system. These immune responses elicited by anthracyclines treated cells could be blunted in the presence of a pan caspase inhibitor Z-VAD-fmk, underlining an important role of caspases in this process. As CD11c+ DC are pivotal in the activation of T-cells, their implication in the immunogenicity of anthracycline treated tumor cells was investigated. Taking advantage of a diphtheria toxin receptor (DTR) transgenic mice expressing DTR in CD11c+ cells (25), it was shown that DCs were indeed required to mediate the immunological responses to dying tumor cells by mediating their phagocytosis. Strikingly, while both mitomycin C-, etoposide- or doxorubicin-treated tumor cells elicited comparable levels of DC maturation, only doxorubicin treated cells were efficiently phagocytosed by DC. This observation formally demonstrated a tight link between phagocytosis of apoptotic bodies and immunogenicity. The molecular mechanisms responsible for the phagocytosis of tumor cells nevertheless remained unexplored until the identification of calreticulin as the key protein in mediating phagocytosis and therefore the immunogenicity of dying cancer cells by Obeid *et al.* [(22), discussed below].

Background of Calreticulin

Calreticulin was first identified as a calcium-binding protein of the muscle sarcoplasmic reticulum (26). This protein was later found to be present in every cell of higher organisms. Under classical conditions, calreticulin is found in the lumen of the reticulum participating in calcium homeostasis and acting as a protein chaperone (27, 28). However, a few reports showed that calreticulin could also be detected in many intracellular compartments and at the plasma membrane (29-31). Indeed, it was recently demonstrated by Henson *et al.* (32) that calreticulin is a marker for phagocytosis on the surface of apoptotic cells. They claimed that membrane calreticulin expression was the second signal (in addition to phosphatidylserine) for apoptotic cells to be phagocytosed (32). Calreticulin was also identified as a marker for the phagocytosis of apoptotic cells in *Drosophila* (33) and was recently shown to be required for the adiponectin-mediated clearance of early apoptotic bodies (34).

Identification of Calreticulin as a Protein Involved in Immunogenic Cell Death

As previous data obtained from Casares *et al.* (21) pointed to a major role of phagocytosis of apoptotic cells in mediating an immune response, the assumption that some

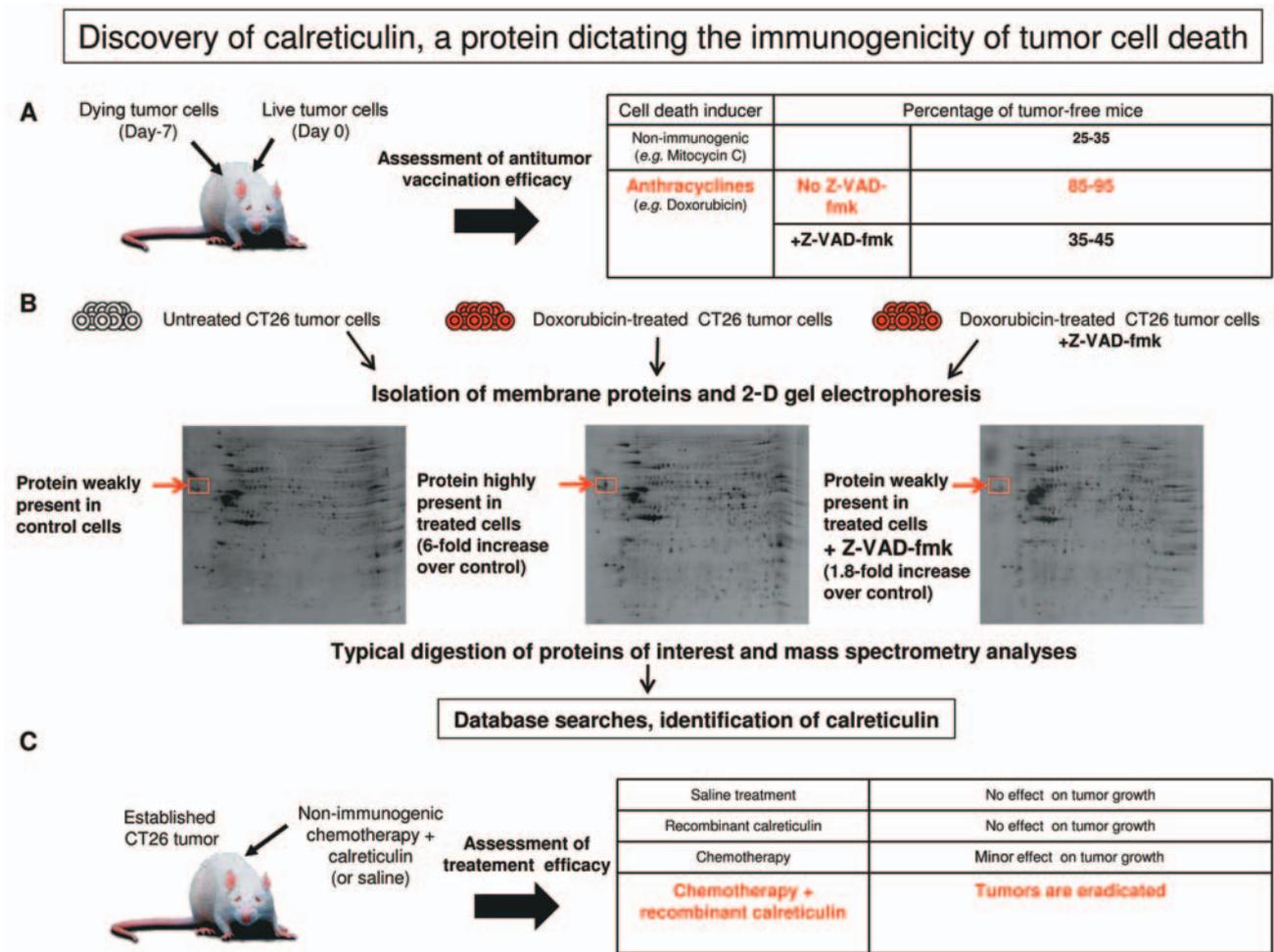


Figure 1. The discovery of the concept of immunogenicity of cell death. The sequential steps which led Obeid *et al.* (22) to the discovery of ectocalreticulin as a predictive factor for the immunogenicity of chemotherapy are detailed. The proteomic analyses of anthracycline-treated CT26 tumors in the presence of Z-VAD-fmk versus live CT26 are shown. The synergistic antitumor effects between non-immunogenic cell death inducers and recombinant calreticulin or inhibitors of PP1/GADD34 are depicted.

induced (or repressed) membrane proteins might account for the immunogenicity of doxorubicin treatment was made. As phagocytosis was shown to be a rapid process occurring within one hour following anthracycline exposure, the differential immunogenicity obtained with various regimens was unlikely to result from transcription regulatory processes. Thus, rather than examining the transcriptome expression induced by treatment, Obeid *et al.* (22) focused on the potential proteomic changes of the external membranes of tumor cells. A proteomic comparison of membranes coming from doxorubicin-treated cells and control cells led to the discovery of calreticulin as shown in Figure 1B. CT26 cells were either left untreated or treated with doxorubicin or treated with doxorubicin and Z-VAD-fmk, a pan caspase inhibitor that reduces the uptake and immunogenicity of doxorubicin-elicited cell death (21).

Membrane proteins were then biotinylated, affinity purified and finally subjected to 2-dimensional electrophoresis. Calreticulin was eventually identified as a protein that was dramatically upregulated by CT26 cells upon treatment with doxorubicin (by a factor of 6), but less (by a factor of 1.8) upon treatment with doxorubicin and Z-VAD-fmk. Another protein identified was ERP57, a calreticulin-interacting chaperone (30).

Use of Calreticulin in Anticancer Therapy

Although proteomic studies of tumor cells provided putative interesting candidates for the improvement of the current management of cancer patients, the functional validation of these candidates is unfortunately missing from most studies. However, Obeid *et al.* (22) provided extensive data to confirm

and extend their discovery of calreticulin as a mediator of immunogenic cell death. Using fluorescence-activated cell sorter and immunofluorescence analyses, they have indeed shown that surface calreticulin (ecto-calreticulin) expression on tumor cells upon treatment with cytotoxic agents was strongly correlated with the ability of dying tumor cells to elicit tumor antigen uptake by DC and subsequent antitumor immunity. By preventing calreticulin expression using a specific small interference RNA, they showed that ecto-calreticulin is responsible for triggering an immune response to idarubicin/doxorubicin/mitoxantrone-treated tumor cells and by adding back recombinant calreticulin to mitomycin C-etoposide-treated tumor cells, they restored antitumor immune responses. Obeid *et al.* (22) finally unraveled the molecular pathway dictating ecto-calreticulin exposure to cell surface. They identified that anthracyclines induced the phosphorylation of eIF2 α , a protein which is hyperphosphorylated in ER stress. Interestingly, the use of compounds which inhibit the PP1/GADD34 complex (involved in eIF2 α dephosphorylation) was sufficient to induce calreticulin exposure. Furthermore, when these inhibitors were injected into CT26 tumors along with a non-immunogenic cell death inducer (etoposide or mitomycin C), synergistic antitumor effects were observed (Figure 1C). These latter results delineate an immunogenic chemotherapy strategy to combat cancer.

Importance of Proteomic Studies in Immunogenic Chemotherapy

Proteomic analyses have traditionally been used to study correlations between proteins expressed in healthy *versus* tumor tissues. These investigations have revealed cellular processes and biochemical networks at the protein level that are responsible for the development of a particular disease. At the clinical level, the characterization of proteins directly involved in the disease process can lead to the design of drugs capable of targeting selective oncogenic pathways in tumor cells. One example of a paradigmatic drug designed to specifically interfere with c-kit/bcr-abl or PDGFR α tyrosine kinases is imatinib mesylate (35-37). The ability of proteomics to detect variations of cell protein expression therefore allows the use of this method to study the molecular events occurring during disease processes (38), identify biomarkers of disease (39, 40), or explore the mechanisms of action of anticancer drugs at the molecular level (41, 42). Nevertheless, because of variations in protein expression, post-translational modifications and compartmentalization, results from proteomic studies should be analysed cautiously in the context of the biological processes involved. Results obtained from whole tumor cell extracts may lead to misinterpretation of results as they give an "average picture" of proteome variations within a cell, while the study of subcellular fractions

would have provided a more accurate picture of cellular alterations within tumor cells upon treatment (43, 44).

The recent paper of Obeid *et al.* (22) is in this regard very interesting as the background information on the importance of phagocytosis in mediating immunogenic chemotherapy has led to the study of membrane fractions rather than whole tumor cell lysates. This strategy allowed the identification of the membrane translocation of calreticulin upon immunogenic cytotoxic treatment of tumor cells. This finding could have clinical implications if the authors could show that calreticulin exposure on tumor cells following patient therapy with defined/selected chemotherapeutic regimen is correlated with long term disease-free survival and immune responses. Calreticulin exposure could easily and routinely be monitored on plasma membranes of tumor cells and could become a predictive factor of an immunogenic chemotherapy (45). The identification of immunogenic proteins expressed by tumor cells could be regarded as a new application of proteomics. Besides its potential use as a predictive factor, ecto-calreticulin could be systematically induced by inhibitors of PP1/GADD34 during non-immunogenic chemotherapies. Moreover, proteomic technology could be extremely useful in determining other proteins within tumor cells that account for immunogenic chemotherapy. For instance, while calreticulin exposure on tumors is now characterized as the first checkpoint to elicit an immune response, the absence of immune response against live cells that express ecto-calreticulin prompts the search for additional proteins responsible for the activity of dying tumor cells. The knowledge of the different actors involved in the immunogenicity of dying tumor cells could aid in the design of future anticancer therapies.

Conclusion

The initial demonstration of immunogenic chemotherapy regimens has challenged the common view that chemotherapy mediates its effects only through cell-autonomous effects. The work of Obeid *et al.* (22) illustrates a utilization of proteomic tools for a new objective: the identification of proteins which are critically involved in the immunogenicity of dying tumor cells. Not only does this work confirm the previous studies of Casares *et al.* (21), but it also provides clinical relevance to these findings by showing that non-immunogenic treatments could be rendered immunogenic using compounds leading to overexpression of calreticulin by tumor cells. These data have indeed arisen great interest among scientists as chemoresistance of tumors could be reversed by the modulation of the PP1/GADD34 complex. In addition, calreticulin could be used to predict the lack of efficacy of a chemotherapeutic regimen *in vitro* or *in vivo*. We further plan (i) to investigate the effects of

calreticulin in humans and (ii) to study the efficacy in humans of inhibitors of PP1/GADD34 as potential enhancers of antitumor responses.

References

- Gao Y, Yang W, Pan M, Scully E, Girardi M, Augenlicht LH, Craft J and Yin Z: Gamma delta T cells provide an early source of interferon gamma in tumor immunity. *J Exp Med* 198: 433-442, 2003.
- Borg C, Terme M, Taieb J, Menard C, Flament C, Robert C, Maruyama K, Wakasugi H, Angevin E, Thielemans K, Le Cesne A, Chung-Scott V, Lazar V, Tchou I, Crepineau F, Lemoine F, Bernard J, Fletcher JA, Turhan A, Blay JY, Spatz A, Emile JF, Heinrich MC, Mecheri S, Tursz T and Zitvogel L: Novel mode of action of c-kit tyrosine kinase inhibitors leading to NK cell-dependent antitumor effects. *J Clin Invest* 114: 379-388, 2004.
- Taieb J, Chaput N, Menard C, Apetoh L, Ullrich E, Bonmort M, Pequignot M, Casares N, Terme M, Flament C, Opolon P, Lecluse Y, Metivier D, Tomasello E, Vivier E, Ghiringhelli F, Martin F, Klatzmann D, Poynard T, Tursz T, Raposo G, Yagita H, Ryffel B, Kroemer G and Zitvogel L: A novel dendritic cell subset involved in tumor immunosurveillance. *Nat Med* 12: 214-219, 2006.
- Crowe NY, Smyth MJ and Godfrey DI: A critical role for natural killer T cells in immunosurveillance of methylcholanthrene-induced sarcomas. *J Exp Med* 196: 119-127, 2002.
- Wiley SR, Schooley K, Smolak PJ, Din WS, Huang CP, Nicholl JK, Sutherland GR, Smith TD, Rauch C, Smith CA *et al*: Identification and characterization of a new member of the TNF family that induces apoptosis. *Immunity* 3: 673-682, 1995.
- Trapani JA and Smyth MJ: Functional significance of the perforin/granzyme cell death pathway. *Nat Rev Immunol* 2: 735-747, 2002.
- Shankaran V, Ikeda H, Bruce AT, White JM, Swanson PE, Old LJ and Schreiber RD: IFN γ and lymphocytes prevent primary tumour development and shape tumour immunogenicity. *Nature* 410: 1107-1111, 2001.
- Zitvogel L, Tesniere A and Kroemer G: Cancer despite immunosurveillance: immunoselection and immunosubversion. *Nat Rev Immunol* 6: 715-727, 2006.
- Matzinger P: Tolerance, danger, and the extended family. *Annu Rev Immunol* 12: 991-1045, 1994.
- Basu S, Binder RJ, Suto R, Anderson KM and Srivastava PK: Necrotic but not apoptotic cell death releases heat shock proteins, which deliver a partial maturation signal to dendritic cells and activate the NF-kappa B pathway. *Int Immunol* 12: 1539-1546, 2000.
- Todryk S, Melcher AA, Hardwick N, Linardakis E, Bateman A, Colombo MP, Stoppacciaro A and Vile RG: Heat shock protein 70 induced during tumor cell killing induces Th1 cytokines and targets immature dendritic cell precursors to enhance antigen uptake. *J Immunol* 163: 1398-1408, 1999.
- Zitvogel L, Casares N, Pequignot MO, Chaput N, Albert ML and Kroemer G: Immune response against dying tumor cells. *Adv Immunol* 84: 131-179, 2004.
- Ghiringhelli F, Apetoh L, Housseau F, Kroemer G and Zitvogel L: Links between innate and cognate tumor immunity. *Curr Opin Immunol* 19: 224-231, 2007.
- Banchereau J and Steinman RM: Dendritic cells and the control of immunity. *Nature* 392: 245-252, 1998.
- Albert ML, Sauter B and Bhardwaj N: Dendritic cells acquire antigen from apoptotic cells and induce class I-restricted CTLs. *Nature* 392: 86-89, 1998.
- Ronchetti A, Iezzi G, Crosti MC, Garancini MP, Protti MP and Bellone M: Role of antigen-presenting cells in cross-priming of cytotoxic T lymphocytes by apoptotic cells. *J Leukoc Biol* 66: 247-251, 1999.
- Hwu P, Bedikian AY and Grimm EA: Challenges of chemosensitivity testing. *Clin Cancer Res* 12: 5258-5259, 2006.
- Mora JR, Bono MR, Manjunath N, Weninger W, Cavanagh LL, Roseblatt M and Von Andrian UH: Selective imprinting of gut-homing T cells by Peyer's patch dendritic cells. *Nature* 424: 88-93, 2003.
- Lang KS, Georgiev P, Recher M, Navarini AA, Bergthaler A, Heikenwalder M, Harris NL, Junt T, Odermatt B, Clavien PA, Pircher H, Akira S, Hengartner H and Zinkernagel RM: Immunoprivileged status of the liver is controlled by Toll-like receptor 3 signaling. *J Clin Invest* 116: 2456-2463, 2006.
- Colonna M: Toll-like receptors and IFN-alpha: partners in autoimmunity. *J Clin Invest* 116: 2319-2322, 2006.
- Casares N, Pequignot MO, Tesniere A, Ghiringhelli F, Roux S, Chaput N, Schmitt E, Hamai A, Hervas-Stubb S, Obeid M, Coutant F, Metivier D, Pichard E, Aucouturier P, Pierron G, Garrido C, Zitvogel L and Kroemer G: Caspase-dependent immunogenicity of doxorubicin-induced tumor cell death. *J Exp Med* 202: 1691-1701, 2005.
- Obeid M, Tesniere A, Ghiringhelli F, Fimia GM, Apetoh L, Perfettini JL, Castedo M, Mignot G, Panaretakis T, Casares N, Metivier D, Larochette N, van Endert P, Ciccocanti F, Piacentini M, Zitvogel L and Kroemer G: Calreticulin exposure dictates the immunogenicity of cancer cell death. *Nat Med* 13(1): 54-61, 2007.
- Scheffer SR, Nave H, Korangy F, Schlote K, Pabst R, Jaffee EM, Manns MP and Gretten TF: Apoptotic, but not necrotic, tumor cell vaccines induce a potent immune response *in vivo*. *Int J Cancer* 103: 205-211, 2003.
- Bartholomae WC, Rininsland FH, Eisenberg JC, Boehm BO, Lehmann PV and Tary-Lehmann M: T cell immunity induced by live, necrotic, and apoptotic tumor cells. *J Immunol* 173: 1012-1022, 2004.
- Jung S, Unutmaz D, Wong P, Sano G, De los Santos K, Sparwasser T, Wu S, Vuthoori S, Ko K, Zavala F, Pamer EG, Littman DR and Lang RA: *In vivo* depletion of CD11c(+) dendritic cells abrogates priming of CD8(+) T cells by exogenous cell-associated antigens. *Immunity* 17: 211-220, 2002.
- Ostwald TJ and MacLennan DH: Isolation of a high affinity calcium-binding protein from sarcoplasmic reticulum. *J Biol Chem* 249: 974-979, 1974.
- Krause KH and Michalak M: Calreticulin. *Cell* 88: 439-443, 1997.
- Sadasivan B, Lehner PJ, Ortman B, Spies T and Cresswell P: Roles for calreticulin and a novel glycoprotein, tapasin, in the interaction of MHC class I molecules with TAP. *Immunity* 5: 103-114, 1996.
- Michalak M, Corbett EF, Mesaeli N, Nakamura K and Opas M: Calreticulin: one protein, one gene, many functions. *Biochem J* 344(Pt 2): 281-292, 1999.
- Bedard K, Szabo E, Michalak M and Opas M: Cellular functions of endoplasmic reticulum chaperones calreticulin, calnexin, and ERp57. *Int Rev Cytol* 245: 91-121, 2005.

- 31 Johnson S, Michalak M, Opas M and Eggleton P: The ins and outs of calreticulin: from the ER lumen to the extracellular space. *Trends Cell Biol* 11: 122-129, 2001.
- 32 Gardai SJ, McPhillips KA, Frasch SC, Janssen WJ, Starefeldt A, Murphy-Ullrich JE, Bratton DL, Oldenborg PA, Michalak M and Henson PM: Cell-surface calreticulin initiates clearance of viable or apoptotic cells through trans-activation of LRP on the phagocyte. *Cell* 123: 321-334, 2005.
- 33 Kuraishi T, Manaka J, Kono M, Ishii H, Yamamoto N, Koizumi K, Shiratsuchi A, Lee BL, Higashida H and Nakanishi Y: Identification of calreticulin as a marker for phagocytosis of apoptotic cells in *Drosophila*. *Exp Cell Res*, 2007, in press.
- 34 Takemura Y, Ouchi N, Shibata R, Aprahamian T, Kirber MT, Summer RS, Kihara S and Walsh K: Adiponectin modulates inflammatory reactions *via* calreticulin receptor-dependent clearance of early apoptotic bodies. *J Clin Invest*, 2007, in press.
- 35 Heinrich MC, Blanke CD, Druker BJ and Corless CL: Inhibition of KIT tyrosine kinase activity: a novel molecular approach to the treatment of KIT-positive malignancies. *J Clin Oncol* 20: 1692-1703, 2002.
- 36 Buchdunger E, O'Reilly T and Wood J: Pharmacology of imatinib (STI571). *Eur J Cancer* 38(Suppl 5): S28-36, 2002.
- 37 Ben-Neriah Y, Daley GQ, Mes-Masson AM, Witte ON and Baltimore D: The chronic myelogenous leukemia-specific P210 protein is the product of the bcr/abl hybrid gene. *Science* 233: 212-214, 1986.
- 38 Patterson SD and Aebersold RH: Proteomics: the first decade and beyond. *Nat Genet* 33(Suppl): 311-323, 2003.
- 39 Chignard N and Beretta L: Proteomics for hepatocellular carcinoma marker discovery. *Gastroenterology* 127: S120-125, 2004.
- 40 Van Eyk JE: Proteomics: unraveling the complexity of heart disease and striving to change cardiology. *Curr Opin Mol Ther* 3: 546-553, 2001.
- 41 Yim EK, Lee KH, Bae JS, Namkoong SE, Um SJ and Park JS: Proteomic analysis of antiproliferative effects by treatment of 5-fluorouracil in cervical cancer cells. *DNA Cell Biol* 23: 769-776, 2004.
- 42 Castagna A, Antonioli P, Astner H, Hamdan M, Righetti SC, Perego P, Zunino F and Righetti PG: A proteomic approach to cisplatin resistance in the cervix squamous cell carcinoma cell line A431. *Proteomics* 4: 3246-3267, 2004.
- 43 Poirier F, Pontet M, Labas V, le Caer JP, Sghiouar-Imam N, Raphael M, Caron M and Joubert-Caron R: Two-dimensional database of a Burkitt lymphoma cell line (DG 75) proteins: protein pattern changes following treatment with 5'-azacytidine. *Electrophoresis* 22: 1867-1877, 2001.
- 44 Poirier F, Joubert-Caron R, Labas V and Caron M: Proteomic analysis of a lymphoma-derived cell line (DG75) following treatment with a demethylating drug: modification of membrane-associated proteins. *Proteomics* 3: 1028-1036, 2003.
- 45 Lake RA and van der Most RG: A better way for a cancer cell to die. *N Engl J Med* 354: 2503-2504, 2006.

Received February 1, 2007

Accepted March 15, 2007