Genetic Variations in the 3′-untranslated Regions of Genes Involved in the Cell Cycle and Apoptosis Pathways Affect Bladder Cancer Risk

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\textbf{Abstract.} Background: Key genes related to cell cycle and apoptosis pathways play critical roles in bladder cancer. Single nucleotide polymorphisms (SNPs) in the 3′-untranslated regions (3′-UTR) of genes may impact microRNA (miRNA)-messenger RNA (mRNA) binding capacity and alter gene expression to contribute to the susceptibility of cancers. However, an association of genetic variations in cell cycle and apoptosis pathways with bladder cancer risk, has not been reported. \textbf{Materials and Methods:} We selected SNPs in the 3′-UTR of cell cycle and apoptosis pathways genes and genotyped them with a case-control study consisting of 578 bladder cancer patients and 1,006 cancer-free subjects. Dual luciferase reporter gene assay was performed to validate the biological function of important SNPs. Results: We found that 5 SNPs might change the binding ability of miRNA to their target genes, among which PPP3CC rs7431 A>G located in the 3′-untranslated regions with the minimum p-value (p=5.75×10^{-4}). Analysis revealed that the rs7431 disrupted miR-212 and miR-132 targeting sites. Logistic regression revealed a significantly decreased risk of bladder cancer associated with the PPP3CC rs7431 A>G polymorphism with an odds ratio (OR) of 0.76 (95% confidence interval (CI)=0.66-0.89, p=5.75×10^{-4}). Luciferase report assay showed that both miR-212 and miR-132 targeting sites, Logistic regression revealed a significantly decreased risk of bladder cancer associated with the PPP3CC rs7431 A>G polymorphism with an odds ratio (OR) of 0.76 (95% confidence interval (CI)=0.66-0.89, p=5.75×10^{-4}). Analysis revealed that the rs7431 disrupted miR-212 and miR-132 targeting sites. Logistic regression revealed a significantly decreased risk of bladder cancer associated with the PPP3CC rs7431 A>G polymorphism with an odds ratio (OR) of 0.76 (95% confidence interval (CI)=0.66-0.89, p=5.75×10^{-4}).

Genetic Variations in the 3′-untranslated Regions of Genes Involved in the Cell Cycle and Apoptosis Pathways Affect Bladder Cancer Risk

Bladder cancer is one of the most frequent malignancies of the urinary tract worldwide (1). In China, the incidence rates of bladder cancer have increased gradually from 2000 to 2011 (2). The overall incidence was 7.68 per 100,000 and mortality rates were 1.99 per 100,000 in the population according to the National Central Cancer Registry (NCCR).
of China 2015 annual report (3). Despite the relatively low incidence of bladder carcinoma compared to other cancer types, the high local recurrence rate requires regular medical examination associated with a significant impact on healthcare resources and the patient’s quality of life (4). Evidence showed that bladder cancer is a complex disease caused by both genetic and environmental risk factors, including cigarette smoking, occupational exposure to carcinogens (5). Even following the same exposure to environmental carcinogenic factors, only a small number of the exposed individuals eventually occur cancers, suggesting that genetic susceptibility in critical genes may play an important role in the etiology of bladder cancer.

Cell-cycle and apoptosis pathways are related to the occurrence and development of tumors (6). Cell cycle is the series of events that happen in a cell contributing to its division and duplication of its DNA to produce two daughter cells (7), and cell apoptosis is a process of programmed cell death that occurs in multicellular organisms (8). Both of the pathways play critical roles in regulating cellular transformation and malignancy. Defective processes of cell cycle and apoptosis affected by key genes have been implicated in bladder cancer (6).

The regulation of cell cycle and apoptosis can be controlled by epigenetics. As an important aspect of epigenetics, microRNAs (miRNAs) are a kind of small noncoding and single-stranded RNA molecules (~22 nt), which normally regulate gene expression through binding the 3'-untranslated regions (3'-UTR) of messenger RNA (mRNA) of target gene at the post-transcriptional level (9). It has been suggested that miRNAs play critical roles in regulating a variety of biological processes, such as organ development, tumorigenesis, cell differentiation, cell proliferation and apoptosis (10). Several studies showed that miRNAs could act as oncogenes and tumor suppressors to influence the occurrence and development of tumor by targeting 3'-UTR of important genes (11, 12).

Single nucleotide polymorphisms (SNPs) are one of the most frequent variations in the human genome. Accumulating evidence have demonstrated that an increasing number of 3'-UTR SNPs located in miRNA binding sites impact miRNA binding ability and affect tumor development (13-15). Therefore, genetic variants in the miRNAs binding sites of key target genes involved in cell cycle and apoptosis pathways may impact miRNA-mRNA binding and alter gene expression. However, the function between genetic variants in miRNA binding sites and bladder cancer risk is still unknown.

Cell-cycle and apoptosis are regulated by epigenetic miRNA and genetic factors that occur in gene 3'-UTR SNPs. We investigated the association between genetic variations of key genes involved in cell cycle and apoptosis pathways and bladder cancer risk with a case-control study including 578 cases and 1,006 controls in the a Chinese population.

Materials and Methods

Study population. This study was approved by the Institutional Review Board of Nanjing Medical University (Nanjing, China). The recruitment of all individuals was described previously (16). Briefly, 578 cases were collected from Nanjing beginning in May 2003, and 1,006 controls shared with ChinaPCa (17). The bladder cancer patients were histologically diagnosed by at least two pathologists, and detailed clinical data were collected by local urologists. Pathological grade and stage were consistent with the 2004 WHO/ISUP classification of bladder cancer. Individuals who smoked daily for >1 year were considered smokers, and the others were considered non-smokers. All participants were provided written informed consent form for their samples and data to be used.

SNPs selection and genotyping. The candidate genes involved in the cell cycle and apoptosis pathways were found using KEGG pathway maps (www.kegg.jp/kegg/pathway.html), and all the SNPs in the 3'-UTR of these genes were selected based on 1000 Genomes Project data. Subsequently, SNPs were filtered if they (a) exhibited genotype call rate of <90%; (b) showed a minor allele frequency (MAF) of ≤0.05 in either cases or controls; or (c) deviated from Hardy-Weinberg equilibrium (HWE) in the controls at p<0.001.

The genomic DNA was extracted from peripheral blood lymphocytes. The selected SNPs were genotyped in 578 cases and 1,006 controls using the Illumina Infinium® BeadChip platform (Illumina, San Diego, CA, USA). Genotype calling was conducted by the GenTrain version 1.0 clustering algorithm in GenomeStudio v2011.1 (Illumina, San Diego, CA, USA).

Bioinformatics analysis of miRNA-mRNA binding ability. The online software miRNASNP (http://www.bioguo.org/miRNASNP/) was used to predict whether 3'-UTR SNPs would change the miRNAs binding ability to the target genes.

Construction of reporter plasmids and luciferase reporter assay. To construct luciferase reporter plasmids, PPP3CC 3'-UTR fragments (A allele or G allele for SNP rs7431) were gene synthesized and inserted at the Xhol/NotI site, downstream of the Renilla luciferase gene in the psiCHECK™-2 vector (Promega, Madison, WI, USA) by Generay Company (Shanghai, China). The nucleotide sequences of the constructed plasmids were confirmed by DNA sequencing method.

In this study, we chose T24 and J82 cell lines for the further luciferase assay. Both cell lines were grown at 37°C in the presence of 5% CO2 in a humidified incubator. T24 and J82 cells were seeded on 24-well plates and then each transient co-transfected reaction contained 0.8 μg constructed vectors, either with A allele or with G allele and 20 pmol chemically synthesized miR-212 or miR-132 or negative control miRNA with Lipofectamine 2000 (Invitrogen, Carlsbad, CA, USA), respectively. Luciferase activity was determined with a Dual-Luciferase Reporter Assay System (Promega, Madison, WI, USA) after 48 h. Relative protein levels were expressed as Renilla luciferase normalized against Firefly luciferase signals. Independent triplicate experiments were performed for each plasmid construct.

Statistical analysis. HWE of the controls’ genotype frequencies was tested using a goodness-of-fit χ² test. Associations between each SNP and risk of bladder cancer were estimated with odds ratios
(ORs) and 95% confidence intervals (CIs) derived from logistic regression analysis under an additive model. All statistical tests were two-sided and a $p$-value $<0.05$ was used as the criterion of statistical significance. The Student’s $t$-test was used to analyze the results of miRNA or mRNA expression in bladder cancer and normal tissues. The miRNA expression levels in bladder cancer tissues and normal tissues were downloaded from the publicly available Gene Expression Omnibus (GEO) database (GSE68594). Statistical analyses were conducted with Statistical Analysis System software (v.9.1 SAS Institute Inc., Cary, NC, USA).

**Results**

**Association between 15 selected SNPs with bladder cancer risk.** There were 230 genes involved in the cell cycle and apoptosis pathways. We obtained 2,562 SNPs in the 3'-UTR of these candidate genes based on the 1000 Genomes Project and then 597 SNPs were selected after quality control. We detected these 597 SNPs genotype distribution in a case-control study and the distribution of the selected characteristics between case and control subjects is summarized in Table I. Finally, we chose 15 significantly different ($p<0.05$) candidate SNPs and the genotype distributions were shown in Table II. The genotype frequencies of these 15 SNPs in controls were in agreement with HWE ($p>0.001$). Among them, we found that the SNP rs7431 A>G with the minimum $p$-value performed a significantly decreased risk of bladder cancer with an OR of 0.76 (95%CI=0.66-0.89, $p=5.75\times10^{-4}$).

**PPP3CC 3'-UTR rs7431 A>G affects PPP3CC expression by regulating the hsa-miR-212 and hsa-miR-132 binding.** We used bioinformatics analysis to determine whether the genetic variants would change the binding ability of miRNA. As a result, we found 5 SNPs might change the binding ability to 3'-UTR of the target genes among 15 candidate SNPs (Table II), and rs7431 had both the minimum $p$-value and the biological effect. To explore whether the SNP rs7431 may influence PPP3CC regulation mediated by miR-212 and miR-132, we constructed luciferase reporter gene assay by using psiCHECK™-2 vector (Figure 1a). Based on the bioinformatics analysis, the PPP3CC rs7431 was located on the target site of miR-212 and miR-132 (Figure 1b). As shown in Figure 1c, both of miR-212 and miR-132 can significantly reduce luciferase activities in T24 cells transfected with rs7431A/G allelic reporter constructs compared to negative control miRNAs. Moreover, vectors within the rs7431 G allele had an increased luciferase activity in adding miR-212 ($p<0.001$) and miR-132 ($p=0.001$) respectively compared with those within the rs7431 A allele in T24 cells. The similar result was also observed in J82 cells ($p=0.001$ and $p=0.002$, respectively). Our findings suggested that both miR-212 and miR-132 could bind and positively regulate the transcription of *PPP3CC* which was more positively influenced by the variant G allele *in vitro*.

From the independent TCGA data, we found the mRNA expression levels of *PPP3CC* significantly decreased in 19 tumor tissues compared with their adjacent normal tissues ($p<0.001$) (Figure 2a). On the contrary, the expression of miR-212/miR-132 significantly increased in tumor tissues compared with normal tissues ($p<0.001$; Figure 2b and c) by GSE68594.

**Discussion**

In the present study, we conducted a case-control study to investigate the association between SNPs in the 3'-UTR of genes involved in the cell cycle and apoptosis pathways and bladder cancer risk. We identified that rs7431 on the miR-212 and miR-132 binding site in *PPP3CC* 3'-UTR was associated with the decreased risk of bladder cancer. The Dual luciferase reporter gene assay indicated that *PPP3CC* rs7431 A to G substitution may decrease the binding ability of miR-212 and miR-132 with *PPP3CC* 3'-UTR. This is the first study to evaluate the possible association of the cell cycle and apoptosis associated genes 3'-UTR SNPs with bladder cancer.

Apoptosis was an initiative cell death process that is controlled by genes (18). It was also a therapeutic target of cancer (19). Genetic variations in apoptosis-regulating genes may play key roles in alterations in cellular apoptotic

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**Table I. Characteristics of selected subjects in bladder cancer cases and controls.**

<table>
<thead>
<tr>
<th>Variables</th>
<th>Cases</th>
<th>Controls</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age, years$^1$</td>
<td>n=578 (%)</td>
<td>n=1006 (%)</td>
</tr>
<tr>
<td>Mean (SD)</td>
<td>64.7 (12.4)</td>
<td>61.5 (9.5)</td>
</tr>
<tr>
<td>Median (range)</td>
<td>66.0 (28.0-90.0)</td>
<td>62.0 (41.0-79.0)</td>
</tr>
<tr>
<td>Gender</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Male</td>
<td>478 (82.7)</td>
<td>1006 (100.0)</td>
</tr>
<tr>
<td>Female</td>
<td>100 (17.3)</td>
<td>0 (0.0)</td>
</tr>
<tr>
<td>Smoking status</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Never</td>
<td>311 (53.8)</td>
<td>454 (45.1)</td>
</tr>
<tr>
<td>Ever</td>
<td>264 (45.7)</td>
<td>506 (50.3)</td>
</tr>
<tr>
<td>Missing</td>
<td>3 (0.5)</td>
<td>46 (4.6)</td>
</tr>
<tr>
<td>Grade$^2$</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Low grade</td>
<td>287 (49.7)</td>
<td>--</td>
</tr>
<tr>
<td>High grade</td>
<td>174 (30.1)</td>
<td>--</td>
</tr>
<tr>
<td>Missing</td>
<td>117 (20.2)</td>
<td>--</td>
</tr>
<tr>
<td>Stage$^2$</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Non-muscle invasive (Ta-T1)</td>
<td>375 (64.9)</td>
<td>--</td>
</tr>
<tr>
<td>Invasive (T2-T4)</td>
<td>152 (26.3)</td>
<td>--</td>
</tr>
<tr>
<td>Missing</td>
<td>51 (8.8)</td>
<td>--</td>
</tr>
</tbody>
</table>

$^1$Age at diagnosis. $^2$Grade and stage in accordance with the 2004 WHO/ISUP classification of bladder cancer.
several genes were related to diseases by affecting miRNA-
exact mechanism about the
PPP3C C
with the
additive model.
the cancer recurrence (22). In our study, we found a
regulated gene expression (15, 23, 24). For example, Song
PPP3C C
chromosome 8. Previous studies have shown that low
expression of
precise mechanism of apoptosis was not fully understood.
involved in the apoptosis process and it was localized on
the process of apoptosis may have a direct or indirect
relationship with the occurrence of a tumor. However, the
apo...
Figure 1. Characterization and functional analyses of the 3′-UTR of PPP3CC. (A) Schematic representation of reporter plasmids containing the PPP3CC 3′-UTR, which was inserted downstream of Renilla luciferase gene in the psiCHECK™-2 vector. (B) Complementarity between miR-212/miR-132 and the PPP3CC 3′-UTR site targeted. The SNP rs7431 was located within the ‘seed region’ of the miR-212/miR-132 binding site. (C) The effect of SNP rs7431 on the interaction between the PPP3CC 3′-UTR and miR-212/ miR-132 in T24 and J82 cells. The luciferase activity of each construct was normalized against the negative control miRNA (NC) transient transfected with constructed vectors with A allele.

Figure 2. Expression of PPP3CC and miR-212/ miR-132 in bladder cancer and normal tissues. (A) PPP3CC mRNA expression levels were analyzed in paired bladder tissues from 19 subjects from TCGA data. The p-values were determined using the paired t-test. The expression levels of (B) miR-212 and (C) miR-132 were evaluated using the GSE68594 database.
Conflicts of Interest

The Authors declare no conflicts of interest.

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References


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