Review

The Significance of Proteomic Biomarkers in Male Breast Cancer

ELENI ZOGRAFOS¹, MARIA GAZOULI¹, GEORGIOS TSANGARIS² and EVANGELOS MARINOS¹

¹Department of Basic Medical Sciences, Laboratory of Biology, School of Medicine, University of Athens, Athens, Greece; ²Proteomics Research Unit, Center of Basic Research II, Biomedical Research Foundation of the Academy of Athens, Athens, Greece

Abstract. Breast cancer in men (MBC) is an uncommon malignancy and accounts for only 1% of all diagnosed breast cancers. By using genomic and transcriptomic approaches, researchers have been able to expand our insight into the genetic basis of breast cancer, by providing new biomarkers. We currently know that gene analysis by itself does not show the complete picture. Along with the genomic approach, proteomics are crucial for the improvement of breast cancer diagnosis, sub-classification, for predicting response to different treatment modalities and for predicting prognosis. There are great challenges in identifying discriminatory proteins and the use of specific techniques along with additional analytical tools is required. A number of techniques allow testing for proteins produced during specific diseases. In this review, an effort is made to summarize the studies and results linked to the implementation of proteomics in the field of MBC detection and diagnosis.

Breast cancer in men (MBC) is an uncommon malignancy, it accounts for only 1% of all diagnosed breast cancers (1). According to the American Cancer Society, approximately 2,350 men will be diagnosed with breast cancer, whilst it is estimated that nearly 440 men will die from the disease in the United States in 2015. In Athens, the capital city of Greece, it has been estimated that there are approximately 20-25 new cases per year (in a male population of around 2 million) (2).

Correspondence to: Maria Gazouli, Ph.D., Assistant Professor of Molecular Biology Department of Basic Medical Sciences, School of Medicine, University of Athens, Michalakopoulou 176, 11527 Athens, Greece. Tel: +30 2107462231, +30 2107462244, e-mail: mgazouli@med.uoa.gr

Key Words: Male breast cancer, proteomics, genetics, biomarkers, review.

Lifetime risk of a male developing breast cancer remains as low as 0.01% (3). The low incidence of breast malignant neoplasms in men may be due to a variety of reasons; it is unlikely for cancer to develop in vestigial parts such as male breasts (4), whereas there is not a continuous endocrine stimulation of the male breast by ovarian hormones (estrogens) (5).

The main risk factors for the development of MBC are aging, high estrogen levels, breast cancer (BRCA) gene mutations, etc. Aging is an important risk factor; the average age of first diagnosis in men is 67 years vs. 62 years in women (6). Moreover, high levels of estrogen have been implicated as a significant risk factor; of note, the imbalance of androgenic vs. estrogenic levels may be caused by a host of factors such as Klinefelter syndrome (7), high body mass index (8), lack of exercise (9), previous liver disease (10), excessive alcohol consumption (11), diabetes (12), infertility (12) and use of exogenous androgens (13), or estrogens (14). Special attention should be paid to the important role for endogenous estradiol in the etiology of male breast cancer, similarly to breast cancer in females, as was recently published by Brinton et al. in a largescale collaborative study (15). Additionally, environmental issues such as exposure to ionizing radiation have been associated with higher risk of MBC (16-18). Finally, a genetic predisposition may also be considered in the etiology of MBC. More specifically, it has been shown that approximately 10% of patients with MBC carry BRCA2 mutations and there is a high relative risk of breast cancer in men carrying these mutations (19). Depletion of BRCA1 protein is associated with increased cell proliferation in MBC similarly to breast cancer in females, although it has been reported that germline BRCA1 mutations are less frequent in men (20). Meijers-Heijboer et al. have indicated that the CHEK2 1100delC variant gives a 10-fold risk of male breast cancer independently of BRCA1 and BRCA2 (21). Mutations in the TP53 gene (22), the androgen receptor gene, PTEN tumor-suppressor gene, and mismatch repair genes (*e.g. hMLH1*) have also been reported in MBC (23, 24), but none of them has been unequivocally associated with an increased risk. It must be kept in mind that mutations in these genes are uncommon in the general population and that it is possible that much of the genetic involvement in breast cancer risk may be due to co-inheritance of several low-risk common variants (25).

As far as the etiology of breast cancer is concerned, it has been well established that cancer arises from successive genetic changes which affect a number of cellular processes, such as apoptosis, angiogenesis, proliferation, growth control and metastasis (26, 27). Gast et al. highlighted that research was guided toward identification of markers (28). By using genomic and transcriptomic approaches, researchers were able to expand our insight into the genetic basis of cancer, by providing new biomarkers (29-31). We currently know that gene analysis in itself does not provide the complete picture. The genomics approach has its limitations. According to Chae et al. genomics "do not capture post-translational modification that affects protein function and stability"(32). The same authors also state that "proteins are the ultimate effector molecule of cellular functions, not genes or messenger RNAs". Therefore, the proteome, rather than the genome, is now considered to provide a more accurate reflection of both the genetic background of the cell, as well as its control over the immediate environment (33). Bearing in mind that proteomic analysis may provide the association between gene sequence and cellular physiology (34), we anticipate proteomics actually to complement gene analyses in its use in the prognosis, and evaluation of disease and its response to treatment (35).

In this review, we summarize studies and results linked to the implementation of proteomics in the field of MBC detection and diagnosis.

Proteomics

The ultimate aim of proteomics is the characterization of the information cascade *via* protein networks. This information may be the cause, or the consequence, of a disease's development (36). With the term "clinical proteomics", we refer to a sub-category of proteomics that deals with the application of proteomic technologies to clinical samples, such as blood, in order to identify unique biomarkers and biosignatures. According to Huijbers *et al.* "a biomarker, or biological marker, is a biomolecule that can be used as an indicator of a disease, based on abnormal presence, absence or changes in genes, RNA, proteins or metabolites" (37). Such proteomic technologies have application in cancer; cancer is characterized by multiple dysregulated proteins and cellular pathways involved in the onset and progression of the disease (38).

Cancer biomarkers may be divided into three main categories, with diagnostic, prognostic, and potentially

predictive applications (39). Tissue, serum, plasma, cerebrospinal fluid, urine, saliva, ascites, nipple fluid, pleural fluid, or any other body fluids can be used as a matrix for the classification of breast cancer, the prediction of its response to therapy (e.g. targeted treatments, hormonal therapy) and for predicting prognosis by the discovery of proteomic biomarkers (32). Circulating biomarkers, such as serum, are ideal for less-invasive diagnostic procedures; the blood compartment, endowed with a protein-rich information archive (40), allows for multiple easy sample collection, thereby enhancing the clinical value of possible biomarkers (41, 42). Additionally, the proteomics approach may be able to identify protein-protein interactions, individual proteins or even driver pathways, leading towards identification of biomarker-based and personalized clinical trials in an effort to increase therapy success rate. With the development of novel targeted therapies in breast cancer, there is an increasing need for development of predictive proteomic biomarkers; in positive clinical trials, good responders may be identified with the application of proteomic techniques.

After significant efforts made in the field of clinical proteomics to discover novel breast cancer biomarkers in women, there is a small number of markers that has made it to clinical practice: uPA/PAI-1, circulating tumor cells, estrogen receptor (ER), progesterone receptor (PR), human epidermal growth factor receptor 2 (HER2), cancer antigen (CA)15-3 and CA27.29 (43). On the other hand, data on male breast protein alterations in breast carcinogenesis are scarce due to the rarity of the disease. Data on proteomics in women should not be extrapolated to men due to the inherent discrepancies between the two (44-48).

Unfortunately, there are great challenges in identifying discriminatory proteins and the use of specific techniques along with additional analytical tools is required. A number of techniques allow testing for proteins produced during specific diseases. Proteomic platforms are divided into antibody- and non antibody-based (32). Antibody-based techniques include enzyme-linked immunosorbent assay (ELISA), western blotting (protein immunoblot), immunohistochemistry (IHC), and protein microarray (chip). Non antibody-based approaches consist of methods based on mass spectrometric (MS) technology. While the former methods require prior knowledge of the proteins that are about to be tested, as well as specific antibodies, the latter do not.

In particular, the most widely used technique for the study of proteomic biomarkers in MBC is IHC. IHC is a conventional assay with high specificity able to assess the expression of proteins. The main advantage of IHC is the fact that it provides data on cellular and spatial localization of the protein, whereas the main disadvantages of the technique are that it is semiquantative, has a low-throughput and is laborintensive (49).

Protein microarray techniques have also been applied in a number of MBC proteomic studies. Protein microarray is a highly sensitive and high-throughput technology used for the evaluation of protein expression and interactions on multiple samples at once. In parallel, samples from more than 1,000 patients can be analyzed in a single array using a validated antibody for each protein of interest. There are distinct types of protein microarrays: analytical microarrays, functional arrays and reverse-phase protein arrays (50). In the first category, antibodies are arrayed on a surface and each array is probed with a solution that contains multiple proteins, allowing for the expression levels of a large number of proteins to be measured in parallel with a set of validated antibodies in a single array. Analytical microarrays can be used for the monitoring of differentially expressed proteins and therefore for clinical diagnostics (51). The arrays on functional protein microarrays contain either intact proteins or protein domains, and are widely used for the study of protein interactions. Lastly, in reverse-phase protein arrays a set of lysed tumor cells is immobilized onto a nitrocellulose surface and then the slide is probed with one validated antibody and later with affinity reagent. By using this technique, it becomes possible to determine the presence of altered proteins, which may indicate disease, making it an eligible platform for the discovery of new biomarkers in MBC (52).

Finally, MS has shown great potential in breast cancer studies. Protein MS is a highly sensitive analytical chemistry technique that generates a mass spectrum of proteins contained in a sample of material; the sample may be gas, solid, or liquid. MS may prove able to discover new molecular markers for early detection, prognosis and prediction of response to various chemotherapeutical agents (53-55). The principle step of MS involves ionizing chemical compounds such as intact proteins ('top-down approach') or peptides derived from enzymatically digested proteins ('bottom-up approach'). The latter approach has the advantage of providing more information per protein, since the ionization of peptides is easier than that of whole proteins (43). Subsequently, the charged molecules (proteins) or charged molecular fragments (peptides) are introduced into a mass analyze which measures their mass-to-charge ratios. Finally, the analysis of the spectra allows scientists to determine not only the protein composition of a biological sample, but also to structurally identify proteins (56). With recent advances in MS, the field of proteomics opens up new horizons for the scientific committee to discover promising tumor alterations as far as classification, prognosis and diagnosis of MBC is concerned.

Immunohistochemistry

According to IHC, breast carcinoma is divided into different subtypes; more specifically, female breast cancer is divided into luminal A and B, HER2-enriched and triple-negative subtype according to the status of ER, PR, HER2 and Ki-67 (57, 58). Of note, different treatment modalities are applied according to this categorization, *i.e.* hormonal treatment (aromatase inhibitors, tamoxifen, fulvestrant) is administered in cases of overexpression of ER and/or PR; anti-HER2 therapy (trastuzumab, pertuzumab, lapatinib, TDM1) is added to conventional therapy in tumors that overexpress HER2, *etc.* (59, 60).

Different independent research groups have tried to evaluate ER, PR, HER2 and Ki-67 status in MBC in order to subclassify the disease; these attempts in the vast majority of cases were retrospective and based on limited number of cases (Table I). ER is expressed in 75-96.9% of MBC cases amongst different published reports (61-70) and PR in 58.8-96% (61-63, 65, 67, 68); HER2 is overexpressed in 1.7-29% of MBC (61, 63, 66, 68, 71, 72). According to these data, the majority of men with breast cancer are classified as having luminal A subtype (65.7-83%) (63, 65, 73). Consequently, it is obvious that this categorization does not add much to the management of MBC and there is an unmet need for additional biomarkers in MBC. At this point, a report published by Johansson et al. (74) tried to sub-classify breast cancer in men in luminal M1 and M2, and suggested that these two distinct subgroups differ from the well-established intrinsic subtypes of breast cancer in women.

Consequently, there has been an effort for the evaluation of other protein markers in MBC using IHC. Androgen receptor (AR) is expressed in the majority of MBC tumors (95-81%) (68, 75), signifying the potential of anti-androgen treatment in MBC. Proteins under androgen control, such as pepsinogen C, are more highly expressed in MBC than in females with breast cancer (76). Apolipoprotein D, another androgen-induced marker, is also overexpressed in MBC (76, 77) and has been positively associated with favorable outcome in men (77). Furthermore, androgen-regulated protein prostate-specific antigen (PSA), which is used as a diagnostic marker for metastatic prostate carcinoma, is also expressed in some cases of MBC (75). Of note, connective tissue growth factor has been found to be a marker for poor prognosis; it is expressed at a high percentage in MBC and has been correlated with high proliferative index and high grade, implicating its crucial role in breast carcinogenesis (78). Additionally, a number of studies revealed a higher percentage of B-cell lymphoma 2 (BCL2) positivity in MBC (94-78.9%) (47, 66, 68, 79), implicating anti-apoptotic mechanisms in MBC (68). Furthermore, p53 immunopositivity was detected in 9-21% of patients with MBC (66-68); p53-negative tumors are more frequent in MBC than in females (80). Overexpression of p53, HER2, and c-MYC protein has been significantly correlated with poor prognosis (81). c-MYC has been detected in 20.8% of MBC cases analyzed (67). p21-Positive tumors are also significantly more frequent in MBC compared to that in females (80). Cyclin D1 is expressed in 58% of MBC cases et al. and according to Rayson et al. PFS was decreased for MBC patients with tumors staining negatively for cyclin D (68). On the other hand, Ki67 was found positively immunostained in 38% of MBC and was correlated with a decrease in PFS (68). Serra et al. examined lysozyme expression in 60 MBC tissues by using IHC: 45% of the sections stained positively for this milk protein. Further analysis confirmed that nodal involvement, as well as lysozyme value, were significant predictors of short-term relapse-free survival (82). Furthermore, according to Ciocca et al. cytokeratins 5/6 and 14 can be potentially used for the identification of a pathologically aggressive MBC form (83). Finally, a retrospective study of 24 paraffin blocks collected from MBC patients revealed expression of both α-smooth muscle actin and CD34 in 22/24 cases, although their detection cannot be considered specific to malignancy (84).

Protein Microarray

The application of protein microarray techniques in MBC biomarker detection is limited to only a few studies. According to a collaborative large-scale biomarker analysis (44), hormone-receptor profiles were compared between breast carcinomas from 251 males and 263 females; patients were matched for age, grade and lymph node status. In this study, tissue microarrays were immunostained for ER α , - β 1, - β 2, and -65, as well as PRa and -b, and AR, augmented by HER2, and CK5/6, -14, -18 and -19, to assist typing. Interestingly enough, in this study, luminal A subtype of breast cancer was the predominant phenotype in males as well as in females, whereas luminal B, basal and HER2 subtype were uncommon in MBC (Table I). Common clusters between male and female patients were revealed by hierarchical clustering, comprising total PR–PRa–PRb and ER β 1/2 clusters. Of note, the presence of ERa subunit on different clusters between the two groups was the most striking finding in this study; more specifically, in women with breast cancer, ERa clustered with PR and its isoforms, whereas in men with breast neoplasm, ERa clustered with AR and ER β isoforms. In another study tissue microarrays were immunostained for BCL2 on 151 MBC patients (85). According to this study, BCL2 was expressed in 94% of cases; more frequently than previously described in women with breast carcinoma. Of note, in the former study, BCL2 expression was not correlated with tumor size, grade, mitotic count or overall survival. Finally, Kornegoor et al. conducted a study on 134 cases of MBC, by IHC of tissue microarrays for ER, PR, AR, HER2, BRST2, cyclin D1, BCL2, p53, p16, p21, Ki67, CK5/6, CK14, and epidermal growth factor receptor (86). In their study, high mitotic count and high grade were correlated with high Ki67, HER2 amplification/overexpression, p53 accumulation, high p21 expression, low PR expression, and low BCL2 expression. Furthermore, according to this study, PR negativity and p53

accumulation were correlated with decreased overall survival. Interestingly enough, in unsupervised hierarchical clustering, four groups with distinctive clinicopathological features were identified; the PR-negative/ER-positive/high-grade cluster was correlated with the poorer overall survival.

Mass Spectrometry

Regarding MBC, only a single study has been performed using MS-based technologies. Chahed et al. performed a thorough proteomic study of protein-expression alterations in MBC (87). Overexpression of multiple proteins in male breast tumors were identified, namely an increase in the expression of structural proteins (CK8, -18 and -19; and tropomyosin 4), glycolytic enzymes (enolase 1), stress related proteins (peroxiredoxin 1 and peptidylprolyl isomerase A), enzymes involved in the synthesis of AMP (adenine phosphoribosyltransferase), heatshock proteins (HSP27), galectin 1 and cathepsin D, nuclear ribonucleoproteins (hnRNP K and A2/B1), ribosomal protein S2 and proteasome β 4. On the other hand, the level of tropomyosin 1 was found to be decreased in MBC tissues, insinuating a possible crucial role for this protein in MBC. Other down-regulated proteins that were identified in MBC were apolipoprotein A1 and annexin A2.

The aforementioned study was performed after the collection of breast tumor tissue sections and non-tumor tissue samples from nine patients with MBC. All patients included in the study had stage IIB at presentation and grade 2. After extraction of proteins and following solubilization, protein extracts were analyzed with 2-dimensional gel electrophoretic analysis. Specifically, both isoelectric focusing and non-equilibrium pH gradient electrophoretic analyses were performed three times. The comparison of each protein spot between healthy and tumor tissues led to the detection of proteins that were differentially expressed in MBC. Subsequently, these altered proteins were isolated; after tryptic digestion of the proteins, matrix-assisted laser desorption/ionization time of flight MS and search in protein databases were used for the identification of each protein. Western blot analyses confirmed elevated levels of CK18, tropomyosin 4, cathepsin D and HSP27 in MBC tumors. Immunohistochemical analyses of tropomyosin 4 were also performed on paraffin MBC sections and the aforementioned results were subsequently confirmed.

Conclusion

MBC is an uncommon disease with biological characteristics distinct from those of female breast cancer. However, due to the rarity of the disease and the limited published literature focusing on MBC, treatment modalities and data regarding prognosis are extrapolated from data based on women with breast cancer. Some studies have tried to provide new insights

Investigated marker	Proteomic technique	Correlation with MBC	Reference
Estrogen receptor	Immunohistochemistry + tissue microarrays	Expressed in 75-96.9% of MBC cases	44, 61-70, 86
Progesterone receptor	Immunohistochemistry + tissue microarrays	Expressed in 58.8- 96% of MBC cases	44, 61-63, 65, 67, 68, 86
Human epidermal growth factor receptor 2	Immunohistochemistry + tissue microarrays	Overexpressed in 1.7- 29% of MBC cases	61, 63, 66, 68, 71, 72, 86
Androven recentor	Immunohistochemistry ± tissue microarrays	Evnressed in 05-81% of MBC cases	44 68 75 86
			11, 00, 11, 00 11, 10, 10, 10
Pepsinogen C	Immunohistochemistry	More positive cells in MBC (/6.4%)	٩/
		than in FBC (50%)	
Apolipoprotein D	Immunohistochemistry	Overexpressed in MBC	76, 77
Prostate-specific antigen	Immunohistochemistry	Expressed in 23% of MBC cases	75
Connective tissue arouth factor	Immunohistochemistry	Expressed in 78% of MRC rases	82
		E	01 70 20 00 00 10
D-Cell lymphoma 2		Expressed III 94-70.9% OI MDC Cases	41,00,00,79,00,00
p-53	Immunohistochemistry + tissue microarrays	Expressed in 9-21% of MBC patients	66-68, 80, 86
		(lower percentage than in FBC)	
c-MYC	Immunohistochemistry	Expressed in 20.8% of MBC cases	67
Cyclin D1	Immunohistochemistry + tissue microarrays	Expressed in 58% of MBC cases	68, 86
MIB-1 (Ki67)	Immunohistochemistry	Expressed in 38% of MBC cases	68
p21	Immunohistochemistry	Expression higher in MBC vs. FBC	80
I veozuma	Imminohietochamietw	avaracced in A5% of MBC occes	60
		captessed III +J/ ULIVIDO Cases	70
Alpha-smooth muscle actin	Immunohistochemistry	Expressed in 91.7% of MBC cases	84
CD34	Immunohistochemistry	Expressed in 91.7% of MBC cases	84
Cytokeratin 8	2-DE and MALDI-TOF/MS	Overexpressed in MBC	87
Cytokeratin 18	2-DE and MALDI-TOF/MS + Western blot	Overexpressed in MBC	87
Cytokeratin 19	2-DE and MALDI-TOF/MS	Overexpressed in MBC	87
Tropomyosin 4	2-DE and MALDI-TOF/MS + western blot +	Overexpressed in MBC	87
•	immunohistochemistry		
Enolase 1	2-DE and MALDI-TOF/MS	Overexnressed in MBC	87
Deroviredovin 1	2-DF and MALDL-TOF/MS	Overexpressed in MBC	87
Doutidulandul i	CHILDLET THE PUP STATE		- C 0
repudyiprotyl isomerase A			10
Adenine phosphoribosyltransterase		Overexpressed in MBC	87
Heat-shock protein 27	2-DE and MALDI-TOF/MS + western blot	Overexpressed in MBC	87
Galectin 1	2-DE and MALDI-TOF/MS	Overexpressed in MBC	87
Cathepsin D	2-DE and MALDI-TOF/MS + western blot	Overexpressed in MBC	87
Nuclear ribonucleoproteins K and A2/B1	2-DE and MALDI-TOF/MS	Overexpressed in MBC	87
Rihosomal protein S2	2-DE and MALDI-TOF/MS	Overexpressed in MBC	87
Drotascoma hata A	2-DF and MAI DI-TOFAAS	Overeversed in MBC	L8
I ropomyosin 1	2-DE and MALDI-IOF/MS	Down-regulated in MBC	8/
Apolipoprotein A1	2-DE and MALDI-TOF/MS	Down-regulated in MBC	87
Annexin A2	2-DE and MALDI-TOF/MS	Down-regulated in MBC	87

187

into the molecular mechanisms of MBC, but these findings must be verified in large-scale prospective well-designed case studies. Of note, for example MS-based technologies seem to be appealing, but unfortunately, to the best of our knowledge only one study on MBC has been published. Furthermore, the clinical application of other techniques in MBC is limited. Hence, there is an unmet need for novel well-designed, collaborative studies, focusing on MBC and evaluating protein biomarkers in this subgroup.

Conflicts of Interest

None.

References

- 1 Jemal A, Siegel R, Ward E, Hao Y, Xu J, Murray T and Thun MJ: Cancer Statistics, 2008. CA Cancer J Clin 58: 71-96, 2008.
- 2 Petridou, E, Giokas, G, Kuper, H, Mucci LA and Trichopoulos D: Endocrine correlates of male breast cancer risk: a case–control study in Athens, Greece. Br J Cancer 83: 1234-1237, 2000.
- 3 American Cancer Society: Cancer Facts and Figures 2015. Atlanta, GA: American Cancer Society, 2015.
- 4 Payson BA and Rosh R: Carcinoma and other neoplasms of the male breast. Radiology *52*: 220-229, 1949.
- 5 Geschickter CF, Lewis D and Hartman CG: Tumors of the breast related to the oestrin hormone. Am J Cancer 21: 828-859, 1934.
- 6 Giordano SH, Cohen DS, Buzdar AU, Perkins G and Hortobagyi GN: Breast carcinoma in men: a population-based study. Cancer 101: 51-57, 2004.
- 7 Swerdlow AJ, Schoemaker MJ, Higgins CD, Wright AF, Jacobs PA and UK Clinical Cytogenetics Group: Cancer incidence and mortality in men with Klinefelter syndrome: a cohort study. J Natl Cancer Inst 97: 1204-1210, 2005.
- 8 Humphries MP, Jordan VC and Speirs V: Obesity and male breast cancer: provocative parallels? BMC Medicine *13*: 134, 2015.
- 9 Hsing AW, McLaughlin JK, Cocco P, Co Chien HT and Fraumeni JF Jr.: Risk factors for male breast cancer (United States). Cancer Causes Control 9: 269-275, 1998.
- 10 Sørensen HT, Friis S, Olsen JH, Thulstrup AM, Mellemkjaer L, Linet M, Trichopoulos D, Vilstrup H and Olsen J: Risk of breast cancer in men with liver cirrhosis. Am J Gastroenterol 93: 231-233, 1998.
- 11 Guénel P, Cyr D, Sabroe S, Lynge E, Merletti F, Ahrens W, Baumgardt-Elms C, Ménégoz F, Olsson H, Paulsen S, Simonato L and Wingren G: Alcohol drinking may increase risk of breast cancer in men: a European population-based case-control study. Cancer Causes Control 15: 571-580, 2004.
- 12 Thomas DB, Jimenez LM, McTiernan A, Rosenblatt K, Stalsberg H, Stemhagen A, Thompson WD, Curnen MG, Satariano W and Austin DF: Breast cancer in men: risk factors with hormonal implications. Am J Epidemiol 135: 734-748, 1992.
- 13 Medras M, Filus A, Jozkow P, Winowski J and Sicinska-Werner T: Breast cancer and long-term hormonal treatment of male hypogonadism. Breast Cancer Res Treat 96: 263-265, 2006.
- 14 Kanhai RC, Hage JJ, van Diest PJ, Bloemena E and Mulder JW: Short-term and long-term histologic effects of castration and

estrogen treatment on breast tissue of 14 male-to-female transsexuals in comparison with two chemically castrated men. Am J Surg Pathol 24: 74-80, 2000.

- 15 Brinton LA, Key TJ, Kolonel LN, Michels KB, Sesso HD, Ursin G, Van Den Eeden SK, Wood SN, Falk RT, Parisi D, Guillemette C, Caron P, Turcotte V, Habel LA, Isaacs CJ, Riboli E, Weiderpass E and Cook MB: Prediagnostic sex steroid hormones in relation to male breast cancer risk. J Clin Oncol 33: 2041-2050, 2015.
- 16 Ron E, Ikeda T, Preston DL and Tokuoka S: Male breast cancer incidence among atomic bomb survivors. J Natl Cancer Inst 97: 603-605, 2005.
- 17 Thomas DB, Rosenblatt K, Jimenez LM, McTiernan A, Stalsberg H, Stemhagen A, Thompson WD, Curnen MG, Satariano W, Austin DF, Greenberg RS, Key C, Kolonel LN and West DW: Ionizing radiation and breast cancer in men (United States). Cancer Causes Control 5: 9-14, 1994.
- 18 Demers PA, Thomas DB, Rosenblatt KzA, Jimenez LM, McTiernan A, Stalsberg H, Stemhagen A, Thompson WD, Curnen MG, Satariano W Austin DF, Isacson P, Greenberg RS, Key C, Kolonel LN and West DW: Occupational exposure to electromagnetic fields and breast cancer in men. Am J Epidemiol *134*: 340-347, 1991.
- 19 Thompson D, Easton D and Breast Cancer Linkage Consortium: Variation in cancer risks, by mutation position, in BRCA2 mutation carriers. Am J Hum Genet 68: 410-419, 2001.
- 20 Gómez-Raposo C, Zambrana Tévar F, Sereno Moyano M, López Gómez M and Casado E: Male breast cancer. Cancer Treat Rev 36: 451-457, 2010.
- 21 Meijers-Heijboer H, van den Ouweland A, Klijn J, Wasielewski M, de Snoo A, Oldenburg R, Hollestelle A, Houben M, Crepin E, van Veghel-Plandsoen M, Elstrodt F, van Duijn C, Bartels C, Meijers C, Schutte M, McGuffog L, Thompson D, Easton D, Sodha N, Seal S, Barfoot R, Mangion J, Chang-Claude J, Eccles D, Eeles R, Evans DG, Houlston R, Murday V, Narod S, Peretz T, Peto J, Phelan C, Zhang HX, Szabo C, Devilee P, Goldgar D, Futreal PA, Nathanson KL, Weber B, Rahman N, Stratton MR and CHEK2-Breast Cancer Consortium: Low-penetrance susceptibility to breast cancer due to CHEK2 1100delC in noncarriers of BRCA1 or BRCA2 mutations. Nat Genet *31*: 55-59, 2002.
- 22 Anelli A, Anelli TF, Youngson B, Rosen PP and Borgen PI: Mutations of the p53 gene in male breast cancer. Cancer 75: 2233-2238, 1995.
- 23 Fackenthal JD1, Marsh DJ, Richardson AL, Cummings SA, Eng C, Robinson BG and Olopade OI: Male breast cancer in Cowden syndrome patients with germline PTEN mutations. J Med Genet 38: 159-164, 2001.
- 24 Boyd J, Rhei E, Federici MG, Borgen I, Watson P, Franklin B, Karr B, Lynch J, Lemon SJ and Lynch HT: Male breast cancer in the hereditary nonpolyposis colorectal cancer syndrome. Breast Cancer Res Treat 53: 87-91, 1999.
- 25 Fletcher O and Houlston RS: Architecture of inherited susceptibility to common cancer. Nat Rev Cancer *10*: 353-361, 2010.
- 26 Mommers EC, van Diest PJ, Leonhart AM, Meijer CJ and Baak JP: Balance of cell proliferation and apoptosis in breast carcinogenesis. Breast Cancer Res Treat 58: 163-169, 1999.
- 27 Reis-Filho J and Lakhani SR: The diagnosis and management of pre-invasive breast disease: genetic alterations in pre-invasive lesions. Breast Cancer Res 5: 313-319, 2003.
- 28 Gast MC, Schellens JH and Beijnen JH: Clinical proteomics in breast cancer: a review. Breast Cancer Res Treat 116: 17-29, 2009.

- 29 van 't Veer LJ, Dai H, van de Vijver MJ, He YD, Hart AA, Mao M, Peterse HL, van der Kooy K, Marton MJ, Witteveen AT, Schreiber GJ, Kerkhoven RM, Roberts C, Linsley PS, Bernards R and Friend SH: Gene-expression profiling predicts clinical outcome of breast cancer. Nature 415: 530-536, 2002.
- 30 Wang Y, Klijn JG, Zhang Y, Sieuwerts AM, Look MP, Yang F, Talantov D, Timmermans M, Meijer-van Gelder ME, Yu J, Jatkoe T, Berns EM, Atkins D and Foekens JA: (2005) Gene-expression profiles to predict distant metastasis of lymph-node-negative primary breast cancer. Lancet 365: 671-679, 2005.
- 31 Vogelstein B and Kinzler KW: Has the breast cancer gene been found? Cell 79(1): 1-3, 1994.
- 32 Chae YK and Gonzalez-Angulo AM: Implications of functional proteomics in breast cancer. Oncologist 19: 328-335, 2014.
- 33 Aebersold R, Anderson L, Caprioli R, Druker B, Hartwell L and Smith R: Perspective: a program to improve protein biomarker discovery for cancer. J Proteome Res 4: 1104-1109, 2005.
- 34 Dove A: Proteomics: Translating genomics into products? Nat Biotechnol *17*: 233-236, 1999.
- 35 Clarke W, Zhang Z and Chan DW: The application of clinical proteomics to cancer and other diseases. Clin Chem Lab Med 41: 1562-1570, 2003.
- 36 Petricoin EF, Zoon KC, Kohn EC, Barrett JC and Liotta LA: Clinical proteomics: translating benchside promise into bedside reality. Nat Rev Drug Discov 1: 683-695, 2002.
- 37 Huijbers A, Velstra B, Dekker TJA, Mesker WE, van der Burgt YEM, Mertens BJ, Deelder AM and Tollenaar RAEM, Proteomic serum biomarkers and their potential application in cancer screening programs. Int J Mol Sci 11: 4175-4193, 2010..
- 38 Khadir A and Tiss A: Proteomics approaches towards early detection and diagnosis of cancer. J Carcinogene Mutagene S14:002, 2013.
- 39 Gam LH: Breast cancer and protein biomarkers. World J Exp Med 2: 86-91, 2012.
- 40 Petricoin EF, Zoon KC, Kohn EC, Barrett JC and Liotta LA: Clinical proteomics: translating benchside promise into bedside reality. Nat Rev Drug Discov *1*: 683-695, 2002.
- 41 Conrads TP, Zhou M, Petricoin EF 3rd, Liotta L and Veenstra TD: Cancer diagnosis using proteomic patterns. Expert Rev Mol Diagn 3: 411-420, 2003.
- 42 Good DM, Thongboonkerd V, Novak J, Bascands JL, Schanstra JP, Coon JJ, Dominiczak A and Mischak H: Body fluid proteomics for biomarker discovery: lessons from the past hold the key to success in the future. J Proteome Res 6: 4549-4555, 2007.
- 43 Zeidan BA, Townsend PA, Garbis SD, Copson E and Cutress RI: Clinical proteomics and breast cancer. Surgeon 13: 271-278, 2015.
- 44 Shaaban AM, Ball GR, Brannan RA, Cserni G, Di Benedetto A, Dent J, Fulford L, Honarpisheh H, Jordan L, Jones JL, Kanthan R, Maraqa L, Litwiniuk M, Mottolese M, Pollock S, Provenzano E, Quinlan PR, Reall G, Shousha S, Stephens M, Verghese ET, Walker RA, Hanby AM and Speirs V: A comparative biomarker study of 514 matched cases of male and female breast cancer reveals gender-specific biological differences. Breast Cancer Res Treat 133: 949-958, 2012.
- 45 Kornegoor R, Verschuur-Maes AH, Buerger H, Hogenes MC, de Bruin PC, Oudejans JJ, van der Groep P, Hinrichs B and van Diest PJ: Molecular subtyping of male breast cancer by immunohistochemistry. Mod Pathol 25: 398-404, 2012.
- 46 Kornegoor R, Moelans CB, Verschuur-Maes AH, Hogenes MC, de Bruin PC, Oudejans JJ, Marchionni L and van Diest PJ:

Oncogene amplification in male breast cancer: analysis by multiplex ligation-dependent probe amplification. Breast Cancer Res Treat *135*: 49-58, 2012.

- 47 Weber-Chappuis K, Bieri-Burger S and Hurlimann J: Comparison of prognostic markers detected by immunohistochemistry in male and female breast carcinomas. Eur J Cancer 2A: 1686-1692, 1996.
- 48 Lacle MM, Kornegoor R, Moelans CB, Maes-Verschuur AH, van der Pol C, Witkamp AJ van der Wall E, Rueschoff J, Buerger H and van Diest PJ: Analysis of copy number changes on chromosome 16q in male breast cancer by multiplex ligationdependent probe amplification. Mod Pathol 26: 1461-1467, 2013.
- 49 Ramos-Vara JA and Miller MA: When tissue antigens and antibodies get along: revisiting the technical aspects of immunohistochemistry-the red, brown, and blue technique. Vet Pathol 51: 42-87, 2014.
- 50 Wingren C and Borrebaeck CA: Antibody-based microarrays. Methods Mol Biol 509: 57-84, 2009.
- 51 Sreekumar A, Nyati MK, Varambally S, Barrette TR, Ghosh D, Lawrence TS and Chinnaiyan AM: Profiling of cancer cells using protein microarrays: discovery of novel radiation-regulated proteins. Cancer Res 61: 7585-7593, 2001.
- 52 Hennessy BT, Lu Y, Gonzalez-Angulo AM, Carey MS, Myhre S, Ju Z, Davies MA, Liu W, Coombes K, Meric-Bernstam F, Bedrosian I, McGahren M, Agarwal R, Zhang F, Overgaard J, Alsner J, Neve RM, Kuo WL, Gray JW, Borresen-Dale AL and Mills GB: A technical assessment of the utility of reverse-phase protein arrays for the study of the functional proteome in non-microdissected human breast cancers. Clin Proteomics *6*: 129-151, 2010.
- 53 Yang WS, Moon HG, Kim HS, Choi EJ, Yu MH, Noh DY and Lee C: Proteomic approach reveals FKBP4 and S100A9 as potential prediction markers of therapeutic response to neoadjuvant chemotherapy in patients with breast cancer. J Proteome Res 11: 1078-1088, 2012.
- 54 Kabbage M, Chahed K, Hamrita B, Guillier CL, Trimeche M, Remadi S, Hoebeke J and Chouchane L: Protein alterations in infiltrating ductal carcinomas of the breast as detected by nonequilibrium pH gradient electrophoresis and mass spectrometry. J Biomed Biotechnol 2008: 564127, 2008.
- 55 Nakagawa T, Huang SK, Martinez SR, Tran AN, Elashoff D, Ye X, Turner RR, Giuliano AE and Hoon DS: Proteomic profiling of primary breast cancer predicts axillary lymph node metastasis. Cancer Res 66: 11825-11830, 2006.
- 56 Hernandez P, Müller M and Appel RD: Automated protein identification by tandem mass spectrometry: issues and strategies. Mass Spectrom Rev 25: 235-254, 2006.
- 57 Perou CM, Sørlie T, Eisen MB, van de Rijn M, Jeffrey SS, Rees CA, Pollack JR, Ross DT, Johnsen H, Akslen LA, Fluge O, Pergamenschikov A, Williams C, Zhu SX, Lønning PE, Børresen-Dale AL, Brown PO and Botstein D: Molecular portraits of human breast tumours. Nature 406: 747-752, 2000.
- 58 Goldhirsch A, Wood WC, Coates AS, Gelber RD, Thürlimann B, Senn HJ and Panel members: Strategies for subtypes-dealing with the diversity of breast cancer: highlights of the St. Gallen International Expert Consensus on the Primary Therapy of Early Breast Cancer 2011. Ann Oncol 22: 1736-1747, 2011.
- 59 Jones RL, Salter J, A'Hern R, Nerurkar A, Parton M, Reis-Filho JS, Smith IE and Dowsett M: Relationship between oestrogen receptor status and proliferation in predicting response and long-term outcome to neoadjuvant chemotherapy for breast cancer. Breast Cancer Res Treat *119*: 315-323, 2010.

- 60 Wang Y, Ikeda DM, Narasimhan B, Longacre TA, Bleicher RJ, Pal S, Jackman RJ and Jeffrey SS: Estrogen receptor-negative invasive breast cancer: imaging features of tumors with and without human epidermal growth factor receptor type 2 overexpression. Radiology 246: 367-375, 2008.
- 61 Masci G, Caruso M, Caruso F, Salvini P, Carnaghi C, Giordano L, Miserocchi V, Losurdo A, Zuradelli M, Torrisi R, Di Tommaso L, Tinterri C, Testori A, Garcia-Etienne CA, Gatzemeier W and Santoro A: Clinicopathological and immunohistochemical characteristics in male breast cancer: a retrospective case series. Oncologist 20: 586-592, 2015.
- 62 Shandiz FH, Tavassoli A, Sharifi N, Khales SA, Kadkhodayan S and Khales SA: Hormone receptor expression and clinicopathologic features in male and female breast cancer. Asian Pac J Cancer Prev 16: 471-474, 2015.
- 63 Nilsson C, Johansson I, Ahlin C, Thorstenson S, Amini RM, Holmqvist M, Bergkvist L, Hedenfalk I and Fjällskog ML: Molecular subtyping of male breast cancer using alternative definitions and its prognostic impact. Acta Oncol 52: 102-109, 2013.
- 64 Schildhaus HU, Schroeder L, Merkelbach-Bruse S, Binot E, Büttner R, Kuhn W and Rudlowski C: Therapeutic strategies in male breast cancer: clinical implications of chromosome 17 gene alterations and molecular subtypes. Breast 22: 1066-1071, 2013.
- 65 Ge Y, Sneige N, Eltorky MA, Wang Z, Lin E, Gong Y and Guo M: Immunohistochemical characterization of subtypes of male breast carcinoma. Breast Cancer Res 11: R28, 2009.
- 66 Muir D, Kanthan R and Kanthan SC: Male versus female breast cancers. A population-based comparative immunohistochemical analysis. Arch Pathol Lab Med 127: 36-41, 2003.
- 67 Mourão Netto M, Logullo AF, Nonogaki S, Brentani RR and Brentani MM: Expression of c-ERBB-2, p53 and c-myc proteins in male breast carcinoma: Comparison with traditional prognostic factors and survival. Braz J Med Biol Res 34: 887-894, 2001.
- 68 Rayson D, Erlichman C, Suman VJ, Roche PC, Wold LE, Ingle JN and Donohue JH: Molecular markers in male breast carcinoma. Cancer 83: 1947-1955, 1998.
- 69 Rogers S, Day CA and Fox SB: Expression of cathepsin D and estrogen receptor in male breast carcinoma. Hum Pathol 24: 148-151, 1993.
- 70 Dawson PJ, Paine TM and Wolman SR: Immunocytochemical characterization of male breast cancer. Mod Pathol 5: 621-625, 1992.
- 71 Rudlowski C, Friedrichs N, Faridi A, Füzesi L, Moll R, Bastert G, Rath W and Büttner R: HER2/neu Gene amplification and protein expression in primary male breast cancer. Breast Cancer Res Treat 84: 215-223, 2004.
- 72 Bloom KJ, Govil H, Gattuso P, Reddy V and Francescatti D: Status of HER-2 in male and female breast carcinoma. Am J Surg 182: 389-392, 2001.
- 73 Aşchie M, Bălţătescu GI and Mitroi A: Clinico-pathological and molecular subtypes of male breast carcinoma according to immunohistochemistry. Rom J Morphol Embryol 54(3 Suppl): 749-755, 2013.
- 74 Johansson I, Nilsson C, Berglund P, Lauss M, Ringnér M, Olsson H, Luts L, Sim E, Thorstensson S, Fjällskog ML and Hedenfalk I: Gene expression profiling of primary male breast cancers reveals two unique subgroups and identifies N-acetyltransferase-1 (NAT1) as a novel prognostic biomarker. Breast Cancer Res 14: R31, 2012.

- 75 Kidwai N, Gong Y, Sun X, Deshpande CG, Yeldandi AV, Rao MS and Badve S: Expression of androgen receptor and prostatespecific antigen in male breast carcinoma. Breast Cancer Res 6: R18-23, 2004.
- 76 Serra C, Vizoso F, Lamelas ML, Rodríguez JC, González LO, Merino AM, Baltasar A, Pérez-Vázquez MT and Medrano J: Comparative study of two androgen-induced markers (apolipoprotein D and pepsinogen C) in female and male breast carcinoma. Int J Surg Investig 2: 183-192, 2000.
- 77 Serra Díaz C, Vizoso F, Lamelas ML, Rodríguez JC, González LO, Baltasar A and Medrano J: Expression and clinical significance of apolipoprotein D in male breast cancer and gynaecomastia. Br J Surg 86: 1190-1197, 1998.
- 78 Lacle MM, van Diest PJ, Goldschmeding R, van der Wall E and Nguyen TQ: Expression of connective tissue growth factor in male breast cancer: clinicopathologic correlations and prognostic value. PLoS ONE 10: e0118957, 2015.
- 79 Pich A, Margaria E and Chiusa L: BCL2 expression in male breast carcinoma. Virchows Arch 433: 229-235, 1998.
- 80 André S, Pinto AE, Laranjeira C, Quaresma M and Soares J: Male and female breast cancer–differences in DNA ploidy, p21 and p53 expression reinforce the possibility of distinct pathways of oncogenesis. Pathobiology 74: 323-327, 2007.
- 81 Pich A, Margaria E and Chiusa L: Oncogenes and male breast carcinoma: c-ERBB-2 and p53 coexpression predicts a poor survival. J Clin Oncol 18: 2948-2956, 2000.
- 82 Serra C, Vizoso F, Alonso L, Rodríguez JC, González LO, Fernández M, Lamelas ML, Sánchez LM, García-Muñiz JL, Baltasar A and Medrano J: Expression and prognostic significance of lysozyme in male breast cancer. Breast Cancer Res 4: R16, 2002.
- 83 Ciocca V, Bombonati A, Gatalica Z, Di Pasquale M, Milos A, Ruiz-Orrico A, Dreher D, Folch N, Monzon F, Santeusanio G, Perou CM, Bernard PS and Palazzo JP: Cytokeratin profiles of male breast cancers. Histopathology 49: 365-370, 2006.
- 84 Kalekou H, Kostopoulos I, Milias S and Papadimitriou CS: Comparative study of CD34, alpha-SMA and h-caldesmon expression in the stroma of gynaecomastia and malebreast carcinoma. Histopathology 47: 74-81, 2005.
- 85 Lacle MM, van der Pol C, Witkamp A, van der Wall E and van Diest PJ: Prognostic value of mitotic index and BCL2 expression in male breast cancer. PLoS One 8: e60138, 2013.
- 86 Kornegoor R, Verschuur-Maes AH, Buerger H, Hogenes MC, de Bruin PC, Oudejans JJ, Hinrichs B and van Diest PJ: Immunophenotyping of male breast cancer. Histopathology 61: 1145-1155, 2012.
- 87 Chahed K, Kabbage M, Hamrita B, Guillier CL, Trimeche M, Remadi S, Ehret-Sabatier L and Chouchane L: Detection of protein alterations in male breast cancer using two-dimensional gel electrophoresis and mass spectrometry: the involvement of several pathways in tumorigenesis. Clin Chim Acta 388: 106-114, 2008.

Received January 5, 2016 Revised February 19, 2016 Accepted February 23, 2016