

Review

MicroRNAs Involved in Small-cell Lung Cancer as Possible Agents for Treatment and Identification of New Targets

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Abstract. *Small-cell lung cancer, a neuro-endocrine type of lung cancers, responds very well to chemotherapy-based agents. However, a high frequency of relapse due to adaptive resistance is observed. Immunotherapy-based treatments with checkpoint inhibitors has resulted in improvement of treatment but the responses are not as impressive as in other types of tumor. Therefore, identification of new targets and treatment modalities is an important issue. After searching the literature, we identified eight down-regulated microRNAs involved in radiation- and chemotherapy-induced resistance, as well as three up-regulated and four down-regulated miRNAs with impacts on proliferation, invasion and apoptosis of small-cell lung cancer cells in vitro. Furthermore, one up-regulated and four down-regulated microRNAs with in vivo activity in SCLC cell xenografts were identified. The identified microRNAs are candidates for inhibition or reconstitution therapy. The corresponding targets are candidates for inhibition or functional reconstitution with antibody-based moieties or small molecules.*

Small-cell lung cancer (SCLC) is an exceptionally lethal malignancy comprising 13-15% of all lung cancer (1), with 250,000 cases diagnosed annually worldwide (1). SCLC is highly sensitive to platinum-based chemotherapy, topoisomerase inhibitor etoposide, and to lurbinectidin, a recently approved DNA binder (2, 3). However, disease

recurrence and metastasis to the brain, adrenal glands, bone and liver after treatment remains an issue (2). Inactivating mutations in retinoblastoma (*RB*) or *TP53* are most commonly observed, however, these alterations are not druggable and, in contrast to non-small-cell lung cancer no tractable drivers or fusion proteins have been observed (4). Monoclonal antibodies nivolumab and pembrolizumab, directed against checkpoint inhibitors, have been approved as first-line treatment of SCLC together with chemotherapy and for treatment of relapsed disease (5). However, the therapeutic benefit is not as pronounced as observed with other types of tumor (5). Furthermore, SCLC subtypes have been identified based on differential expression of transcription factors achaete-scute homolog (ASCL1), neurogenic differentiation factor (NeuroD1), yes-associated protein (YAP1) and POU class 2 homeobox 3 (POU2F3) (6). These subtypes might respond to drugs with different vulnerability (6). Several agents targeting T-cell immunoreceptor with Ig and ITIM domains (TIGIT), cytotoxic T-lymphocyte antigen 4 (CTLA4), or cyclin-dependent kinases 4 and 6 are in phase III clinical studies or under Food and Drug Administration review (7). Nevertheless, there is an urgent need to identify new targets and treatment modalities for SCLC. Here, we focus on microRNAs (miRs) as therapeutic agents and as tools for identification of SCLC-related targets for therapeutic intervention.

MicroRNAs – Role in Oncology

miRs are synthesized by RNA polymerase II in the nucleus as precursor RNAs, processed and exported into the cytoplasm (8-10). One strand of a 22 nucleotides (nts) complex is maintained (guide strand), the other strand (passenger strand) is discarded (8-10). The guide strand binds to the 3'-untranslated region of corresponding mRNAs and induces their degradation or inhibits their translation (8-10). A single miR can interact with several different mRNAs and therefore can interfere with several pathways and has the potential to rewire oncogenic pathways (11). miRs can exert an oncogenic or tumor-suppressive role, depending on the context (12). A

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tumor-suppressive role is mediated by *miR-16-1* and *miR-15a* by targeting anti-apoptotic protein BCL2 apoptosis regulator (BCL2). Their deletion in mice causes B-cell chronic lymphocytic leukemia corresponding to the disease in humans and its cytogenetic characteristics (13, 14). The oncogenic role of miRs was demonstrated by induction of hepatocellular carcinoma through liver-specific expression of *miR-221* in transgenic mice (15). miRs can have an impact on all stages of carcinogenesis, including metastasis and anti-tumoral immune response (16). We recently summarized the impact of miRs on growth and metastasis of hepatocellular carcinoma (17), pancreatic cancer (18), non-small-cell lung carcinoma (19), breast cancer (20) and prostate cancer (21). In this review, we focus on the role of miRs with respect to chemoresistance, tumor growth and metastasis of SCLC.

miRs Involved in Chemoresistance and Radioresistance of SCLC

All of the miRs discussed are down-regulated in SCLC-cancer related cell lines or clinical specimen in comparison to corresponding controls.

miR-7. *miR-7* (Figure 1) affects the multidrug-resistance protein ATP-binding cassette subfamily C member 1 (ABCC1) (22), inward-rectifier potassium ion channel 2.1 (KIR2.1) (23) and poly (ADP-ribose) polymerase 1 (PARP1) (24).

ABCC1 expression is inversely correlated with *miR-7* (22). ABCC1 is a transmembrane drug transporter containing three membrane-spanning domains and two cytosolic nucleotide-binding domains (25) and is expressed in many types of multidrug-resistant cancer (26). Overexpression of ABCC1 is predictive for resistance to chemotherapy in SCLC (27, 28). A low-level expression of *miR-7* correlated with shorter overall survival in patients with SCLC (22). In the SCLC cell line H69AR, *miR-7* down-regulation was shown to be responsible for resistance to adriamycin and etoposide (22, 29).

miR-7 also targets KIR2.1, a member of the classical inward rectifying potassium channel family (23, 30-32). *KIR2.1* was up-regulated five-fold in H69AR cells in comparison to H69 SCLC cells (23). KIR2.1 induced resistance to apoptosis following exposure to chemotherapeutic drugs (23). Overexpression of KIR2.1 in H69 and H466 SCLC cells enhanced their growth in immuno-deficient mice (23). Up-regulation of *miR-7* sensitized H69AR cells to adriamycin, cisplatin and etoposide (23). RAS-protein kinase C–mitogen-activated protein kinase (MEK) signaling was identified as an important inducer of KIR2.1, which was down-regulated by RAS-protein kinase C inhibitor staurosporine and MEK inhibitor UO126 (23).

PARP1 was identified as a target in doxorubicin-resistant SCLC cell line H69AR in comparison to H69 parental cells (24). PARP1 was resolved as a target of *miR-7* (24, 34).

Inhibition of *miR-7* resulted in increased homologous repair in doxorubicin-resistant SCLC cells (24). *miR-7* reduced expression of breast cancer susceptibility protein 1 (BRCA1) and repair protein RAD51 homolog1 (RAD51), and disrupted homologous recombination-based repair, leading to doxorubicin resistance by targeting PARP1 (24). PARP1 has a multi-faceted role in DNA repair and chromatin remodeling (35). PARP1 inhibitors are approved anticancer agents based on a synthetic-lethality based mode of action (36-38).

miR-22. *miR-22* (Figure 1) was down-regulated in NCI-466 SCLC cells and inhibited radiosensitivity by targeting Werner helicase-interacting protein-1 (WRNIP1) (39). WRNIP1 is an ATPase which can protect replication forks and co-operates with RAD51 to safeguard the integrity and maintenance of the genome (40-42). Overexpression of *miR-22* promoted apoptosis and inhibited migration of NCI-466 cells (39).

miR-24-3p. Autophagy is a strategy by which resistance to chemotherapy is conferred (43, 44). Etoposide- and cisplatin-resistant SCLC cells exhibited increased autophagy (45). *miR-24-3p* (Figure 1) was down-regulated in SCLC cells and expression of autophagy-related 4A cysteine peptidase (ATG4A) was blocked (45). Expression of *miR-24-3p* can suppress autophagy of SCLC cells by directly targeting ATG4A (45). It has been shown that inhibitors of autophagy can sensitize chemoresistant cells to anticancer therapy in clinical trials (45, 46).

miR-100. *miR-100* (Figure 1) was shown to target homeobox transcription factor HOXA1, which was associated with poor prognosis in patients with SCLC, and its down-regulation mediated chemoresistance (47). HOXA1 was found to be expressed in 46% (29/63) of tumors from patients with SCLC. Expression of *miR-100* in multidrug-resistant SCLC cell line H69AR reversed resistance to cisplatin and etoposide (47). HOXA1 is involved in progression and prognosis of several types of tumor. It mediates tumor proliferation and poor prognosis in gastric cancer *via* cyclin D1 (48); enhances proliferation, invasion and metastasis of prostate cancer cells (49); and correlates with poor prognosis in patients with hepatocellular carcinoma (50).

miR-138. *miR-138* (Figure 1) was down-regulated in SCLC tissues and three corresponding cell lines (51). In NCI-H2081 SCLC cells, *miR-138* reduced cell growth and inhibited cell-cycle progression (51). Histone H2A variant X (H2AX) was identified as a target of *miR-138* (51). *H2AX* knockdown achieved a similar effect as observed for *miR-138* overexpression, whilst its induction abolished *miR-138*-mediated SCLC cell growth and inhibition of cell-cycle progression (51). Expression of *miR-138* was shown to confer SCLC cells with greater DNA-repair capacity and reduced

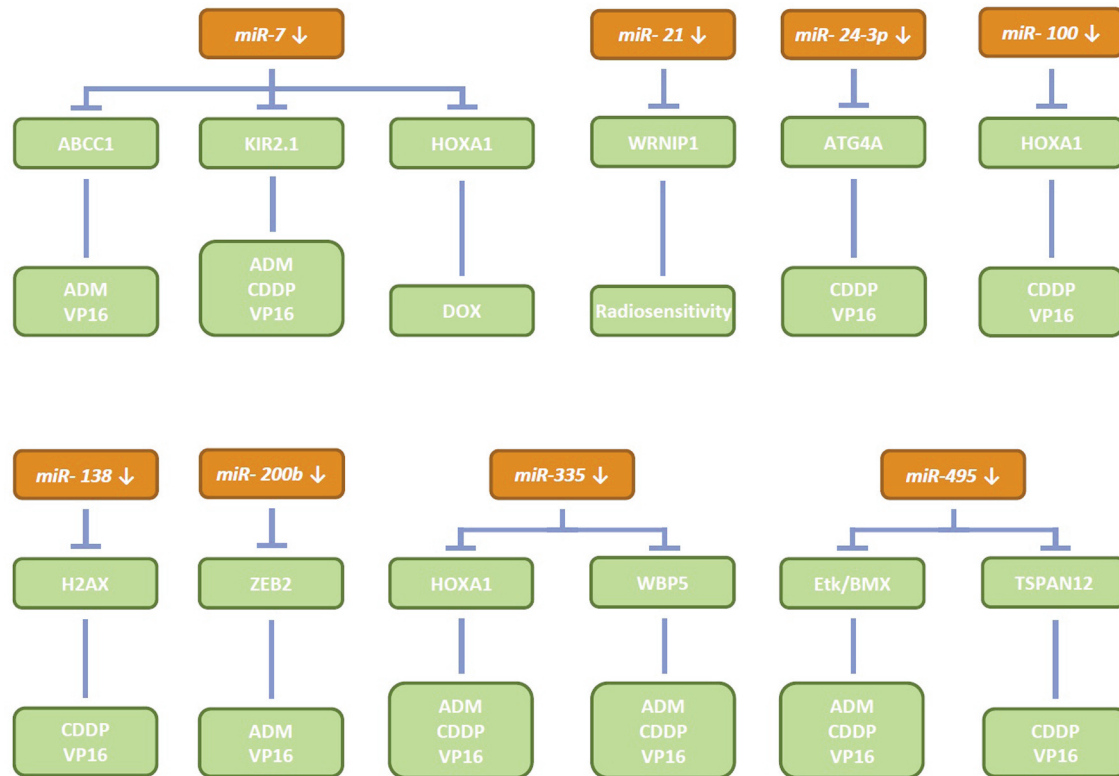


Figure 1. miRs involved in chemo- and radio-resistance of small-cell lung cancer (SCLC) cells. Downward arrows indicate down-regulation of miRs in SCLC in comparison to controls. ABCC1: ATB binding cassette subfamily C member; ADM: Adriamycin; ATG4A: autophagy-related protein 4; BMX: cytoplasmic tyrosine kinase BMX; CDDP: cisplatin; ETK: non-receptor tyrosine kinase Etk; H2AX: histone H2AX; HOXA1: homeobox protein HOXA1; KIR2.1: inward rectifier ion channel 2.1; PARP1: poly (ADP ribose)-polymerase 1; TSPAN 12: tetraspanin 12; VP16: etoposide; WBP5: ww domain-binding protein 1; WRNIP1: ATPase WRNIP1; ZEB2: zinc finger E-box-binding homeobox 2.

their resistance to chemotherapeutic agents (51). H2AX is involved in double-stranded DNA repair, chromatin remodeling and contributes to nucleosome formation (52-55).

miR-200b. *miR-200b* (Figure 1) targets zinc finger E-box homeobox2 (ZEB2), which correlated with poor pathologic stage and shorter survival (35). ZEB2 was found to be expressed in 23.5% (16/68) of cases of SCLC (56). Inhibition of ZEB2 expression making use of small-interfering RNA (siRNA), sensitized SCLC-related cells to chemotherapeutic drugs by enhancing drug-induced apoptosis accompanied by S-phase arrest (56). ZEB2 is a transcription factor with eight zinc fingers and a homeodomain (57). ZEB2 has been identified as a regulator of nervous system development (58, 59). In cancer, ZEB2 plays an instrumental role in epithelial mesenchymal transition (EMT), cancer-stem cell traits, apoptosis, survival, tumor recurrence and metastasis (60).

miR-335. *miR-335* (Figure 1) was found to target WW domain-binding protein 5 (WBP5), expression of which was

10-fold increased in H69AR compared to H69 SCLC cells. WBP5 induced multidrug resistance by promoting cell proliferation and inhibiting apoptosis in H69AR cells (61). Expression of WBP5 was associated with shorter survival in patients with SCLC (61). WW binding domains are typically 35-40 amino acids in length and can interact with a variety of different peptide ligands, including motifs with core proline-rich sequences (62). WBP5 was shown to be involved in multidrug resistance of SCLC through the Hippo pathway [WBP5-tyrosine kinase ABL-mammalian Ste-20-like kinase (MST2)-yes-associated protein1 (YAP1)] pathway (61). WBP5 can induce nuclear accumulation of YAP1, a transcription factor which induces genes involved in development and survival (63). Inhibition of YAP1 by verteporfin was shown to blunt multidrug resistance in H69AR cells (61). WBP5 can bind to ABL, an upstream activator of ser-thr kinase MST2 of the Hippo pathway (64,65). It was shown that WBP5 promotes tumor growth and resistance of H69 cells to adriamycin and cisplatin in nude mice (61).

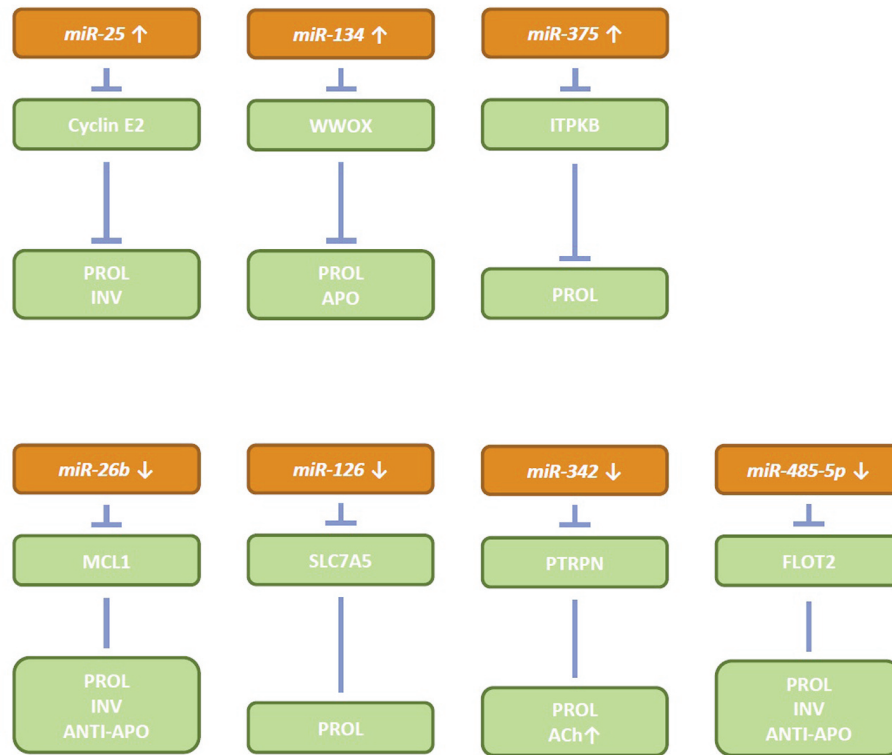


Figure 2. miRs Up- and down-regulated in small-cell lung cancer with in vitro activity. Upward arrows indicate increased expression and downward arrows point to decreased expression of the corresponding miRs in comparison to control tissues. ACh: Acetyl-choline; APO: apoptosis; FLOT2: flotillin 2; PTRPN: protein tyrosine phosphatase receptor type N; INV: invasion; ITPKB: inositol-triphosphate-3 kinase B; MCL1: myeloid cell differentiation protein 1; PROL: proliferation; SLC7A5: solute carrier family 7A5; WWOX: WW containing oxidoreductase.

miR-335 was also down-regulated in SCLC cell lines H69AR and H446DDP (66). Overexpression of *miR-335* inhibited migration of H69AR and H446DDP cells *in vitro* and their tumor growth *in vivo*, whereas its inhibition resulted in opposite effects (66). PARP1 was identified as a direct target of *miR-335* (66). Chemoradiosensitivity of SCLC cells was increased by down-regulation of *PARP1* and nuclear factor κB (66). Down-regulation of *miR-335* resulted in resistance to adriamycin, cisplatin and etoposide in SCLC cell lines H69AR and H446DDP (66). PARP1 detects single-strand DNA breaks and recruits other enzymes involved in DNA repair (67, 68).

miR-495. *miR-495* (Figure 1) was down-regulated in SCLC and inhibited chemoresistance by targeting endothelial tyrosine kinase/bone marrow X kinase (ETK/BMX) (69) and tetraspanin 12 (TSPN12) (70). Functional assays were performed in SCLC cell lines NCI-H446, NCI-H69 and their multidrug-resistant derivatives H446AR and H69AR (69, 70). *miR-495* was expressed at a lower level in SCLC compared to normal lung tissues (69, 70).

miR-495 inhibited apoptosis induced by chemotherapeutic agents such as adriamycin, cisplatin and etoposide by

targeting ETK/BMX (69). In nude mice, antagomirs directed against *miR-495* induced rapid growth of xenografts derived from H69 and H446 cells (69). Down-regulation of *miR-495* promoted proliferation, migration invasion and tumor growth of H446 and H69 SCLC *in vitro* and *in vivo* (69). ETK/BMX has been shown to mediate drug resistance in SCLC (71), to regulate the cytoskeleton and migration (72), and to up-regulate vascular endothelial growth factor (73). ETK/BMX has also been identified as a mediator of resistance in acute myeloid leukemia (74) and as a regulator of multiple tyrosine kinases in hormone-refractory prostate cancer (75).

miR-495 was also found to target TSPAN12, which is related to resistance to cisplatin and etoposide (60). TSPAN12 promoted proliferation, migration and tumor growth in drug-resistant SCLC cells H466AR and H69AR (60). TSPAN12 belongs to the tetraspanin family of transmembrane receptors characterized by four transmembrane domains and two extracellular loops (76). Tetraspanins are involved in signaling platforms by forming tetraspanin-enriched microdomains (77). Tetraspanins can mediate tumor-promoting but also metastasis-inhibitory processes (78-80).

miRs Up-regulated in SCLC With Activity in Preclinical *In Vitro* Systems

miR-25. *miR-25* (Figure 2) was up-regulated in SCLC cell lines and tissues (81). Down-regulation of *miR-25* induced cell-cycle arrest and inhibited invasive capability of H510 SCLC cells (81). Overexpression of *miR-25* reversed the effect of *miR-25* down-regulation in H510 cells (81). *miR-25* acted as an oncogene in SCLC cell lines (81). Cyclin E2 has been identified as a direct target of *miR-25*. These findings seem to be counterintuitive since cyclin E has been identified as a regulator of S-phase activity by binding to and activating cyclin-dependent kinase 2 and by phosphorylation of pocket proteins initiating a cascade of events that leads to the expression of S-phase-specific genes (82, 83). A role of cyclin E in DNA replication, control of genomic stability and regulation of the centrosome cycle has also been reported (82, 83). Cyclin E2 is aberrantly expressed in many types of tumors and is increased in cancer-derived cell lines (84). Overexpression of cyclin E in transgenic mice was shown to induce cancer by acting as a dominant oncogene (85). Due to its role in proliferation and apoptosis, cyclin E2 may be an important target for cancer therapy (86). However, it was shown that cyclin E is dispensable for the development of higher eukaryotes and for the division of eukaryotic cells (85). In any case, down-regulation of cyclin E2 in SCLC as reported in (81) might activate a novel tumor-promoting pathway which has to be resolved in further detail.

miR-134. In H69 SCLC cells, *miR-134* (Figure 2) promoted growth, inhibited apoptosis and activated the extracellular signal-regulated kinase 1/2 (ERK1/2) pathway (87). WW domain-containing oxidoreductase (WWOX) has been identified as a direct target of *miR-134* (87). WWOX has two WW domains responsible for protein–protein interactions and a short dehydrogenase/reductase domain which catalyses conversion of low-molecular-weight ligands, most likely steroids (88). Ectopic expression of WWOX inhibited anchorage-dependent growth of MDA-MB-435 and T47D breast cancer cells and attenuates tumorigenicity of MDA-MB-435 cells *in vivo* (89). In lung cancer, WWOX gene restoration prevented tumor growth *in vitro* and *in vivo* (89). WWOX localizes to the Golgi apparatus and behaves as a tumor suppressor (90). WWOX is frequently down-regulated in human tumors (91, 92).

miR-375. *miR-375* (Figure 2) was found to be up-regulated in lung adenocarcinoma and SCLC, and down-regulated in lung squamous cell carcinoma (93). *miR-375* promoted proliferation of NCI-H82 SCLC cells (93). Inositol-triphosphate-3 kinase B (ITPKB) was identified as a target of *miR-375* (93). ITPKB regulates inositol phosphate metabolism by phosphorylation of second messenger

inositol-1,4,5 triphosphate (94, 95). ITPKB is associated with the Ca signaling pathway and is enriched at actin filaments and invaginations of the nuclear envelope (96). ITPKB also regulates immune functions and is required for B- and T-cell development (96). The role of *miR-375* and down-regulation of *ITPKB* in SCLC remains to be investigated in further detail.

miRs Down-regulated in SCLC With Activity in Preclinical *In Vitro* Systems

miR-26a. Low level expression of *miR-26a* (Figure 2) was detected in SCLC cell lines NCI-H196, NCI-H466 and NCI-H1688 in comparison to MRC5 non-transformed control cells (97). Transfection of these cell lines with a *miR-26a* mimic suppressed proliferation, migration and colony formation (97). Myeloid cell leukemia protein 1 (MCL1) has been identified as a target of *miR-26a* (97). MCL1 is a member of the BCL2 family and plays a role in inhibition of apoptosis induced by tumor necrosis factor-related apoptosis-inducing ligand (98, 99). Inhibition of MCL1 with small molecules has been pursued in several types of cancer, such as myeloma, follicular lymphoma and advanced SCLC in advanced clinical studies (100-102). MCL1 inhibition has been shown to be effective against a subset of SCLCs with high MCL1 and low B-cell lymphoma-extra large (BCL-XL) expression (101).

miR-126. *miR-126* (Figure 2) inhibited proliferation of H69 SCLC cells by causing delay in the G₁ phase of the cell-cycle (102). *miR-126* has been identified as a direct target of solute carrier family 7, member 5 (SLC7A5) (102). Suppression of SLC7A5 by RNAi delayed SCLC cells in the G₁ phase (103). SLC7A5 is part of cluster of differentiation 98 (CD98), and also referred to as large neutral amino acid transporter 1. The other component of CD98 is the CD98 heavy subunit protein encoded by the SCL3A2 gene. CD98 preferentially transports branched chain and aromatic amino acids and is overexpressed in several types of cancer (103-105). SCL7A5 can activate mechanistic target of rapamycin (mTOR), which phosphorylates p70S6 kinase and eukaryotic translation factor 4E-binding protein 1 (4EBP1), resulting in production of growth-promoting proteins (106). mTOR is activated in a large percentage of SCLCs and genetic alterations in the phosphatidylinositol-4,5-bisphosphate 3-kinase/AKT serine/threonine kinase 1/mTOR pathway have been identified in 36% of patients with SCLC (107).

miR-342. Protein tyrosine phosphatase receptor type N (PTPRN), also known as islet antigen 2 (IA-2), was identified as a target of *miR-342* (Figure 2) in SCLC cell lines NCI-H82 and NCI-345 (108). Down-regulation of *PTPRN* by siRNA suppressed SCLC growth as well as cell acetyl choline (ACh)

content and secretion (109). ACh rescued the inhibitory effects of *PTPRN* siRNA and of *miR-342* mimic on SCLC proliferation (109). ACh is an autocrine growth factor which facilitates SCLC growth (109). *PTPRN* is a transmembrane tyrosine receptor phosphatase and has an important role in secretion of hormones and neurotransmitters in SCLC cell lines, such as follicle-stimulating hormone, insulin, luteinizing hormone, dopamine, renin and norepinephrine (110, 111). *PTPRN* is highly expressed in tumors and cell lines of neuro-endocrine origin (112). It also has been identified as an auto-antigen that is reactive with sera of patients with insulin-dependent diabetes mellitus (112).

miR-485-5p. *miR-485-5p* (Figure 2) was reduced in SCLC tissues compared to adjacent normal tissues (113). *miR-485-5p* inhibited proliferation, migration and invasion of NCI-H466 and NCI-485-5p SCLC cell lines (113). Flotillin 2 (*FLOT2*) has been identified as a target of *miR-485-5p* (113). *FLOT2* was found to be up-regulated in SCLC tissues and correlated with worse prognosis (113). *FLOT1* and -2 are lipid-raft marker proteins which assemble into heterotetramers, forming molecular scaffolds to regulate clustering at the plasma membrane (114, 115). They are involved in signal transduction, nerve regeneration, endocytosis and lymphocyte activation (114, 115). Up-regulation of *FLOT2* is related to lymph node metastasis and poor prognosis in patients with solid tumors (116).

Dysregulated miRs With Activity in Preclinical In Vivo Models of SCLC

Up-regulated miRs

miR-665. Inhibition of *miR-665* (Figure 3) attenuated proliferation, invasion and migration of NCI-H446 SCLC cells (117). *In vivo*, inhibition of *miR-665* led to attenuation of tumor growth (117). Lethal giant larvae protein homolog-1 (*LLGL1*) was identified as a target of *miR-665* (117). *LLGL1* is part of the cytoskeletal network and is associated with non-muscle myosin II heavy chain (117). Overexpression of *LLGL1* inhibited proliferation and migration, and increased cellular adhesion and apoptosis (118, 119). Loss of *LLGL1* reduced cellular adhesion and dissemination in colorectal cancer, melanoma and gastric cancer; its reduced expression has been noted in lung squamous cell carcinoma (120-123).

Down-regulated miRs

miR-216a-5p. *miR-216a-5p* (Figure 3) reduced proliferation and migration of H69 SCLC cells (124). *miR-216a-5p* targeted *BCL2* and modulated *BCL2*-like protein (*BAX*) and *BCL2* antagonist of cell death (*BAD*) (124). *In vivo* inhibition of *miR-216a-5p* promoted tumor growth of H69-derived xenografts in mice, whereas a *miR-216* mimic inhibited it (124). *BCL2* is an anti-apoptotic protein which

is expressed in SCLC (125, 126). *BCL2* inhibitor venetoclax was shown to be active in preclinical SCLC-related *in vitro* and *in vivo* systems with high *BCL2* expression (127). Venetoclax is approved for chronic lymphocytic leukemia and small lymphocytic leukemia, and is also part of a combination therapy for acute myeloid leukemia (128). Currently clinical studies of treatment of relapsed or refractory SCLC with oral venetoclax in combination with irinotecan and venetoclax in combination with atezolizumab are underway (NCI04543916).

miR-335. Investigations into the role of *miR-335* (Figure 3) were performed with SCLC cell lines SBC-3 and SBC-5. The latter gives rise to bone metastasis in immuno-deficient mouse models, SBC-3 does not. Reduced expression of *miR-335* in SBC-5 in comparison to SBC-3 cells was observed (129). Overexpression of *miR-335* in transfected SBC-5 cells reduced proliferation, migration and colony formation. Skeletal lesions from *miR-335*-transfected SBC-5 cells were not observed in immunodeficient mice (129). Insulin-like growth factor receptor 1 (*IGF-1R*) and osteoblast receptor activator of nuclear κ B ligand (*RANKL*) were identified as targets for *miR-335* (129). *IGF-1R* promotes proliferation, invasion, migration and inhibits apoptosis of tumor cells (130). *IGF-1R* knock-out mice exhibit reduced bone metastasis of breast cancer xenografts (131). Prerequisite for osteolytic metastases is the activation of osteoclasts. Osteoblasts secrete *RANKL* which interacts with osteoclast precursors displaying *RANK* receptor on their surface, resulting in their maturation into functional osteoclasts. Osteoblasts also produce osteoprotegerin, a soluble decoy receptor which can block *RANK/RANKL* signaling (132-134). *miR-335* inhibits *IGF-1R* and *RANKL*, two validated mediators of bone metastasis.

miR-450. Down-regulation of *miR-450* (Figure 3) correlated with reduced survival in patients with SCLC (135). *miR-450* inhibited proliferation and invasion of H510A SCLC cells and growth as xenografts implanted into immunodeficient mice (135). Interferon regulatory factor 2 (*IRF2*) was identified as a target of *miR-450* (135). Overexpression of *IRF2* in H510A cells abrogated the inhibitory effects of *miR-450* (136). *IRF2* is a member of *IRF* protein family which possess an *N*-terminal DNA binding domain characterized by five well-conserved tryptophan-rich repeats recognizing IFN-stimulated response elements and a *C*-terminal region which mediates interactions with family members, transcription factors and co-factors conferring specific activities on each *IRF* (136, 137). *IRF2* acts as an oncogene and is involved in regulation of histone 4 gene transcription (138, 139). Overexpression of *IRF2* promote the growth of pancreatic cancer cells (140). In colorectal cancer, *IRF2* has been identified as a driver of immune suppression and immune therapy resistance (141).

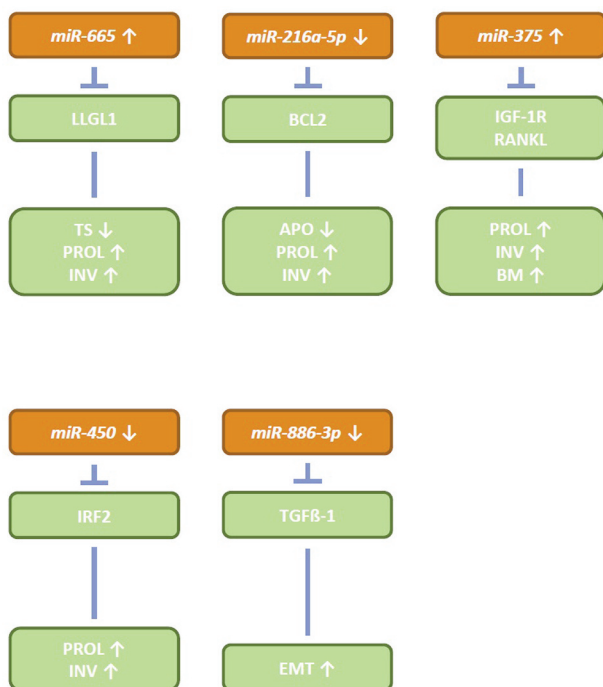


Figure 3. miRNAs Up- and down-regulated with activity in preclinical small-cell lung cancer-related *in vivo* models. Upward arrows: Up-regulated miRNAs; downward arrows: down-regulated miRNAs. APO: Apoptosis; BCL2: BCL2 apoptosis regulator; BM: bone metastasis; EMT: epithelial–mesenchymal transition; IGF-1R: insulin-like growth factor receptor 1; INV: invasion; IRF2: interferon regulatory factor 2; LLGL1: lethal giant larvae protein homolog 1; PROL: proliferation; RANK: receptor activator of NFκB ligand; RANKL: RANK ligand; TGFβ1: transforming growth factor β1; TS: tumor suppressor.

miR-886-3p. *miR-886-3p* (Figure 3) was down-regulated in SCLC (141). In NCI-H446 and NCI-H1688, overexpression of *miR-886-3p* induced mesenchymal–epithelial transition, a change in cellular phenotype from spindle shape to round shape (141). Transforming growth factor β1 (TGFβ1) has been identified as a target of *miR-886-3p* (141). *miR-886-3p*-mediated mesenchymal epithelial transition was induced by suppression of TGFβ (141). Intratumoral injection of a *miR-886-3p* conjugate to cholesterol resulted in necrosis of tumor tissue and suppression of intramuscular invasion (141). Suppression of lung cancer xenograft growth of NCI-H446 cells was observed after systemic delivery of a cholesterol conjugated *miR-886-3p* mimic after tail vein injection of tumor cells. Targeting EMT by either inhibition of TGFβ or by administration of *miR-886-3p* may be a concept for treatment of SCLC which has to be validated in further detail. EMT is a basic principle of tumor progression (142–144).

Conclusion

We identified miRNAs which affect chemoresistance and radioresistance, as well as *in vitro* and *in vivo* properties of SCLC cell lines. Up-regulated miRNAs are candidates for inhibition or reconstitution of the corresponding targets. Down-regulated miRNAs are candidates for reconstitution therapy or inhibition of the corresponding targets with small molecules or antibody-related entities.

Up-regulated miRNAs can be inhibited with miR antagonists, which are single-stranded RNAs composed of 12–25 nucleotides complementary to the corresponding mRNA or with RNA sponges (145, 146). The latter are composed of multiple miR-binding sites competing with binding of miRNAs to corresponding mRNA (145, 146). In the case of down-regulated miRNAs, reconstitution therapy is the indicated therapeutic intervention (147, 148) or re-expression of the corresponding targets, an approach which faces druggability issues due to nonspecific interactions.

Eight down-regulated miRNAs were found to mediate chemo/radioresistance (Figure 1). They are candidates for reconstitution therapy. PARP1 (*miR-335*) can be inhibited by several approved small molecules and is a validated target (35, 36). ETK (BMX) (*miR-495*), TSPAN12 (*miR-495*) and KIR2.1 (*miR-7*) are druggable with small molecules or antibody-derived entities. However, the role of the identified miRNAs in resistance of relapsed SCLC needs to be validated in more detail.

Three up-regulated and four down-regulated miRNAs affecting proliferation, invasion and apoptosis of SCLC cell lines *in vitro* were identified (Figure 2). MCL1, which is targeted by *miR-26a*, seems to be a promising target. MCL1 inhibition has been shown to be effective in a subset of preclinical SCLC-related *in vitro* models with high MCL1 and low BCL-xL expression (101). PTPRN (*miR-342*) and SCL7A5 (*miR-126*) are druggable targets and the corresponding miRNAs are candidates for miR-inhibitory agents. However, more target validation experiments are necessary to resolve the relevance of the latter targets.

Furthermore, one up-regulated and four down-regulated miRNAs with efficacy in preclinical SCLC-related *in vivo* models were identified (Figure 3). The down-regulated miRNAs are candidates for substitution therapy. BCL2 (*miR-216-5p*) is inhibited by venetoclax and it has been shown that venetoclax is effective in preclinical *in vivo* models with high BCL2 expression (127). *miR-335* targets IGF-1R and RANKL, which mediate proliferation, invasion and bone metastasis of SCLC and both represent druggable targets (130, 132). *miR-886-3p* inhibits TGFβ1, a possible target for interfering with EMT (143, 144). For these miRNAs and corresponding targets, more target validation experiments in non-small-cell lung carcinoma-related systems are necessary in order to substantiate their role in SCLC.

Regarding miR-based therapy, many technical hurdles which are not discussed in detail here have been identified. Issues are targeting of miRs to tumor cells, efficacy of intracellular escape, removal by the reticulo-endothelial system, excretion by the kidneys, pharmaco-kinetic and pharmaco-dynamic issues, immunogenicity, toxicity and cytokine-release syndrome (149-154). Recently, the field has experienced several set-backs, mainly due to toxicity issues (155). It remains to be seen whether miRs are tools for further target identification and whether miR-based therapy is a viable strategy for treatment of SCLC.

Conflicts of Interest

AN is and UHW was an employee of Roche.

Authors' Contributions

AN and UHW jointly designed and prepared the article.

References

- Gazdar AF, Bunn PA and Minna JD: Small-cell lung cancer: what we know, what we need to know and the path forward. *Nat Rev Cancer* 17(12): 725-737, 2017. PMID: 29077690. DOI: 10.1038/nrc.2017.87
- Pietanza MC, Byers LA, Minna JD and Rudin CM: Small cell lung cancer: will recent progress lead to improved outcomes? *Clin Cancer Res* 21(10): 2244-2255, 2015. PMID: 25979931. DOI: 10.1158/1078-0432.CCR-14-2958
- Farago AF, Drapkin BJ, Lopez-Vilarino de Ramos JA, Galmarini CM, Núñez R, Kahatt C and Paz-Ares L: ATLANTIS: a Phase III study of lurbinectedin/doxorubicin *versus* topotecan or cyclophosphamide/doxorubicin/vincristine in patients with small-cell lung cancer who have failed one prior platinum-containing line. *Future Oncol* 15(3): 231-239, 2019. PMID: 30362375. DOI: 10.2217/fon-2018-0597
- Gazdar AF and Minna JD: Small cell lung cancers made from scratch. *J Exp Med* 216(3): 476-478, 2019. PMID: 30760489. DOI: 10.1084/jem.20182216
- Iams WT, Porter J and Horn L: Immunotherapeutic approaches for small-cell lung cancer. *Nat Rev Clin Oncol* 17(5): 300-312, 2020. PMID: 32055013. DOI: 10.1038/s41571-019-0316-z
- Rudin CM, Poirier JT, Byers LA, Dive C, Dowlati A, George J, Heymach JV, Johnson JE, Lehman JM, MacPherson D, Massion PP, Minna JD, Oliver TG, Quaranta V, Sage J, Thomas RK, Vakoc CR and Gazdar AF: Molecular subtypes of small cell lung cancer: a synthesis of human and mouse model data. *Nat Rev Cancer* 19(5): 289-297, 2019. PMID: 30926931. DOI: 10.1038/s41568-019-0133-9
- Dawkins JBN and Webster RM: The small-cell lung cancer drug market. *Nat Rev Drug Discov* 19(8): 507-508, 2020. PMID: 32235872. DOI: 10.1038/d41573-020-00057-5
- Bartel DP: Metazoan MicroRNAs. *Cell* 173(1): 20-51, 2018. PMID: 29570994. DOI: 10.1016/j.cell.2018.03.006
- Li Z and Rana TM: Therapeutic targeting of microRNAs: current status and future challenges. *Nat Rev Drug Discov* 13(8): 622-638, 2014. PMID: 25011539. DOI: 10.1038/nrd4359
- Rupaimoole R and Slack FJ: MicroRNA therapeutics: towards a new era for the management of cancer and other diseases. *Nat Rev Drug Discov* 16(3): 203-222, 2017. PMID: 28209991. DOI: 10.1038/nrd.2016.246
- Peter ME: Targeting of mRNAs by multiple miRNAs: the next step. *Oncogene* 29(15): 2161-2164, 2010. PMID: 20190803. DOI: 10.1038/ncr.2010.59
- Garzon R, Calin GA and Croce CM: MicroRNAs in Cancer. *Annu Rev Med* 60: 167-179, 2009. PMID: 19630570. DOI: 10.1146/annurev.med.59.053006.104707
- Calin GA, Cimmino A, Fabbri M, Ferracin M, Wojcik SE, Shimizu M, Taccioli C, Zanesi N, Garzon R, Aqeilan RI, Alder H, Volinia S, Rassenti L, Liu X, Liu CG, Kipps TJ, Negrini M and Croce CM: MiR-15a and miR-16-1 cluster functions in human leukemia. *Proc Natl Acad Sci USA* 105(13): 5166-5171, 2008. PMID: 18362358. DOI: 10.1073/pnas.0800121105
- Calin GA, Dumitru CD, Shimizu M, Bichi R, Zupo S, Noch E, Alder H, Rattan S, Keating M, Rai K, Rassenti L, Kipps T, Negrini M, Bullrich F and Croce CM: Frequent deletions and down-regulation of micro-RNA genes miR15 and miR16 at 13q14 in chronic lymphocytic leukemia. *Proc Natl Acad Sci USA* 99(24): 15524-15529, 2002. PMID: 12434020. DOI: 10.1073/pnas.242606799
- Callegari E, Elamin BK, Giannone F, Milazzo M, Altavilla G, Fornari F, Giacomelli L, D'Abundo L, Ferracin M, Bassi C, Zagatti B, Corrà F, Miotto E, Lupini L, Bolondi L, Gramantieri L, Croce CM, Sabbioni S and Negrini M: Liver tumorigenicity promoted by microRNA-221 in a mouse transgenic model. *Hepatology* 56(3): 1025-1033, 2012. PMID: 22473819. DOI: 10.1002/hep.25747
- Ali Syeda Z, Langden SSS, Munkhzul C, Lee M and Song SJ: Regulatory mechanism of MicroRNA expression in cancer. *Int J Mol Sci* 21(5): 1723, 2020. PMID: 32138313. DOI: 10.3390/ijms21051723
- Weidle UH, Schmid D, Birzele F and Brinkmann U: MicroRNAs involved in metastasis of hepatocellular carcinoma: Target candidates, functionality and efficacy in animal models and prognostic relevance. *Cancer Genomics Proteomics* 17(1): 1-21, 2020. PMID: 31882547. DOI: 10.21873/cgp.20163
- Weidle UH, Birzele F and Nopora A: Pancreatic ductal adenocarcinoma: MicroRNAs affecting tumor growth and metastasis in preclinical *in vivo* models. *Cancer Genomics Proteomics* 16(6): 451-464, 2019. PMID: 31659100. DOI: 10.21873/cgp.20149
- Weidle UH, Birzele F and Nopora A: MicroRNAs as potential targets for therapeutic intervention with metastasis of non-small cell lung cancer. *Cancer Genomics Proteomics* 16(2): 99-119, 2019. PMID: 30850362. DOI: 10.21873/cgp.20116
- Weidle UH, Dickopf S, Hintermair C, Kollmorgen G, Birzele F and Brinkmann U: The role of micro RNAs in breast cancer metastasis: Preclinical validation and potential therapeutic targets. *Cancer Genomics Proteomics* 15(1): 17-39, 2018. PMID: 29275360. DOI: 10.21873/cgp.20062
- Weidle UH, Epp A, Birzele F and Brinkmann U: The functional role of prostate cancer metastasis-related Micro-RNAs. *Cancer Genomics Proteomics* 16(1): 1-19, 2019. PMID: 30587496. DOI: 10.21873/cgp.20108
- Liu H, Wu X, Huang J, Peng J and Guo L: miR-7 modulates chemoresistance of small cell lung cancer by repressing MRP1/ABCC1. *Int J Exp Pathol* 96(4): 240-247, 2015. PMID: 26108539. DOI: 10.1111/iep.12131

- 23 Liu H, Huang J, Peng J, Wu X, Zhang Y, Zhu W and Guo L: Upregulation of the inwardly rectifying potassium channel Kir2.1 (KCNJ2) modulates multidrug resistance of small-cell lung cancer under the regulation of miR-7 and the Ras/MAPK pathway. *Mol Cancer* 14: 59, 2015. PMID: 25880778. DOI: 10.1186/s12943-015-0298-0
- 24 Lai J, Yang H, Zhu Y, Ruan M, Huang Y and Zhang Q: MiR-7-5p-mediated downregulation of PARP1 impacts DNA homologous recombination repair and resistance to doxorubicin in small cell lung cancer. *BMC Cancer* 19(1): 602, 2019. PMID: 31215481. DOI: 10.1186/s12885-019-5798-7
- 25 Zaman GJ, Versantvoort CH, Smit JJ, Eijndems EW, de Haas M, Smith AJ, Broxterman HJ, Mulder NH, de Vries EG and Baas F: Analysis of the expression of MRP, the gene for a new putative transmembrane drug transporter, in human multidrug resistant lung cancer cell lines. *Cancer Res* 53(8): 1747-1750, 1993. PMID: 8467491.
- 26 Robey RW, Pluchino KM, Hall MD, Fojo AT, Bates SE and Gottesman MM: Revisiting the role of ABC transporters in multidrug-resistant cancer. *Nat Rev Cancer* 18(7): 452-464, 2018. PMID: 29643473. DOI: 10.1038/s41568-018-0005-8
- 27 Hsia TC, Lin CC, Wang JJ, Ho ST and Kao A: Relationship between chemotherapy response of small cell lung cancer and P-glycoprotein or multidrug resistance-related protein expression. *Lung* 180(3): 173-179, 2002. PMID: 12177731. DOI: 10.1007/s004080000091
- 28 Kuo TH, Liu FY, Chuang CY, Wu HS, Wang JJ and Kao A: To predict response chemotherapy using technetium-99m tetrofosmin chest images in patients with untreated small cell lung cancer and compare with p-glycoprotein, multidrug resistance related protein-1, and lung resistance-related protein expression. *Nucl Med Biol* 30(6): 627-632, 2003. PMID: 12900288. DOI: 10.1016/s0969-8051(03)00058-1
- 29 Mirski SE, Gerlach JH and Cole SP: Multidrug resistance in a human small cell lung cancer cell line selected in adriamycin. *Cancer Res* 47(10): 2594-2598, 1987. PMID: 2436751.
- 30 Raab-Graham KF, Radeke CM and Vandenberg CA: Molecular cloning and expression of a human heart inward rectifier potassium channel. *Neuroreport* 5(18): 2501-2505, 1994. PMID: 7696590. DOI: 10.1097/00001756-199412000-00024
- 31 Hibino H, Inanobe A, Furutani K, Murakami S, Findlay I and Kurachi Y: Inwardly rectifying potassium channels: their structure, function, and physiological roles. *Physiol Rev* 90(1): 291-366, 2010. PMID: 20086079. DOI: 10.1152/physrev.00021.2009
- 32 Pancrazio JJ, Viglione MP, Tabbara IA and Kim YI: Voltage-dependent ion channels in small-cell lung cancer cells. *Cancer Res* 49(21): 5901-5906, 1989. PMID: 2477149.
- 33 Giovannardi S, Forlani G, Balestrini M, Bossi E, Tonini R, Sturani E, Peres A and Zippel R: Modulation of the inward rectifier potassium channel IRK1 by the Ras signaling pathway. *J Biol Chem* 277(14): 12158-12163, 2002. PMID: 11809752. DOI: 10.1074/jbc.M110466200
- 34 Tsoukalas N, Aravantinou-Fatorou E, Baxevanos P, Tolia M, Tsapakidis K, Galanopoulos M, Lontos M and Kyrgias G: Advanced small cell lung cancer (SCLC): new challenges and new expectations. *Ann Transl Med* 6(8): 145, 2018. PMID: 29862234. DOI: 10.21037/atm.2018.03.31
- 35 Ray Chaudhuri A and Nussenzweig A: The multifaceted roles of PARP1 in DNA repair and chromatin remodelling. *Nat Rev Mol Cell Biol* 18(10): 610-621, 2017. PMID: 28676700. DOI: 10.1038/nrm.2017.53
- 36 Lord CJ, Tutt AN and Ashworth A: Synthetic lethality and cancer therapy: lessons learned from the development of PARP inhibitors. *Annu Rev Med* 66: 455-470, 2015. PMID: 25341009. DOI: 10.1146/annurev-med-050913-022545
- 37 Lord CJ and Ashworth A: PARP inhibitors: Synthetic lethality in the clinic. *Science* 355(6330): 1152-1158, 2017. PMID: 28302823. DOI: 10.1126/science.aam7344
- 38 Slade D: PARP and PARG inhibitors in cancer treatment. *Genes Dev* 34(5-6): 360-394, 2020. PMID: 32029455. DOI: 10.1101/gad.334516.119
- 39 Jiang W, Han X, Wang J, Wang L, Xu Z, Wei Q, Zhang W and Wang H: miR-22 enhances the radiosensitivity of small-cell lung cancer by targeting the WRNIP1. *J Cell Biochem* 120(10): 17650-17661, 2019. PMID: 31190355. DOI: 10.1002/jcb.29032
- 40 Kawabe Yi, Branzei D, Hayashi T, Suzuki H, Masuko T, Onoda F, Heo SJ, Ikeda H, Shimamoto A, Furuichi Y, Seki M and Enomoto T: A novel protein interacts with the Werner's syndrome gene product physically and functionally. *J Biol Chem* 276(23): 20364-20369, 2001. PMID: 11301316. DOI: 10.1074/jbc.C100035200
- 41 Kawabe Y, Seki M, Yoshimura A, Nishino K, Hayashi T, Takeuchi T, Iguchi S, Kusa Y, Ohtsuki M, Tsuyama T, Imamura O, Matsumoto T, Furuichi Y, Tada S and Enomoto T: Analyses of the interaction of WRNIP1 with Werner syndrome protein (WRN) *in vitro* and in the cell. *DNA Repair (Amst)* 5(7): 816-828, 2006. PMID: 16769258. DOI: 10.1016/j.dnarep.2006.04.006
- 42 Leuzzi G, Marabitti V, Pichierri P and Franchitto A: WRNIP1 protects stalled forks from degradation and promotes fork restart after replication stress. *EMBO J* 35(13): 1437-1451, 2016. PMID: 27242363. DOI: 10.15252/embj.201593265
- 43 Levy JMM, Towers CG and Thorburn A: Targeting autophagy in cancer. *Nat Rev Cancer* 17(9): 528-542, 2017. PMID: 28751651. DOI: 10.1038/nrc.2017.53
- 44 Yang ZJ, Chee CE, Huang S and Sinicrope FA: The role of autophagy in cancer: therapeutic implications. *Mol Cancer Ther* 10(9): 1533-1541, 2011. PMID: 21878654. DOI: 10.1158/1535-7163.MCT-11-0047
- 45 Pan B, Chen Y, Song H, Xu Y, Wang R and Chen L: Mir-24-3p downregulation contributes to VP16-DDP resistance in small-cell lung cancer by targeting ATG4A. *Oncotarget* 6(1): 317-331, 2015. PMID: 25426560. DOI: 10.18632/oncotarget.2787
- 46 Amaravadi RK, Lippincott-Schwartz J, Yin XM, Weiss WA, Takebe N, Timmer W, DiPaola RS, Lotze MT and White E: Principles and current strategies for targeting autophagy for cancer treatment. *Clin Cancer Res* 17(4): 654-666, 2011. PMID: 21325294. DOI: 10.1158/1078-0432.CCR-10-2634
- 47 Xiao F, Bai Y, Chen Z, Li Y, Luo L, Huang J, Yang J, Liao H and Guo L: Downregulation of HOXA1 gene affects small cell lung cancer cell survival and chemoresistance under the regulation of miR-100. *Eur J Cancer* 50(8): 1541-1554, 2014. PMID: 24559685. DOI: 10.1016/j.ejca.2014.01.024
- 48 Yuan C, Zhu X, Han Y, Song C, Liu C, Lu S, Zhang M, Yu F, Peng Z and Zhou C: Elevated HOXA1 expression correlates with accelerated tumor cell proliferation and poor prognosis in gastric cancer partly via cyclin D1. *J Exp Clin Cancer Res* 35: 15, 2016. PMID: 26791264. DOI: 10.1186/s13046-016-0294-2
- 49 Wang H, Liu G, Shen D, Ye H, Huang J, Jiao L and Sun Y: HOXA1 enhances the cell proliferation, invasion and metastasis

- of prostate cancer cells. *Oncol Rep* 34(3): 1203-1210, 2015. PMID: 26135141. DOI: 10.3892/or.2015.4085
- 50 Zha TZ, Hu BS, Yu HF, Tan YF, Zhang Y and Zhang K: Overexpression of HOXA1 correlates with poor prognosis in patients with hepatocellular carcinoma. *Tumour Biol* 33(6): 2125-2134, 2012. PMID: 22864671. DOI: 10.1007/s13277-012-0472-6
- 51 Yang H, Luo J, Liu Z, Zhou R and Luo H: MicroRNA-138 regulates DNA damage response in small cell lung cancer cells by directly targeting H2AX. *Cancer Invest* 33(4): 126-136, 2015. PMID: 25699650. DOI: 10.3109/07357907.2015.1006329
- 52 Matthaios D, Hountis P, Karakitsos P, Bouros D and Kakolyris S: H2AX a promising biomarker for lung cancer: a review. *Cancer Invest* 31(9): 582-599, 2013. PMID: 24164298. DOI: 10.3109/07357907.2013.849721
- 53 Kuo LJ and Yang LX: Gamma-H2AX - a novel biomarker for DNA double-strand breaks. *In Vivo* 22(3): 305-309, 2008. PMID: 18610740.
- 54 Podhorecka M, Skladanowski A and Bozko P: H2AX phosphorylation: Its role in DNA damage response and cancer therapy. *J Nucleic Acids* 2010: 920161, 2010. PMID: 20811597. DOI: 10.4061/2010/920161
- 55 Ivashkevich A, Redon CE, Nakamura AJ, Martin RF and Martin OA: Use of the γ -H2AX assay to monitor DNA damage and repair in translational cancer research. *Cancer Lett* 327(1-2): 123-133, 2012. PMID: 22198208. DOI: 10.1016/j.canlet.2011.12.025
- 56 Fang S, Zeng X, Zhu W, Tang R, Chao Y and Guo L: Zinc finger E-box-binding homeobox 2 (ZEB2) regulated by miR-200b contributes to multi-drug resistance of small cell lung cancer. *Exp Mol Pathol* 96(3): 438-444, 2014. PMID: 24769353. DOI: 10.1016/j.yexmp.2014.04.008
- 57 Bürglin TR and Affolter M: Homeodomain proteins: an update. *Chromosoma* 125(3): 497-521, 2016. PMID: 26464018. DOI: 10.1007/s00412-015-0543-8
- 58 Hegarty SV, Sullivan AM and O'Keeffe GW: Zeb2: A multifunctional regulator of nervous system development. *Prog Neurobiol* 132: 81-95, 2015. PMID: 26193487. DOI: 10.1016/j.pneurobio.2015.07.001
- 59 Epifanova E, Babaev A, Newman AG and Tarabykin V: Role of Zeb2/Sip1 in neuronal development. *Brain Res* 1705: 24-31, 2019. PMID: 30266271. DOI: 10.1016/j.brainres.2018.09.034
- 60 Fardi M, Alivand M, Baradaran B, Farshdousti Hagh M and Solali S: The crucial role of ZEB2: From development to epithelial-to-mesenchymal transition and cancer complexity. *J Cell Physiol* : , 2019. PMID: 30773635. DOI: 10.1002/jcp.28277
- 61 Tang R, Lei Y, Hu B, Yang J, Fang S, Wang Q, Li M and Guo L: WW domain binding protein 5 induces multidrug resistance of small cell lung cancer under the regulation of miR-335 through the Hippo pathway. *Br J Cancer* 115(2): 243-251, 2016. PMID: 27336605. DOI: 10.1038/bjc.2016.186
- 62 Sudol M, Chen HI, Bougeret C, Einbond A and Bork P: Characterization of a novel protein-binding module—the WW domain. *FEBS Lett* 369(1): 67-71, 1995. PMID: 7641887. DOI: 10.1016/0014-5793(95)00550-s
- 63 Zanconato F, Cordenonsi M and Piccolo S: YAP/TAZ at the roots of cancer. *Cancer Cell* 29(6): 783-803, 2016. PMID: 27300434. DOI: 10.1016/j.ccell.2016.05.005
- 64 Liu W, Wu J, Xiao L, Bai Y, Qu A, Zheng Z and Yuan Z: Regulation of neuronal cell death by c-Abl-Hippo/MST2 signaling pathway. *PLoS One* 7(5): e36562, 2012. PMID: 22590567. DOI: 10.1371/journal.pone.0036562
- 65 Harvey KF, Zhang X and Thomas DM: The Hippo pathway and human cancer. *Nat Rev Cancer* 13(4): 246-257, 2013. PMID: 23467301. DOI: 10.1038/nrc3458
- 66 Luo Y, Tong L, Meng H, Zhu W, Guo L, Wei T and Zhang J: MiR-335 regulates the chemo-radioresistance of small cell lung cancer cells by targeting PARP-1. *Gene* 600: 9-15, 2017. PMID: 27871924. DOI: 10.1016/j.gene.2016.11.031
- 67 Sachdev E, Tabatabai R, Roy V, Rimel BJ and Mita MM: PARP Inhibition in Cancer: An Update on Clinical Development. *Target Oncol* 14(6): 657-679, 2019. PMID: 31625002. DOI: 10.1007/s11523-019-00680-2
- 68 O'Connor MJ: Targeting the DNA Damage Response in Cancer. *Mol Cell* 60(4): 547-560, 2015. PMID: 26590714. DOI: 10.1016/j.molcel.2015.10.040
- 69 Wei T, Zhu W, Fang S, Zeng X, Huang J, Yang J, Zhang J and Guo L: miR-495 promotes the chemoresistance of SCLC through the epithelial-mesenchymal transition via Etk/BMX. *Am J Cancer Res* 7(3): 628-646, 2017. PMID: 28401017.
- 70 Ye M, Wei T, Wang Q, Sun Y, Tang R, Guo L and Zhu W: TSPAN12 promotes chemoresistance and proliferation of SCLC under the regulation of miR-495. *Biochem Biophys Res Commun* 486(2): 349-356, 2017. PMID: 28302484. DOI: 10.1016/j.bbrc.2017.03.044
- 71 Guo L, Zhou Y, Sun Y and Zhang F: Non-receptor tyrosine kinase Etk regulation of drug resistance in small-cell lung cancer. *Eur J Cancer* 46(3): 636-641, 2010. PMID: 20004564. DOI: 10.1016/j.ejca.2009.11.009
- 72 Abassi YA, Rehn M, Ekman N, Alitalo K and Vuori K: p130Cas Couples the tyrosine kinase Bmx/Etk with regulation of the actin cytoskeleton and cell migration. *J Biol Chem* 278(37): 35636-35643, 2003. PMID: 12832404. DOI: 10.1074/jbc.M306438200
- 73 Chau CH, Chen KY, Deng HT, Kim KJ, Hosoya K, Terasaki T, Shih HM and Ann DK: Coordinating Etk/Bmx activation and VEGF upregulation to promote cell survival and proliferation. *Oncogene* 21(57): 8817-8829, 2002. PMID: 12483534. DOI: 10.1038/sj.onc.1206032
- 74 van Oosterwijk JG, Buelow DR, Drenberg CD, Vasilyeva A, Li L, Shi L, Wang YD, Finkelstein D, Shurtleff SA, Janke LJ, Pounds S, Rubnitz JE, Inaba H, Pabla N and Baker SD: Hypoxia-induced upregulation of BMX kinase mediates therapeutic resistance in acute myeloid leukemia. *J Clin Invest* 128(1): 369-380, 2018. PMID: 29227282. DOI: 10.1172/JCI91893
- 75 Chen S, Cai C, Sowalsky AG, Ye H, Ma F, Yuan X, Simon NI, Gray NS and Balk SP: BMX-mediated regulation of multiple tyrosine kinases contributes to castration resistance in prostate cancer. *Cancer Res* 78(18): 5203-5215, 2018. PMID: 30012673. DOI: 10.1158/0008-5472.CAN-17-3615
- 76 Serru V, Dessen P, Boucheix C and Rubinstein E: Sequence and expression of seven new tetraspans. *Biochim Biophys Acta* 1478(1): 159-163, 2000. PMID: 10719184. DOI: 10.1016/s0167-4838(00)00022-4
- 77 Yáñez-Mó M, Barreiro O, Gordon-Alonso M, Sala-Valdés M and Sánchez-Madrid F: Tetraspanin-enriched microdomains: a functional unit in cell plasma membranes. *Trends Cell Biol* 19(9): 434-446, 2009. PMID: 19709882. DOI: 10.1016/j.tcb.2009.06.004
- 78 Hemler ME: Tetraspanin proteins promote multiple cancer stages. *Nat Rev Cancer* 14(1): 49-60, 2014. PMID: 24505619. DOI: 10.1038/nrc3640

- 79 Zöller M: Tetraspanins: push and pull in suppressing and promoting metastasis. *Nat Rev Cancer* 9(1): 40-55, 2009. PMID: 19078974. DOI: 10.1038/nrc2543
- 80 Sala-Valdés M, Ailane N, Greco C, Rubinstein E and Boucheix C: Targeting tetraspanins in cancer. *Expert Opin Ther Targets* 16(10): 985-997, 2012. PMID: 22880813. DOI: 10.1517/14728222.2012.712688
- 81 Zhao Z, Liu J, Wang C, Wang Y, Jiang Y and Guo M: MicroRNA-25 regulates small cell lung cancer cell development and cell cycle through cyclin E2. *Int J Clin Exp Pathol* 7(11): 7726-7734, 2014. PMID: 25550809.
- 82 Hwang HC and Clurman BE: Cyclin E in normal and neoplastic cell cycles. *Oncogene* 24(17): 2776-2786, 2005. PMID: 15838514. DOI: 10.1038/sj.onc.1208613
- 83 Santamaria D and Ortega S: Cyclins and CDKS in development and cancer: lessons from genetically modified mice. *Front Biosci* 11: 1164-1188, 2006. PMID: 16146805. DOI: 10.2741/1871
- 84 Gudas JM, Payton M, Thukral S, Chen E, Bass M, Robinson MO and Coats S: Cyclin E2, a novel G1 cyclin that binds Cdk2 and is aberrantly expressed in human cancers. *Mol Cell Biol* 19(1): 612-622, 1999. PMID: 9858585. DOI: 10.1128/MCB.19.1.612
- 85 Möröy T and Geisen C: Cyclin E. *Int J Biochem Cell Biol* 36(8): 1424-1439, 2004. PMID: 15147722. DOI: 10.1016/j.biocel.2003.12.005
- 86 Mazumder S, DuPree EL and Almasan A: A dual role of cyclin E in cell proliferation and apoptosis may provide a target for cancer therapy. *Curr Cancer Drug Targets* 4(1): 65-75, 2004. PMID: 14965268. DOI: 10.2174/1568009043481669
- 87 Chen T, Gao F, Feng S, Yang T and Chen M: MicroRNA-134 regulates lung cancer cell H69 growth and apoptosis by targeting WWOX gene and suppressing the ERK1/2 signaling pathway. *Biochem Biophys Res Commun* 464(3): 748-754, 2015. PMID: 26166818. DOI: 10.1016/j.bbrc.2015.07.021
- 88 Pospiech K, Pluciennik E and Bednarek AK: WWOX Tumor suppressor gene in breast cancer, a historical perspective and future directions. *Front Oncol* 8: 345, 2018. PMID: 30211123. DOI: 10.3389/fonc.2018.00345
- 89 Bednarek AK, Keck-Waggoner CL, Daniel RL, Laflin KJ, Bergsagel PL, Kiguchi K, Brenner AJ and Aldaz CM: WWOX, the FRA16D gene, behaves as a suppressor of tumor growth. *Cancer Res* 61(22): 8068-8073, 2001. PMID: 11719429.
- 90 Fabbri M, Iliopoulos D, Trapasso F, Aqeilan RI, Cimmino A, Zanasi N, Yendamuri S, Han SY, Amadori D, Huebner K and Croce CM: WWOX gene restoration prevents lung cancer growth *in vitro* and *in vivo*. *Proc Natl Acad Sci USA* 102(43): 15611-15616, 2005. PMID: 16223882. DOI: 10.1073/pnas.0505485102
- 91 Schrock MS and Huebner K: WWOX: a fragile tumor suppressor. *Exp Biol Med (Maywood)* 240(3): 296-304, 2015. PMID: 25538133. DOI: 10.1177/1535370214561590
- 92 Baryła I, Styczeń-Binkowska E and Bednarek AK: Alteration of WWOX in human cancer: a clinical view. *Exp Biol Med (Maywood)* 240(3): 305-314, 2015. PMID: 25681467. DOI: 10.1177/1535370214561953
- 93 Jin Y, Liu Y, Zhang J, Huang W, Jiang H, Hou Y, Xu C, Zhai C, Gao X, Wang S, Wu Y, Zhu H and Lu S: The expression of miR-375 is associated with carcinogenesis in three subtypes of lung cancer. *PLoS One* 10(12): e0144187, 2015. PMID: 26642205. DOI: 10.1371/journal.pone.0144187
- 94 Chamberlain PP, Sandberg ML, Sauer K, Cooke MP, Lesley SA and Spraggon G: Structural insights into enzyme regulation for inositol 1,4,5-trisphosphate 3-kinase B. *Biochemistry* 44(44): 14486-14493, 2005. PMID: 16262249. DOI: 10.1021/bi051256q
- 95 Irvine RF and Schell MJ: Back in the water: the return of the inositol phosphates. *Nat Rev Mol Cell Biol* 2(5): 327-338, 2001. PMID: 11331907. DOI: 10.1038/35073015
- 96 Sauer K and Cooke MP: Regulation of immune cell development through soluble inositol-1,3,4,5-tetrakisphosphate. *Nat Rev Immunol* 10(4): 257-271, 2010. PMID: 20336153. DOI: 10.1038/nri2745
- 97 Yu JG, Ji CH and Shi MH: MicroRNA-26b suppresses tumorigenicity and promotes apoptosis in small cell lung cancer cells by targeting myeloid cell leukemia 1 protein. *Kaohsiung J Med Sci* 34(11): 593-605, 2018. PMID: 30392566. DOI: 10.1016/j.kjms.2018.06.005
- 98 Quinn BA, Dash R, Azab B, Sarkar S, Das SK, Kumar S, Oyesanya RA, Dasgupta S, Dent P, Grant S, Rahmani M, Curiel DT, Dmitriev I, Hedvat M, Wei J, Wu B, Stebbins JL, Reed JC, Pelliccia M, Sarkar D and Fisher PB: Targeting Mcl-1 for the therapy of cancer. *Expert Opin Investig Drugs* 20(10): 1397-1411, 2011. PMID: 21851287. DOI: 10.1517/13543784.2011.609167
- 99 Perciavalle RM and Opferman JT: Delving deeper: MCL-1's contributions to normal and cancer biology. *Trends Cell Biol* 23(1): 22-29, 2013. PMID: 23026029. DOI: 10.1016/j.tcb.2012.08.011
- 100 Xiang W, Yang CY and Bai L: MCL-1 inhibition in cancer treatment. *Onco Targets Ther* 11: 7301-7314, 2018. PMID: 30425521. DOI: 10.2147/OTT.S146228
- 101 Yasuda Y, Ozasa H, Kim YH, Yamazoe M, Ajimizu H, Yamamoto Funazo T, Nomizo T, Tsuji T, Yoshida H, Sakamori Y, Nakajima N, Menju T, Yoshizawa A, Date H and Hirai T: MCL1 inhibition is effective against a subset of small-cell lung cancer with high MCL1 and low BCL-X_L expression. *Cell Death Dis* 11(3): 177, 2020. PMID: 32152266. DOI: 10.1038/s41419-020-2379-2
- 102 Miko E, Margitai Z, Czimmerer Z, Várkonyi I, Dezsó B, Lányi A, Bacsó Z and Scholtz B: miR-126 inhibits proliferation of small cell lung cancer cells by targeting SLCTA5. *FEBS Lett* 585(8): 1191-1196, 2011. PMID: 21439283. DOI: 10.1016/j.febslet.2011.03.039
- 103 Salisbury TB and Arthur S: The regulation and function of the L-type amino acid transporter 1 (LAT1) in cancer. *Int J Mol Sci* 19(8): 2373, 2018. PMID: 30103560. DOI: 10.3390/ijms19082373
- 104 Häfliger P and Charles RP: The L-type amino acid transporter LAT1-An emerging target in cancer. *Int J Mol Sci* 20(10): 2428, 2019. PMID: 31100853. DOI: 10.3390/ijms20102428
- 105 Lu X: The role of large neutral amino acid transporter (LAT1) in cancer. *Curr Cancer Drug Targets* 19(11): 863-876, 2019. PMID: 31376820. DOI: 10.2174/1568009619666190802135714
- 106 Nicklin P, Bergman P, Zhang B, Triantafellow E, Wang H, Nyfeler B, Yang H, Hild M, Kung C, Wilson C, Myer VE, MacKeigan JP, Porter JA, Wang YK, Cantley LC, Finan PM and Murphy LO: Bidirectional transport of amino acids regulates mTOR and autophagy. *Cell* 136(3): 521-534, 2009. PMID: 19203585. DOI: 10.1016/j.cell.2008.11.044
- 107 Pardo OE, Arcaro A, Salerno G, Tetley TD, Valovka T, Gout I and Seckl MJ: Novel cross talk between MEK and S6K2 in FGF-2 induced proliferation of SCLC cells. *Oncogene* 20(52): 7658-7667, 2001. PMID: 11753643. DOI: 10.1038/sj.onc.1204994

- 108 Xu H, Cai T, Carmona GN, Abuhatzira L and Notkins AL: Small cell lung cancer growth is inhibited by miR-342 through its effect of the target gene IA-2. *J Transl Med* 14(1): 278, 2016. PMID: 27670444. DOI: 10.1186/s12967-016-1036-0
- 109 Song P, Sekhon HS, Jia Y, Keller JA, Blusztajn JK, Mark GP and Spindel ER: Acetylcholine is synthesized by and acts as an autocrine growth factor for small cell lung carcinoma. *Cancer Res* 63(1): 214-221, 2003. PMID: 12517800.
- 110 Xie H, Notkins AL and Lan MS: IA-2, a transmembrane protein tyrosine phosphatase, is expressed in human lung cancer cell lines with neuroendocrine phenotype. *Cancer Res* 56(12): 2742-2744, 1996. PMID: 8665506.
- 111 Lan MS, Lu J, Goto Y and Notkins AL: Molecular cloning and identification of a receptor-type protein tyrosine phosphatase, IA-2, from human insulinoma. *DNA Cell Biol* 13(5): 505-514, 1994. PMID: 8024693. DOI: 10.1089/dna.1994.13.505
- 112 Hu YF, Zhang HL, Cai T, Harashima S and Notkins AL: The IA-2 interactome. *Diabetologia* 48(12): 2576-2581, 2005. PMID: 16273344. DOI: 10.1007/s00125-005-0037-y
- 113 Gao F, Wu H, Wang R, Guo Y, Zhang Z, Wang T, Zhang G, Liu C and Liu J: MicroRNA-485-5p suppresses the proliferation, migration and invasion of small cell lung cancer cells by targeting flotillin-2. *Bioengineered* 10(1): 1-12, 2019. PMID: 30836864. DOI: 10.1080/21655979.2019.1586056
- 114 Gauthier-Rouvière C, Bodin S, Comunale F and Planchon D: Flotillin membrane domains in cancer. *Cancer Metastasis Rev* 39(2): 361-374, 2020. PMID: 32297092. DOI: 10.1007/s10555-020-09873-y
- 115 Liu XX, Liu WD, Wang L, Zhu B, Shi X, Peng ZX, Zhu HC, Liu XD, Zhong MZ, Xie D, Zeng MS and Ren CP: Roles of flotillins in tumors. *J Zhejiang Univ Sci B* 19(3): 171-182, 2018. PMID: 29504311. DOI: 10.1631/jzus.B1700102
- 116 Liu FT, Qu QG and Zhu ZM: Up-regulation of Flot-2 protein is related to lymph node metastasis and poor prognosis in human solid tumors. *Minerva Chir* 72(2): 146-156, 2017. PMID: 27981826. DOI: 10.23736/S0026-4733.16.07261-8
- 117 Liu R, Zhang L, Xu Z and Cui Y: [MiR-665 promotes the biological behavior of small cell lung cancer by targeting LLGL1]. *Zhongguo Fei Ai Za Zhi* 23(4): 223-232, 2020. PMID: 3222154. DOI: 10.3779/j.issn.1009-3419.2020.104.03
- 118 Tsuruga T, Nakagawa S, Watanabe M, Takizawa S, Matsumoto Y, Nagasaka K, Sone K, Hiraie H, Miyamoto Y, Hiraie O, Minaguchi T, Oda K, Yasugi T, Yano T and Taketani Y: Loss of Hg1-1 expression associates with lymph node metastasis in endometrial cancer. *Oncol Res* 16(9): 431-435, 2007. PMID: 18074678. DOI: 10.3727/000000007783980855
- 119 Song J, Peng XL, Ji MY, Ai MH, Zhang JX and Dong WG: Hg1-1 induces apoptosis in esophageal carcinoma cells both *in vitro* and *in vivo*. *World J Gastroenterol* 19(26): 4127-4136, 2013. PMID: 23864775. DOI: 10.3748/wjg.v19.i26.4127
- 120 Schimanski CC, Schmitz G, Kashyap A, Bosserhoff AK, Bataille F, Schäfer SC, Lehr HA, Berger MR, Galle PR, Strand S and Strand D: Reduced expression of Hg1-1, the human homologue of *Drosophila* tumour suppressor gene *lgl*, contributes to progression of colorectal cancer. *Oncogene* 24(19): 3100-3109, 2005. PMID: 15735678. DOI: 10.1038/sj.onc.1208520
- 121 Kuphal S, Wallner S, Schimanski CC, Bataille F, Hofer P, Strand S, Strand D and Bosserhoff AK: Expression of Hg1-1 is strongly reduced in malignant melanoma. *Oncogene* 25(1): 103-110, 2006. PMID: 16170365. DOI: 10.1038/sj.onc.1209008
- 122 Desuki A, Staib F, Gockel I, Moehler M, Lang H, Biesterfeld S, Maderer A, Galle PR, Berger MR and Schimanski CC: Loss of *LLGL1* expression correlates with diffuse gastric cancer and distant peritoneal metastases. *Can J Gastroenterol Hepatol* 2019: 2920493, 2019. PMID: 31058107. DOI: 10.1155/2019/2920493
- 123 Matsuzaki T, Takekoshi S, Toriumi K, Kitatani K, Nitou M, Imamura N, Ogura G, Masuda R, Nakamura N and Iwazaki M: Reduced expression of Hg1-1 contributes to the progression of lung squamous cell carcinoma. *Tokai J Exp Clin Med* 40(4): 169-177, 2015. PMID: 26662669.
- 124 Sun Y, Hu B, Wang Y, Li Z, Wu J, Yang Y, Wei Y, Peng X, Chen H, Chen R, Jiang P, Fang S, Yu Z and Guo L: miR-216a-5p inhibits malignant progression in small cell lung cancer: involvement of the Bcl-2 family proteins. *Cancer Manag Res* 10: 4735-4745, 2018. PMID: 30425570. DOI: 10.2147/CMAR.S178380
- 125 Ben-Ezra JM, Kornstein MJ, Grimes MM and Krystal G: Small cell carcinomas of the lung express the Bcl-2 protein. *Am J Pathol* 145(5): 1036-1040, 1994. PMID: 7977636.
- 126 Jiang SX, Sato Y, Kuwao S and Kameya T: Expression of bcl-2 oncogene protein is prevalent in small cell lung carcinomas. *J Pathol* 177(2): 135-138, 1995. PMID: 7490679. DOI: 10.1002/path.1711770206
- 127 Lochmann TL, Floros KV, Naseri M, Powell KM, Cook W, March RJ, Stein GT, Greninger P, Maves YK, Saunders LR, Dylla SJ, Costa C, Boikos SA, Levenson JD, Souers AJ, Krystal GW, Harada H, Benes CH and Faber AC: Venetoclax is effective in small-cell lung cancers with high BCL-2 expression. *Clin Cancer Res* 24(2): 360-369, 2018. PMID: 29118061. DOI: 10.1158/1078-0432.CCR-17-1606
- 128 Lochmann TL, Bouck YM and Faber AC: BCL-2 inhibition is a promising therapeutic strategy for small cell lung cancer. *Oncoscience* 5(7-8): 218-219, 2018. PMID: 30234143. DOI: 10.18632/oncoscience.455
- 129 Gong M, Ma J, Guillemette R, Zhou M, Yang Y, Yang Y, Hock JM and Yu X: miR-335 inhibits small cell lung cancer bone metastases via IGF-IR and RANKL pathways. *Mol Cancer Res* 12(1): 101-110, 2014. PMID: 23966614. DOI: 10.1158/1541-7786.MCR-13-0136
- 130 Riedemann J and Macaulay VM: IGF1R signalling and its inhibition. *Endocr Relat Cancer* 13 Suppl 1: S33-S43, 2006. PMID: 17259557. DOI: 10.1677/erc.1.01280
- 131 Hiraga T, Myoui A, Hashimoto N, Sasaki A, Hata K, Morita Y, Yoshikawa H, Rosen CJ, Mundy GR and Yoneda T: Bone-derived IGF mediates crosstalk between bone and breast cancer cells in bony metastases. *Cancer Res* 72(16): 4238-4249, 2012. PMID: 22738911. DOI: 10.1158/0008-5472.CAN-11-3061
- 132 Weidle UH, Birzele F, Kollmorgen G and Rieger R: Molecular mechanisms of bone metastasis. *Cancer Genomics Proteomics* 13(1): 1-12, 2016. PMID: 26708594.
- 133 Dougall WC: Molecular pathways: osteoclast-dependent and osteoclast-independent roles of the RANKL/RANK/OPG pathway in tumorigenesis and metastasis. *Clin Cancer Res* 18(2): 326-335, 2012. PMID: 22031096. DOI: 10.1158/1078-0432.CCR-10-2507
- 134 Yin JJ, Pollock CB and Kelly K: Mechanisms of cancer metastasis to the bone. *Cell Res* 15(1): 57-62, 2005. PMID: 15686629. DOI: 10.1038/sj.cr.7290266
- 135 Liu F, Yu X, Huang H, Chen X, Wang J, Zhang X and Lin Q: Upregulation of microRNA-450 inhibits the progression of lung cancer *in vitro* and *in vivo* by targeting interferon regulatory factor

2. *Int J Mol Med* 38(1): 283-290, 2016. PMID: 27246609. DOI: 10.3892/ijmm.2016.2612
- 136 Yanai H, Negishi H and Taniguchi T: The IRF family of transcription factors: Inception, impact and implications in oncogenesis. *Oncoimmunology* 1(8): 1376-1386, 2012. PMID: 23243601. DOI: 10.4161/onci.22475
- 137 Taniguchi T, Ogasawara K, Takaoka A and Tanaka N: IRF family of transcription factors as regulators of host defense. *Annu Rev Immunol* 19: 623-655, 2001. PMID: 11244049. DOI: 10.1146/annurev.immunol.19.1.623
- 138 Nguyen H, Mustafa A, Hiscott J and Lin R: Transcription factor IRF-2 exerts its oncogenic phenotype through the DNA binding/transcription repression domain. *Oncogene* 11(3): 537-544, 1995. PMID: 7630638.
- 139 Vaughan PS, van der Meijden CM, Aziz F, Harada H, Taniguchi T, van Wijnen AJ, Stein JL and Stein GS: Cell cycle regulation of histone H4 gene transcription requires the oncogenic factor IRF-2. *J Biol Chem* 273(1): 194-199, 1998. PMID: 9417064. DOI: 10.1074/jbc.273.1.194
- 140 Liao W, Overman MJ, Boutin AT, Shang X, Zhao D, Dey P, Li J, Wang G, Lan Z, Li J, Tang M, Jiang S, Ma X, Chen P, Katkhuda R, Korphaisarn K, Chakravarti D, Chang A, Spring DJ, Chang Q, Zhang J, Maru DM, Maeda DY, Zebala JA, Kopetz S, Wang YA and DePinho RA: KRAS-IRF2 axis drives immune suppression and immune therapy resistance in colorectal cancer. *Cancer Cell* 35(4): 559-572.e7, 2019. PMID: 30905761. DOI: 10.1016/j.ccell.2019.02.008
- 141 Shen J, Zhou W, Bi N, Song YM, Zhang FQ, Zhan QM and Wang LH: MicroRNA-886-3P functions as a tumor suppressor in small cell lung cancer. *Cancer Biol Ther* 19(12): 1185-1192, 2018. PMID: 30230945. DOI: 10.1080/15384047.2018.1491505
- 142 Santamaria PG, Moreno-Bueno G, Portillo F and Cano A: EMT: Present and future in clinical oncology. *Mol Oncol* 11(7): 718-738, 2017. PMID: 28590039. DOI: 10.1002/1878-0261.12091
- 143 Kalluri R and Weinberg RA: The basics of epithelial-mesenchymal transition. *J Clin Invest* 119(6): 1420-1428, 2009. PMID: 19487818. DOI: 10.1172/JCI39104
- 144 Liu H, Zhang X, Li J, Sun B, Qian H and Yin Z: The biological and clinical importance of epithelial-mesenchymal transition in circulating tumor cells. *J Cancer Res Clin Oncol* 141(2): 189-201, 2015. PMID: 24965746. DOI: 10.1007/s00432-014-1752-x
- 145 Nguyen DD and Chang S: Development of novel therapeutic agents by inhibition of oncogenic MicroRNAs. *Int J Mol Sci* 19(1): 65, 2017. PMID: 29280958. DOI: 10.3390/ijms19010065
- 146 Ling H, Fabbri M and Calin GA: MicroRNAs and other non-coding RNAs as targets for anticancer drug development. *Nat Rev Drug Discov* 12(11): 847-865, 2013. PMID: 24172333. DOI: 10.1038/nrd4140
- 147 Gambari R, Brognara E, Spandidos DA and Fabbri E: Targeting oncomiRNAs and mimicking tumor suppressor miRNAs: New trends in the development of miRNA therapeutic strategies in oncology (Review). *Int J Oncol* 49(1): 5-32, 2016. PMID: 27175518. DOI: 10.3892/ijo.2016.3503
- 148 Broderick JA and Zamore PD: MicroRNA therapeutics. *Gene Ther* 18(12): 1104-1110, 2011. PMID: 21525952. DOI: 10.1038/gt.2011.50
- 149 Juliano RL, Ming X and Nakagawa O: Cellular uptake and intracellular trafficking of antisense and siRNA oligonucleotides. *Bioconjug Chem* 23(2): 147-157, 2012. PMID: 21992697. DOI: 10.1021/bc200377d
- 150 Malek A, Merkel O, Fink L, Czubyko F, Kissel T and Aigner A: *In vivo* pharmacokinetics, tissue distribution and underlying mechanisms of various PEI(-PEG)/siRNA complexes. *Toxicol Appl Pharmacol* 236(1): 97-108, 2009. PMID: 19371615. DOI: 10.1016/j.taap.2009.01.014
- 151 Dowdy SF: Overcoming cellular barriers for RNA therapeutics. *Nat Biotechnol* 35(3): 222-229, 2017. PMID: 28244992. DOI: 10.1038/nbt.3802
- 152 Bennett CF: Therapeutic antisense oligonucleotides are coming of age. *Annu Rev Med* 70: 307-321, 2019. PMID: 30691367. DOI: 10.1146/annurev-med-041217-010829
- 153 Saliminejad K, Khorram Khorshid HR, Soleymani Fard S and Ghaffari SH: An overview of microRNAs: Biology, functions, therapeutics, and analysis methods. *J Cell Physiol* 234(5): 5451-5465, 2019. PMID: 30471116. DOI: 10.1002/jcp.27486
- 154 Lee SWL, Paoletti C, Campisi M, Osaki T, Adriani G, Kamm RD, Mattu C and Chiono V: MicroRNA delivery through nanoparticles. *J Control Release* 313: 80-95, 2019. PMID: 31622695. DOI: 10.1016/j.jconrel.2019.10.007
- 155 Jones D: Setbacks shadow microRNA therapies in the clinic. *Nat Biotechnol* 36(10): 909-910, 2018. PMID: 30307922. DOI: 10.1038/nbt1018-909

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